(19) World Intellectual Property Organization

International Bureau



(43) International Publication Date 26 February 2004 (26.02.2004)

(10) International Publication Number WO 2004/016768 A2

(51) International Patent Classification7:

C12N

(US). SHRIVER, Mark, D. [US/US]; 720 Hemlock St.,

(21) International Application Number:

PCT/US2003/026229

(22) International Filing Date: 19 August 2003 (19.08.2003)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

60/404,357 60/467,613 19 August 2002 (19.08.2002) 2 May 2003 (02.05.2003) US

(63) Related by continuation (CON) or continuation-in-part (CIP) to earlier applications:

US Filed on

60/404,357 (CIP) 19 August 2002 (19.08.2002)

US

60/467,613 (CIP)

Filed on

2 May 2003 (02.05.2003)

- (71) Applicant (for all designated States except US): DNAPRINT GENOMICS, INC. [US/US]; 900 Cocoanut Avenue, Sarasota, FL 34236 (US).
- (72) Inventors; and
- (75) Inventors/Applicants (for US only): FRUDAKIS, Tony, N. [US/US]; 3707 Plumosa Terrace, Bradenton, FL 34210

Boalsburg, PA 16827 (US).

- (74) Agent: HAILE, Lisa, A.; Gray Cary Ware & Freidenrich LLP, 4365 Executive Drive, Suite 1100, San Diego, CA 92121-2133 (US).
- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

without international search report and to be republished upon receipt of that report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: COMPOSITIONS AND METHODS FOR INFERRING ANCESTRY

(57) Abstract: Ancestry informative marker (AIMs) polynucleotides that contain a single nucleotide polymorphism or that contain an insertion or deletion, are provided, as are methods of using panels of the AIMs to draw an inference as to a trait of an individual. The trait can be, for example, biogeographical ancestry, a pigmentation trait, responsiveness to a drug, or susceptibility to a disease. Also provided are methods of determining the proportional ancestry of an individual. Reagents and kits also are provided.

COMPOSITIONS AND METHODS FOR INFERRING ANCESTRY

BACKGROUND OF THE INVENTION

FIELD OF THE INVENTION

[0001] The invention relates generally to the identification of genetic markers predictive of an individual's biogeographical ancestry, and more specifically to combinations of single nucleotide polymorphisms useful as ancestry informative markers (AIMs), which allow an inference as to a trait of an individual, algorithms for identifying such AIMs, and methods of using such AIMs to infer a trait of an individual, including an individual's ancestry, responsiveness of an individual to a drug, and predisposition of an individual to a disease.

BACKGROUND INFORMATION

[0002] The majority (80-90%) of the genetic variation among human individuals is interindividual, and only a relatively small proportion (10-20%) is due to population differences (Nei, In Molecular Population Genetics (Columbia University Press, New York) 1987; Cavalli-Sforza et al., In The History and Geography of Human Genes (Princeton University Press, Princeton NJ) 1994; Deka et al., Electrophoresis 16:1659-1664, 1995; Rosenberg et al., Science 298:2381-2385, 2002; Akey et al., BioTechniques 30:348-367, 2001; Akey et al., Hum. Genet. 108:516-520, 2002). Most populations share alleles and those alleles that are most frequent in one population are also frequent in others. There are very few classical markers (e.g., blood group, serum protein, and immunological markers) or DNA genetic markers that are population-specific or have large frequency differentials among geographically and ethnically defined populations (Roychodhury and Nei, In Human Polymorphic Genes: World Distribution (Oxford University Press, New York) 1988; Dean et al., Amer. J. Hum. Genet. 55:788-808, 1994; Cavalli-Sforza et al., supra, 1994, Akey et al., supra, 2001, 2002). Despite this apparent lack of unique genetic markers, there are marked physical and physiological differences among human populations that presumably reflect genetic adaptation to unique ecological conditions, random genetic drift, and sex selection. In contemporary populations, these differences are evident in morphological differences between ethnic groups, as well as in differences in drug responsiveness and in susceptibility and resistance to disease.

[0003] On a basic level, human population structure can be represented in terms of BioGeographical Ancestry (BGA), which is the heritable component of "race" or heritage, and which is relevant on any scale of resolution. For example, on a crude level, BGA can be determined for 2 groups (e.g., European vs. others); or on a fine level, e.g., it can refer to "race" in terms of 4 groups such as IndoEuropeans, East Asians, sub-Saharan African and Native American; or on a finer level, e.g., it can refer to ethnicity within the European group (for example, Mediterranean or Scandinavian); or on a still finer level, e.g., it can even refer to groups of families within ethnic groups, such as groups of O'Reilly's descendent from a set of common ancestors within the Irish group. The measurement of BGA is relevant for most any type of genetics or epidemiological study design. For example, BGA is an important component in the variability of drug response (Burroughs et al., J. Natl. Med. Assoc. 94:1-26, 2002). The reason for this relationship is that genetic drift, geographical and/or reproductive isolation, and regional selective pressures have molded the allele frequencies of our ancestors for compatibility with alkaloids, tannins (self-defense chemicals), and other xenobiotics found in indigenous diets. Most drugs are derived from such chemicals and, therefore, it is no coincidence that the family of enzymes that allow humans to detoxify drugs are found at different frequencies in different populations. This scenario is not unique to drug responsiveness, and many other parts of the genome that are unrelated to drug responsiveness are subject to these same types of pressures.

[0004] Investigators generally have been concerned with identifying gene variants that cause a disease (the so called "phenotypically active" loci), rather than identifying gene variants that are simply correlated with disease. As such, whatever the trait being examined, and for most study designs involving unrelated individuals, it has been considered important to control for population structure so as to avoid identifying markers of structure that correlate with trait value in a given sample rather than those in linkage disequilibrium (LD) with phenotypically active loci (Risch et al., *Genome Biology* 3:1-12, 2001 Wang et al., *Amer. J. Hum. Genet.* 71:1227-1234, 2002; Burroughs et al., *supra*, 2002; Rao and Chakraborty, *Amer. J. Hum. Genet.* 26:444-453, 1974). There are two sources of population structure in a sample collection: 1) sampling effects, which can create structure even if sampling is performed from homogeneous populations, and 2) natural human demography.

3

The first source of population structure is a nuisance for genetics studies, and associations found from a study due to this type of structure are generally considered an artifact of the collection process rather than a reflection of human demography. Most geneticists generally consider the second kind of structure to be a nuisance as well. As such, associations identified as being due to population structure have been considered spurious findings or artifacts, and have generally been discarded; only findings due to true linkage or LD have been published, as such markers are considered linked to biologically relevant genes.

[0005] Much effort has been directed to quantifying both types of population structure (above) in groups of individuals. Such methods essentially measure the departure from expected levels of heterozygosity within a group of samples as an indication of structure (though none of these methods are capable of reading within-individual structure). Many common diseases exhibit locus and/or allelic heterogeneity as a function of BGA, and many authors have suggested that inappropriate attention to population structure during the study design step has produced at least some of the so-called "false positive" results implicated in the rash of irreproducible Common Disease/Common Variant results obtained to date (Terwilliger et al., Curr. Opin. Genet. Devel. 12:726-734, 2002). In order to control for the influence of population structure, several tests are appropriate (Cockerham, Evolution 23:72-83, 1969; Cockerham, Genetics 74:679-700, 1973; Wier and Cockerham, Evolution 38:1358-1370, 1984; Long, Genetics 112:629-647, 1986, Excoffier et al., Genetics 131:343-359, 1992). These methods can be grouped in two main categories - genomic control methods (Devlin and Roeder, Biometrics 55:997-1004, 1999), and structured association (SA) methods (Pritchard and Donelly, Theor. Popul. Biol. 60:227-237, 2001). Both methods require genotyping of a panel of unlinked markers to estimate and correct for the effect of genetic structure, but they are usually applied for sample collections. However, should a pool of samples fail such a test, it is usually not clear which samples should be eliminated to rectify the problem. An equally vexing problem with this method is that it is often applied for a study sample after the creation of expensive data, thus creating a circular logic problem in addition to an economic problem; these methods are usually employed to extract information on population structure using the characteristics of the data within which associations are sought.

4

[0006] In order to minimize the influence of population structure from the outset of an effort to identify phenotypically active loci, where structure or admixture is not to be used as a statistical fuel, it is generally desirable to qualify samples based on crude population stratifications such as BGA so that cases and controls can be matched and homogenized in composition. For example, it is not uncommon in the execution of case-control studies to ensure equal proportions or "racial homogeneity" within and between cases and controls. However, for most research purposes, the subjective methods used to measure population affiliation are unsatisfactory. As currently measured using biographical questionnaires, little knowledge of population structure other than the obvious is obtained, and only basic connections between population structure and drug response can be apparent and/or controlled. Consistency is a significant problem with the self-reporting of race on questionnaires, and one that the Food and Drug Administration is attempting to address during the clinical trial design process. However, using such subjective and imprecise methods of data collection, consistency can be a difficult end to achieve.

[0007] Rather than reformulating how questions are asked on questionnaires, consistency can be better addressed by replacing the subjective nature of the exercise with objective, reproducible scientific methods. Standardization and objectivity is of paramount importance for the collection of race data because its measure can be as subjective as that of any other human attribute. The self-reporting of race is not as trivial an exercise as the self-reporting of gender, and many people do not know their race or are of sufficient admixture that they have trouble classifying themselves into a single group. Such a scenario is particularly common in countries such as the United States, in which numerous cultures have been combined due to immigration. For example, a woman of mainly sub-Saharan African descent, raised in Puerto Rico, may describe herself as Hispanic. Though she socio-culturally identifies with Hispanics, however, her xenobiotic metabolism and drug target polymorphisms may more likely be associated with those shared among other sub-Saharans. By using nonanthropologic designations that describe the socio-cultural construct of society, current guidelines for considering information on race in the study design process can effect poor predictive power and false positive results. Where a person was raised and lives, and the cultural or sociological customs they observe, may have an impact on how that person responds to a

5

drug or proclivity to develop a disease. Thus, non-biological metrics are required, but the evidence suggests that BGA also has an impact and, therefore, needs to be measured in a scientifically accurate and reproducible manner.

[8000] Genetic markers present in a person's DNA provide the best opportunity to reliably determine the BGA an individual, and it has long been recognized that such a means is possible. For example, Reed (Science 244:575-576, 1973) and Neel (Mutat. Res. 26:319-328, 1974) referred to such markers as "private", and used them to estimate mutation rates. Reed (supra, 1973) used the term "ideal" (in reference to the utility of the markers in individual ancestry estimation) to describe hypothetical genetic marker loci at which different alleles are fixed in different populations. Chakraborty et al. (Ethnic. Dis. 1:245-256, 1991) referred to variants that are found in only one population as "unique alleles", and showed how allele frequencies could be inverted to provide a likelihood estimate of population, or BGA affiliation. The most useful "unique alleles" for the inference of BGA are those that also have large differences in allele frequency among populations (Reed, supra, 1973; Chakraborty et al., Genetics 130:231-243, 1992; Stephens et al., Amer. J. Hum. Genet. 55:809-824, 1994), and that have been referred to as "population-specific alleles" (PSAs, Shriver et al., Amer. J. Hum. Genet. 60:957-964, 1997; Parra et al., Amer. J. Hum. Genet. 63:1839-1851, 1998), but which are now referred to as "Ancestry Informative Markers" (AIMs; Shriver et al., Hum. Genet. 112:387-399, 2003, Frudakis et al., J. Forens. Sci. 48(4) 771-782, 2003).

[0009] Within the field of forensics, statistical methods that use simple tandem repeats (STRs) to infer the highest level of ancestry in a particular individual (majority BGA using proportional ancestry notation) can be fairly robust in terms of estimating majority BGA affiliation. Although STR tests can effectively resolve majority ancestral origin in most cases, an unacceptable number (5-10%) of classifications are ambiguous. Aside from sampling errors caused by rare alleles, and the fact that STRs were not selected from the genome for their ability to resolve population affiliation (i.e., STR allele frequency differentials are not necessarily nor optimally informative for this purpose), the major reason the high level of ambiguity likely is due to admixture, which is clearly a factor of the genetic variation for many human populations (Parra et al., supra, 1998, Cavalli-Sforza and Bodmer,

6

In *The genetics of human populations* (Dover Publications, NY; see pages 387-507) 1999; Rosenberg et al., *supra*, 2002). For a given study design, whether using self-reported information or DNA marker testing, and whether attempting to solve a pharmacogenomic or forensic problem, classifying a patient into a single group sacrifices the subtle, but not insignificant, information related to population structure and sub-structure; for example, there is no allowance to assign a person of 50% African and 50% European affiliation into a group. Unfortunately, markers and methods for allowing an accurate inference as to the BGA for more than just two groups at a time for an individual have not yet been described. Thus, a need exists for robust markers useful for inferring BGA, and for methods of identifying and using such markers. The present invention satisfies this need, and provides additional advantages.

SUMMARY OF THE INVENTION

[0010] The present invention provides methods and compositions for measuring, with a desired predetermined level of confidence, within individual population structure, which, as disclosed herein, allows inferences to be drawn, for example, as to ancestry, pigmentation traits, drug responsiveness, and disease susceptibility of the individual. By way of example, the present methods and compositions were used in a forensics capacity, wherein DNA samples obtained at the crime scenes of a serial murder/rapist in Louisiana were examined. Based on psychological profiling, police were of the belief that the serial killer was a Caucasian male, and had tested the DNA of over 1,000 Caucasian men without finding a match. The police then turned to the inventors, who, using the compositions and methods of the invention, determined that the individual committing the crimes was African American and, more specifically, had a proportional and confidence qualified ancestry of 85% sub-Saharan African and 15% Native American. Based on this result and additional results as disclosed herein, the police were further advised that the average African American is of 20% IndoEuropean ancestry, that greater levels of IndoEuropean ancestry correlate with lighter skin tone, and, therefore, that the person committing the crimes was likely an African American with average to darker than average skin tone. Within two months of refocusing their efforts based on this information, the police arrested an African American man of

7

average skin tone (for African Americans); DNA testing determined that he was the person whose DNA was found at the crime scenes.

[0011] Accordingly, the present invention relates to a method of inferring, with a predetermined level of confidence, a trait of an individual. Such a method can be performed, for example, by contacting a sample, which includes nucleic acid molecules of a test individual, with hybridizing oligonucleotides, wherein the hybridizing nucleotides can detect nucleotide occurrences of single nucleotide polymorphisms (SNPs) of a panel of at least about ten ancestry informative markers (AIMs) indicative of a population structure correlated with the trait, and wherein said contacting is performed under conditions suitable for detecting the nucleotide occurrences of the AIMs of the individual by the hybridizing oligonucleotides; and identifying, with a predetermined level of confidence, a population structure that correlates with the nucleotide occurrences of the AIMs in the individual, wherein the population structure correlates with a trait. As disclosed herein, a panel of at least about ten AIMs (e.g., 8, 9, 10, 11, 12, 13, 14, 15, 20, 25, 30, or more) is examined in practicing a method of the invention. Generally, the greater number of AIMs examined, the greater the confidence level of an inference made using the method.

loo12] A trait for which an inference is made according to a method of the invention can be any trait, including a trait for which an ethnic predisposition is known or suspected to occur and a trait for which it known that no ethnic predisposition occurs or for which it is not known or unclear as to whether there is an ethnic predisposition. In one embodiment, the trait is biogeographical ancestry (BGA). In one aspect, the panel of AIMs used to examine BGA includes AIMs as set forth in SEQ ID NOS:1 to 71. In another aspect, the panel includes AIMs as set forth in SEQ ID NOS:7, 21, 23, 27, 45, 54, 59, 63, and 72 to 152; in SEQ ID NOS:3, 8, 9, 11, 12, 33, 40, 59, 63, and 153 to 239; or in SEQ ID NOS:1, 8, 11, 21, 24, 40, 172, and 240 to 331, as well as a panel containing combinations of AIMs as set forth in SEQ ID NOS:1 to 331. As disclosed herein, AIMs useful in practicing a method of the invention can, but need not, be linked to a gene linked to the trait (i.e., a gene known to be involved in the trait phenotype) and generally are not in linkage disequilibrium with the gene (or locus). For example, an AIM useful for inferring drug responsiveness of an individual

according to a method of the invention need not be linked to a gene involved in responsiveness to the drug (e.g., a drug metabolism gene or a drug transport gene such as a cytochrome P450 gene or P-glycoprotein gene). Similarly, an AIM useful for inferring a pigmentation trait of an individual according to a method of the invention need not be linked to a gene involved in pigmentation (e.g., a tyrosinase gene or a melanocortin-1 receptor gene). Thus, in one aspect, at least one (e.g., 1, 2, 3, 4, or 5) AIM of a panel is not linked to a gene involved in the trait for which an inference is being made.

Where BGA is the trait for which an inference is being made, an individual being [0013]examined can have an ancestry that includes any one or a combination of ancestral groups, including, for example, a proportion of sub-Saharan African ancestry, Native American ancestry, IndoEuropean ancestry, East Asian ancestry, Middle Eastern ancestry, Pacific Islander ancestry, or a combination including one or more of these ancestries. As such, the proportional ancestry of an individual can comprise one ancestry (e.g., 100% IndoEuropean ancestry), or any proportion of two, three, four, or more ancestral groups. As such, a test individual (or individual of known proportional ancestry) can have, for example, a proportion of at least three ancestral groups, which can include proportions of sub-Saharan African ancestry and two other ancestries, or can include proportions of sub-Saharan African and IndoEuropean ancestral groups and a third ancestry; or Native American and IndoEuropean ancestral groups and a third ancestry; or East Asian and Native American ancestral groups and a third ancestry; or IndoEuropean and East Asian ancestral groups and a third ancestry; or can include proportions of Native American, East Asian, and IndoEuropean ancestral groups, or of sub-Saharan African, Native American, and IndoEuropean ancestral groups, and the like.

[0014] In another embodiment, a trait of a test individual for which an inference is being made is responsiveness of the individual to a drug, particularly a therapeutic drug. As such, a method of the invention provides a tool for realizing personalized medicine. A drug for which an inference can be made as to whether a test individual will be responsive, in either a positive or negative manner, can be, for example, a cancer chemotherapeutic agent such as paclitaxel, or a drug such as a statin, which can be useful for maintaining or lowering

cholesterol levels. In one aspect of this embodiment, AIMs of the panel of AIMs used to practice the method includes AIMs of genes other than genes known to be involved in melanin synthesis or metabolism.

[0015] In still another embodiment, a trait of a test individual for which an inference is being made is a susceptibility or predisposition of the individual to a disease. As disclosed herein, various traits are associated with population structure at a continental level, whereas other traits are associated with population structure at finer levels. As such, a method of the invention can provide an means for making an inference with respect to a trait such as disease susceptibility for diseases such as diabetes, hypertension, and cancers that are known to have an ethnic predisposition (i.e., known to occur with higher frequencies in individuals of certain ethnic/ancestral groups), as well as for disease such as such as alcoholism, or schizophrenia, Parkinson's disease, and other neurological disorders, which do not (or at least are not known to) have an ethnic predisposition.

[0016] In yet another embodiment, a trait of a test individual for which an inference is being made is a pigmentation trait. The pigmentation trait can be any such trait including, for example, eye color or shade, skin color, hair color, or a combination thereof. In one aspect of this embodiment, AIMs of the panel of AIMs used to practice the method includes AIMs of genes other than genes known to be involved in melanin synthesis or metabolism, or other aspects of pigmentation.

[0017] A method of inferring a trait of a test individual by determining a population structure that correlates with nucleotide occurrences of AIMs in the individual can further include identifying, with a predetermined level of confidence, a sub-population structure of the population structure, wherein the sub-population structure correlates with a trait. For example, a population structure of an individual can correlate to an intercontinental group with which, by inference, the individual shares ancestry, for example, IndoEuropean, and a sub-population structure can further correlate with an intracontinental group with which the individual shares IndoEuropean ancestry, for example, Mediterranean ethnicity.

10

[0018] The hybridizing oligonucleotides useful in the methods of the invention can be oligonucleotide probes or oligonucleotide primers. Oligonucleotide probes useful in the present methods can hybridize to a nucleotide sequence that includes the SNP position for an AIM, wherein the nucleotide at the position of the hybridizing oligonucleotide that corresponds to the position of the SNP for the AIM either matches or does not match the nucleotide occurrence at the SNP position. Additional oligonucleotide probes useful in the methods of the invention include oligonucleotide probes that hybridize to a polynucleotide sequence adjacent to and upstream and/or adjacent to and downstream of the SNP position, and that can, but need not, include a nucleotide corresponding to the nucleotide position of the SNP, and wherein such a corresponding nucleotide, when present in the probe, can, but need not match the nucleotide occurrence at the SNP.

[0019] Oligonucleotide primers useful in the methods of the invention include oligonucleotide primers useful for a primer extension reaction, as well as oligonucleotide primers that, in combination, allow for amplification of template polynucleotide comprising the AIM. Such amplification primer pairs generally include a forward primer and a reverse primer useful for amplification of a template polynucleotide comprising an AIM of interest. It will be recognized, however, that 2, 3, 4, or more different forward primers can be used with a common reverse primer for amplification of different template polynucleotides comprising the AIM (e.g., in a multiplex reaction) and a common gene sequence (e.g., AIMs of a family of related gene sequences) or for generating amplification products of different sizes from a single template. Similarly, one common forward primer can be used with one or a plurality of different reverse primers.

[0020] Accordingly, in one embodiment, a method of the invention is performed using oligonucleotide primers. In one aspect of this embodiment, the method includes contacting the sample with the oligonucleotide primers and with a polymerase, under condition suitable for generation of a primer extension product. In such a method, the nucleotide occurrence of a SNP can be determined by detecting the presence of the primer extension product, or by sequencing the primer extension product (or a product thereof) and identifying the nucleotide at the position corresponding to the position of the SNP. In another aspect of this

11

embodiment, the method includes contacting the sample with oligonucleotide primers that comprise amplification primer pairs and with a polymerase, under condition suitable for generation of an amplification product. In such a method, the nucleotide occurrence of a SNP can be determined by detecting the presence of the amplification product, or by sequencing the amplification product (or a product thereof) and identifying the nucleotide at the position corresponding to the position of the SNP.

[0021] The methods of the invention are particularly adaptable to being performed in a high throughput format, including in a multiplex format, thus allowing examination of a large number of AIMs and/or a large number of samples of test individuals, as well as controls, in parallel. As such, the methods can be performed using a format in which the samples being examined are arranged in an array, particularly an addressable array, e.g., on in wells in a tray or on a glass slide or silicon chip, and can be partly or fully automated using robotics. Where a multiplex platform is used, it will be recognized that the AIMs examined need not necessarily be those having the greatest delta values for the particular trait, but also can be selected to balance the delta value with the compatibility of primers in a multiplex set, for example, to select AIMs such that hybridizing oligonucleotides (e.g., amplification primer pairs) can be designed that can be used in a single reaction for examining a panel of AIMs but that do not substantially cross-hybridize with AIMs other than the target AIM for which the hybridizing oligonucleotides are designed.

[0022] The present invention also relates to a method of estimating, with a predetermined level of confidence, proportional ancestry of at least two ancestral groups of a test individual. Such a method can be performed, for example, by contacting a sample, which includes nucleic acid molecules of the test individual, with hybridizing oligonucleotides that can detect nucleotide occurrences of SNPs of a panel of at least about ten AIMs that are indicative of BGA for each ancestral group examined, wherein the contacting is under conditions suitable for detecting the nucleotide occurrences of the AIMs of the test individual by the hybridizing oligonucleotides; and identifying, with a predetermined level of confidence, a population structure that correlates with, or is most likely given, the nucleotide

occurrences of the AIMs of each of the ancestral groups examined, wherein the population structure is indicative of proportional ancestry.

[0023] The proportional ancestry estimated according to a method of the invention can be a proportion of any ancestral group, including, for example, a proportion of sub-Saharan African, Native American, IndoEuropean, East Asian, Middle Eastern, or Pacific Islander ancestral group, and generally is a combination of two or more of such ancestral groups. Thus, the proportional ancestry of a test individual can include proportions of sub-Saharan African and IndoEuropean ancestral groups (e.g., 80% sub-Saharan African and 20% IndoEuropean; or 60% sub-Saharan African, 20% IndoEuropean, and 20% of a third ancestral group); or can include proportions of Native American and IndoEuropean ancestral groups; East Asian and Native American ancestral groups; IndoEuropean and East Asian ancestral groups; and the like. Similarly, the proportional ancestry can include proportions of Native American, East Asian, and IndoEuropean ancestral groups; sub-Saharan African, Native American, and IndoEuropean ancestral groups; sub-Saharan African, Native American, and IndoEuropean ancestral groups; sub-Saharan African, Native American, and East Asian ancestral groups; and the like.

[0024] A panel of AIMs useful for estimating proportional ancestry of an individual can include AIMs as set forth in SEQ ID NOS:1 to 331, for example, AIMs as set forth in SEQ ID NOS:1 to 71, which can be useful for determining proportional ancestries including IndoEuropean, sub-Saharan African, East Asian, and Native American; or AIMs as set forth in SEQ ID NOS:7, 21, 23, 27, 45, 54, 59, 63, and 72 to 152, which can be useful for determining proportional ancestry of East Asians and sub-Saharan Africans; or in SEQ ID NOS:3, 8, 9, 11, 12, 33, 40, 59, 63, and 153 to 239, which can be useful for determining proportional ancestry of East Asians and IndoEuropeans; or in SEQ ID NOS:1, 8, 11, 21, 24, 40, 172, and 240 to 331, which can be useful for determining proportional ancestry of IndoEuropeans and sub-Saharan Africans.

[0025] In one embodiment, an estimate is made wherein the proportional ancestry includes proportions of three ancestral groups. In one aspect of this embodiment, identifying a population structure that correlates with, or is most likely given, the nucleotide occurrences of the AIMs of the test individual is practiced by performing a likelihood determination for

affiliation with each of a sub-Saharan African ancestral group, a Native American ancestral group, an IndoEuropean ancestral group, and an East Asian ancestral group; thereafter selecting three ancestral groups having a greatest likelihood value; determining a likelihood of all possible proportional affiliations among the three ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with the nucleotide occurrences of the AIMs of the test individual is identified; and identifying a single proportional combination of maximum likelihood.

[0026] In another aspect of this embodiment, identifying a population structure that correlates with, or is most likely given, the nucleotide occurrences of the AIMs is practiced by performing six two-way comparisons comprising likelihood determinations for affiliation between each group with each other group; thereafter selecting three ancestral groups having a greatest likelihood value; determining a likelihood of all possible proportional affiliations among the three ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with, or is most likely given, the nucleotide occurrences of the AIMs of the test individual is identified; and identifying a single proportional combination of maximum likelihood.

[0027] In still another aspect of the embodiment wherein an estimate is made wherein the proportional ancestry includes proportions of three ancestral groups, the method is practiced by performing three three-way comparisons among the groups; determining a likelihood of all possible proportional affiliations among the three ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with, or is most likely given, the nucleotide occurrences of the AIMs of the test individual is identified; and identifying a single proportional combination of maximum likelihood. In another aspect of this embodiment, the method can further include generating a graphical representation of the comparison of the three ancestral groups, wherein the graphical representation comprises a triangle with each ancestral group independently represented by a vertex of the triangle, and wherein the maximum likelihood value of proportional affiliation for an individual comprises a point within the triangle. If desired, the graphical

representation can further include a confidence contour that indicates a level of confidence associated with estimating the proportional ancestry.

In another embodiment, an estimate is made wherein the proportional ancestry [0028] includes proportions of four ancestral groups. In various aspects of this embodiment, identifying a population structure that correlates with, or is most likely given, the nucleotide occurrences of the AIMs of the test individual is practiced by performing six two-way comparisons, or by performing three three-way comparisons, or by performing one four-way comparison among the groups; determining a likelihood of all possible proportional affiliations among the four ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with, or is most likely given, the nucleotide occurrences of the AIMs of the test individual is identified; and identifying a single proportional combination of maximum likelihood. In one aspect of this embodiment, the method can further include generating a graphical representation of the comparison of the three ancestral groups, wherein the graphical representation comprises a pyramid with each ancestral group independently represented by a vertex of the pyramid, and wherein the maximum likelihood value of proportional affiliation for an individual comprises a point within the pyramid. If desired, the graphical representation can further include a confidence contour comprising a sphere around the point, wherein the sphere indicates a level of confidence associated with estimating the proportional ancestry.

[0029] The method of estimating, with a predetermined level of confidence, proportional ancestry of at least two ancestral groups of a test individual by identifying a population structure indicative of the proportional ancestry can further include identifying a sub-population structure indicative of ethnicity associated with one of the ancestral groups for which the test individual has a proportional ancestry. According to this method, a sub-population structure of the population structure that correlates with the nucleotide occurrences of the AIMs in the test individual is identified, wherein the sub-population structure correlates with ethnicity of the test individual. Such a method of identifying a sub-population structure can be performed, for example, by identifying those chromosomes of the test individual that contain the AIMs indicative of affiliation with a BioGeographical

ancestral group (where the individual is proportionally affiliated with more than one BioGeographical Ancestry group), contacting a sample including nucleic acid molecules of the test individual with second hybridizing oligonucleotides that can detect nucleotide occurrences of SNPs of a second panel of AIMs, wherein the AIMs of the second panel are informative for ethnicity within one of these groups and are present on the same chromosomes of the test individual that contain the AIMs indicative of the larger (intercontinental) ancestral group within which the ethnicity occurs; and identifying a sub-population structure that correlates with the nucleotide occurrences of the AIMs of the second panel, wherein the sub-population is indicative of ethnicity of the ancestral group of the test individual.

[0030] According to such a method, using hybridizing oligonucleotides specific for the first panel of AIMs (e.g., AIMs of the 71 exemplified AIMs; SEQ ID NOS:1 to 71), a test individual can be determined to be 60% IndoEuropean (IE) and 40% East Asian. In such a case, only a fraction of the total possible AIMs that can be indicative of the IE ancestral group will have been positive (if all were positive, the individual would have been 100% IE) and, therefore, only some of the individuals chromosomes or chromosomal regions will be of IndoEuropean origin. The chromosomes of the individual containing the positive AIMs for IE are then identified, and second hybridizing oligonucleotides specific for a second panel of AIMs are selected (e.g., from a group of 1000 or so AIMs that cover all 23 pairs of human chromosomes), wherein the AIMs of the second panel are limited to those that are highly variable in allele frequencies between IE ethnic groups and, therefore, indicative of IE ethnicity, and also are present on the chromosomes for which the first panel AIMs were IE positive. A sub-population structure that correlates with the nucleotide occurrences of the AIMs of the second panel is then identified, thus indicating an ethnicity with respect to the IE ancestral group of the test individual, for example, that the IE ancestral group derives from a Northern European, a Mediterranean, a Middle Eastern, or a South Asian Indian ethnicity. As such, the method provides a means to identify the ethnic origin of particular chromosomes (e.g., a Mediterranean origin of chromosomes previously determined to be of IndoEuropean origin) that contain AIMs that correlate with a population structure indicative of

IndoEuropean BioGeographical Ancestry, and further contain AIMs that correlate more specifically with a sub-population structure indicative of Mediterranean ethnicity.

[0031] In another embodiment, the method of estimating proportional ancestry of a test individual can include generating an ancestral map of the world, wherein locations of populations having a proportional ancestry corresponding to the proportional ancestry of the test individual are indicated on the ancestral map. As such, the method can supplement genealogical information. For example, the method can further include overlaying the ancestral map with a genealogical map, wherein the genealogical map indicates locations of populations having geopolitical relevance with respect to the test individual, and statistically combining the information of the ancestral map and genealogical map to obtain a most likely estimate of family history of the test individual.

[0032] Identifying a population structure that correlates with, or is most likely given, the nucleotide occurrences of the AIMs, according to a method of the invention, can be performed by comparing the nucleotide occurrences of the AIMs of the test individual with known proportional ancestries corresponding to nucleotide occurrences of AIMs indicative of BGA. The known proportional ancestries corresponding to nucleotide occurrences of AIMs indicative of BGA can be contained in a table or other list, and the nucleotide occurrences of the test individual can be compared to the table or list visually, or can be contained in a database, and the comparison can be made electronically, for example, using a computer. Further, each of the known proportional ancestries corresponding to nucleotide occurrences of AIMs indicative of BGA can be associated with a photograph of a person from whom the known proportional ancestry was determined, thus providing a means to further infer physical characteristics of a test individual. In one aspect, the photograph is a digital photograph, which comprises digital information that can be contained in a database that can further contain a plurality of such digital information of digital photographs, each of which is associated with a known proportional ancestry corresponding to nucleotide occurrences of AIMs indicative of BGA of the person in the photographs.

[0033] In another aspect, a method of the invention can further include identifying a photograph of a person having a proportional ancestry corresponding to the proportional

17

ancestry of the test individual. Such identifying can be done by manually looking through one or more files of photographs, wherein the photographs are organized, for example, according to the nucleotide occurrences of AIMs of the person in the photograph. Identifying the photograph also can be performed by scanning a database comprising a plurality of files, each file containing digital information corresponding to a digital photograph of a person having a known proportional ancestry, and identifying at least one photograph of a person having nucleotide occurrences of AIMs indicative of BGA that correspond to the nucleotide occurrences of AIMs indicative of BGA of the test individual.

[0034] Accordingly, the present invention also relates to an article of manufacture, which is at least one photograph of a person having a known proportional ancestry corresponding to a population structure comprising nucleotide occurrences of AIMs indicative of BGA, as well as to a plurality of such articles, each article of the plurality comprising one (or more) photograph(s) of a person having a known proportional ancestry corresponding to a population structure comprising nucleotide occurrences of AIMs indicative of BGA. The article can be contained in a file, or a plurality of the articles can be contained in a filed, for example, a file containing a plurality of photographs of different persons, wherein the some or all of the persons have the same or different known proportional ancestries that correspond to a population structure comprising nucleotide occurrences of AIMs indicative of BGA.

[0035] Accordingly, a plurality of such articles is provided, as is a plurality of files, each file of which can contain one or more articles, i.e., photographs, which can be of one or more persons having the same or different known proportional ancestries that correspond to a population structure comprising nucleotide occurrences of AIMs indicative of BGA. For example, different files of the plurality each can contain one (or more) photograph(s) of one person having a known proportional ancestry corresponding to a population structure comprising nucleotide occurrences of AIMs indicative of BGA. Different files of the plurality also can contain photographs of two or more different persons, each of whom has the same or substantially the same proportional ancestry corresponding to a population structure comprising nucleotide occurrences of AIMs indicative of BGA. As such, a plurality of files can contain files, each of which contains one or more photographs of one or more

persons, and when containing one or more photographs of two or more different persons, the different persons can have the same or different known proportional ancestries.

[0036] In one embodiment, the article of manufacture, i.e., the photograph of a person having a known proportional ancestry corresponding to a population structure comprising nucleotide occurrences of AIMs indicative of BGA, is a digital photograph, which comprises digital information. As such, the digital information of the digital photograph, or of a plurality of digital photograph articles of manufacture of the invention can be contained in a database. As such, the present invention further provides a plurality of the articles of manufactures, including at least two digital photographs each of which comprises digital information. In one aspect of this embodiment, the digital information for one or a plurality of the articles is contained in a database, which can be contained in any medium suitable for containing such a database, including, for example, computer hardware or software, a magnetic tape, or a computer disc such as floppy disc, CD, or DVD. As such, the database can be accessed through a computer, which can contain the database therein, can accept a medium containing the database, or can access the database through a wired or wireless network, e.g., an intranet or internet.

[0037] The present invention also relates a kit, which contains a plurality of hybridizing oligonucleotides, each hybridizing oligonucleotide including at least fifteen contiguous nucleotides of a polynucleotide as set forth in SEQ ID NOS:1 to 331, or a polynucleotide complementary thereto, and the plurality including at least five of such oligonucleotides, each based on different polynucleotides as set forth in SEQ ID NOS:1 to 331. In one embodiment, the hybridizing oligonucleotides that include at least fifteen contiguous nucleotides of at least five polynucleotides as set forth in SEQ ID NOS:1 to 71, or polynucleotides complementary to any of SEQ ID NOS:1 to 71.

[0038] The hybridizing polynucleotides of a kit of the invention can include probes, which are useful for detecting a particular AIM, including a particular nucleotide occurrence at the SNP position or DIP (deletion/insertion polymorphism) position of the AIM; can include primers, including primers useful for a primer extension reaction and primer pairs useful for a nucleic acid amplification reaction; or can include combinations of such probes and primers.

In one embodiment, a hybridizing oligonucleotide of the plurality includes a nucleotide corresponding to nucleotide position of the AIM (e.g., nucleotide 50 of any of SEQ ID NOS:1 to 34 and most others, nucleotide 56 of SEQ ID NO:35, nucleotide 44 of SEQ ID NO:50, or nucleotide 26 of SEQ ID NO:56), or to a nucleotide sequence complementary thereto, such a hybridizing oligonucleotide being useful as a probe to identify the presence or absence of a particular nucleotide occurrence at the SNP position of the AIM.

[0039] In another embodiment, the kit contains at least one pair of hybridizing oligonucleotides useful for detecting the nucleotide occurrence(s) at the SNP (or DIP) position of an AIM. In one aspect of this embodiment, a pair of hybridizing oligonucleotides includes one oligonucleotide that hybridizes upstream and adjacent to the SNP position of an AIM and a second oligonucleotide that hybridizes downstream of and adjacent to the SNP (or DIP) position of the AIM, wherein one or the other of the pair further contains a nucleotide complementary to a nucleotide occurrence suspected of being at the SNP (or DIP) position of the AIM (i.e., one of the polymorphic nucleotides), such a pair of hybridizing oligonucleotides being useful in an oligonucleotide ligation assay. In another aspect of this embodiment, a pair of hybridizing oligonucleotides includes an amplification primer pair, including a forward primer and a reverse primer, such a pair of hybridizing oligonucleotides being useful for amplifying a portion of polynucleotide that includes the SNP (or DIP) position of the AIM.

[0040] A kit of the invention can further contain additional reagents useful for practicing a method of the invention. As such, the kit can contain one or more polynucleotides comprising an AIM, including, for example, a polynucleotide containing an AIM for which a hybridizing oligonucleotide or pair of hybridizing oligonucleotides of the kit is designed to detect, such polynucleotide(s) being useful as controls. Further, hybridizing oligonucleotides of the kit can be detectably labeled, or the kit can contain reagents useful for detectably labeling one or more of the hybridizing oligonucleotides of the kit, including different detectable labels that can be used to differentially label the hybridizing oligonucleotides; such a kit can further include reagents for linking the label to hybridizing oligonucleotides, or for detecting the labeled oligonucleotide, or the like. A kit of the invention also can contain, for

example, a polymerase, particularly where hybridizing oligonucleotides of the kit include primers or amplification primer pairs; or a ligase, where the kit contains hybridizing oligonucleotides useful for an oligonucleotide ligation assay. In addition, the kit can contain appropriate buffers, deoxyribonucleotide triphosphates, etc., depending, for example, on the particular hybridizing oligonucleotides contained in the kit and the purpose for which the kit is being provided.

BRIEF DESCRIPTION OF THE DRAWINGS

segments are shuffled by recombination over time in an admixed population. Initially, the parental populations have chromosomal segments that are continuous with respect to AIMs along the segment. In the first filial (F1) generation all persons have one complete chromosomal segment from each parental population. In the F2 generation, many more combinations are possible. The relative likelihood of the non-recombinant vs. the recombinant genotypes shown in F2 is dependent on the size of the chromosomal segment. Segments of the order of the size of human chromosomes will average several recombination events in a single meiosis (one recombination is equally likely every 50 cM of genetic distance). F3 shows an example of a likely genotype for a person with two parents from the F2 generation. F(N) x F1 diagrams a genotype of a person with one F(N) parent and one F1 parent; and F(N) x F2 diagrams a genotype of a person with one F(N) parent and one F2 parent.

[0042] Figures 2A and 2B show triangle graphs generated using the algorithm described in Example 6 (see, also, Table 12). NAM, Native American; AFR, sub-Saharan African; EUR, IndoEuropean.

[0043] Figure 2A illustrates extension of a line from the NAM vertex to the opposite leg of the triangle, wherein the opposite leg represents 0% Native American ancestry. A circle is shown at the position of the estimated proportional ancestry (see Figure 2B), with the hatch mark on the line indicated the percent of Native American ancestry (approximately 15%).

[0044] Figure 2B shows additional lines drawn from the AFR and EUR vertices. The position on each line corresponding to the position of the circle represents the proportion of each respective ancestry; i.e., 15% Native American, 60% IndoEuropean, and 25% African.

[0045] Figure 3 shows a triangle plot depicting one approach to illustrate the value and precision of individual ancestry estimates. Typical distributions of three populations are shown (European Americans: filled squares; African-Americans: open triangles; and an African/Native American population: open circles). Also shown is a single individual with likelihood intervals represented as concentric rings surrounding the point estimate (filled circle). Like a topological map, each concentric ring represents a decrease in the likelihood by 1 log unit (10 times less likely). In this example, the individual has a likelihood interval space that is symmetrical and circular. Interval spaces will take many shapes depending on the admixture proportions of the subject in question and the allele frequencies of the markers that have been typed.

[0046] Figure 4 provides a triangular plot showing average admixture estimates for three African-American samples (filled circles: WASH-Washington DC, AFCAR-AfroCaribbeans, and BOG-Bogalusa), a European-American sample (open circle: SCO-State College), and a Spanish-American sample (open diamond: SLV-San Luis Valley CO). In parenthesis is shown the average African (AFR), IndoEuropean (EUR), and Native American (NAM) genetic contribution to each sample.

[0047] Figures 5A and 5B show the genetic structure in US resident populations.

[0048] Figure 5A shows the percentage of unlinked AIMs showing significant association. Expected values are based on a 5% significance level. Values for the Washington DC sample are based on 33 AIMs, for San Luis Valley CO on 19 AIMs, and for State College PA on 34 AIMs.

[0049] Figure 5B shows the correlation between individual ancestry estimates based on independent subsets of informative markers. Average correlation is based on 100 replicates.

The total number of markers is the same as for Figure 5A. The corresponding p values are indicated at the bottom of the graph.

[0050] Figures 6A and 6B show the triangle plots for a father (Figure 6A) and mother (Figure 6B).

[0051] Figures 7A to 7C show the triangle plots for each of three children of the father and mother represented in Figure 6.

[0052] Figure 8 shows the distribution of AIMs in the genome (chrom. number, chromosome number).

[0053] Figures 9A and 9B demonstrate the robustness of BGA admixture proportion analysis using AIMs (see Example 2). The confidence (contour lines) of the maximum likelihood estimate (MLE; point) is predictably affected by the elimination of AIMs informative for a particular pair-wise comparison. The first contour line extending from the MLE defines the triangle plot space within which the likelihood is 2 times lower than that of the MLE, and the second contour line defines the space in which the likelihood is 5 times lower than the MLE.

[0054] Figure 9A shows the MLE and confidence contours obtained using 71 AIMs; actual percentages are indicated.

[0055] Figure 9B shows the results obtained after eliminating those AIMs used to obtain the results shown in Figure 9A from the analysis that are informative for East Asian-Native American distinction. The MLE is relatively unaffected, and the confidence contours along the East Asian-Indo European (European) and Native American-European axes remain undistorted, but the confidence contours are distorted along the East Asian-Native American axis.

[0056] Figure 10 shows the BGA admixture proportions determined for each of eight individuals of a family pedigree. Circles represent females, squares males and the BGA affiliation for each individual is shown as a fraction where the numerator represents

IndoEuropean BGA and the denominator represents Native American BGA. None of the individuals harbored sub-Saharan African or East Asian BGA except as indicated by the asterisk (*), which indicates that the individual was determined to be of 4% East Asian BGA.

[0057] Figure 11 shows a family tree demonstrating how a Chinese great grandparent in an otherwise IndoEuropean family tree can produce a grandchild with IndoEuropean/East Asian ancestry. The individuals that are 100% East Asian (Chinese) are shown with shading; the admixture results for the male (square) at the bottom of the pedigree (short arrow) are of interest. The grandparent indicated by the long arrow is about a 50%/50% East Asian/IndoEuropean mix, and her daughter, the subject's mother, is expected to be a 25%/75% East Asian/IndoEuropean mix (see Example 3).

[0058] Figure 12 shows the distribution of all SNPs available for genotyping by chromosomal arm for a group of patients treated for elevated cholesterol levels.

[0059] Figure 13 shows the distribution of SNPs in Caucasian individuals taking LipitorTM (n = 180) for whom response was known in terms of cholesterol (lip TC), low density lipoprotein (lip LDL), liver transaminase AST-SGOT (lip SGOT) and ALT-GPT (lip GPT) measurements. SNPs with delta values of significance (>0.20) among the various trait classes were selected. For example, in about 70% of patients, LipitorTM causes a decrease in LDL. For any given SNP, the delta value (δ) is the difference in minor allele frequency among those individuals for whom LDL decreased by at least 20% versus those for whom LDL did not change.

[0060] Figure 14 shows a similar analysis as for Figure 13, except that response is measured following treatment with $Zocor^{TM}$ (n = 150), and only total cholesterol (zoc TC) and LDL (zoc LDL) were examined.

[0061] Figure 15 shows a distribution of SNPs ($\delta > 0.11$) among chromosome for 1,000 individuals of known eye color.

[0062] Figure 16 shows a distribution of SNPs ($\delta > 0.11$) among chromosome for 1,000 individuals of known hair eye color.

24

DETAILED DESCRIPTION OF THE INVENTION

[0063] The present invention is based on the identification of ancestry informative markers (AIMs) useful for inferring a level of population structure of an individual, which, in turn, allows an inference as to various traits of the individual. Further, the AIMs of the present invention are demonstrated to correlate with a trait, regardless of whether the marker is in linkage disequilibrium with a gene or locus known to be involved in the trait. As such, the AIMs of the present invention are distinguishable from previously described markers, which only were considered useful if they were linked with a trait, i.e., if the marker was physically close to a gene known to be involved in the trait as characterized, for example, in having a low cross-over percentage with respect to gene (or locus) known to be involved in (or associated with) the trait. In contrast, there is no requirement that the markers (AIMs) useful in the present methods be in linkage disequilibrium with a gene/trait and, in fact, AIMs that are disclosed herein as correlating with a trait can be located on different chromosomes from each other and from a gene/locus known to be associated with the trait.

[0064] AIMs are genetic loci that show alleles with high frequency differences between populations. AIMs are exemplified herein generally by single nucleotide polymorphisms (SNPs; see, e.g., SEQ ID NO:1), as well as by deletion/insertion polymorphisms (DIPs; see, e.g., SEQ ID NO:363). As disclosed herein, AIMs can be used to estimate BioGeographical Ancestry (BGA) of an individual or collection of individuals at the population level (in terms of races), at the sub-population level (in terms of ethnicities), and at the micro-group level (in terms of familial lines within ethnic groups), as well as at a practical, phenotypically qualified level (e.g., cases and controls). Such ancestry estimates at the subgroup and individual level can be directly instructive regarding the genetics of phenotypes that are different qualitatively or in frequency between populations, including, for example, the likelihood that an individual will respond to a particular medication or the propensity of an individual to develop a disease. Ancestry estimates also can provide a compelling foundation for the use of Admixture Mapping (AM) methods to identify the genes underlying these traits.

[0065] As exemplified herein, a panel of 71 AIMs (SEQ ID NOS:1 to 71) was identified from an examination of over 800 candidate AIMs (see, also, SEQ ID NOS:72 to 331), and

methods were developed to examine these AIMS as a means to obtain accurate estimates of proportional ancestry. The methods and markers of the invention have been validated in studies using skin pigmentation as a model phenotype (see, also, Intl. Publ. No. WO 02/097047 (PCT/US02/16789), which is incorporated herein by reference). Initial markers were genotyped in two population samples with primarily African ancestry, African Americans from Washington D.C. and an African Caribbean sample from England, and in a sample of European Americans from Pennsylvania (see Example 1). In the two African population samples, very strong correlations were observed between estimates of individual ancestry and skin pigmentation as measured by reflectometry ($R^2 = 0.21$, p < 0.0001 for the African-American sample and $R^2 = 0.16$, p < 0.0001 for the British African-Caribbean sample). These correlations confirmed the validity of the ancestry estimates and also indicated the high level of population structure related to admixture, which characterizes these populations and is detectable using other tests to identify genetic structure. These results demonstrate that an estimate of an individual's ancestry can be made based on a DNA analysis using a relatively small number of well defined genetic markers (AIMs).

[0066] The methods and genetic markers disclosed herein provide tools for several distinct purposes, including, for example, 1) for the estimation of ancestry proportions in individuals from their DNA; 2) for the estimation of genetic structure for the control of study designs commonly used for genetic research; 3) for the construction of physical profiles through the inference of characteristics related to ancestry, which may have implications in forensic investigations; 4) for the identification of disease predisposition, referred to as "Mapping by Ancestry Linkage Disequilibrium" (MALD); and 5) for predicting a significant portion of an individual patient's response to prescription and over-the-counter medications. As such, the present invention provides, for example, 1) statistical methods for the determination of ancestral proportions from genetic sequences within individuals and examples of use;
2) several hundred AIMs culled from the publicly available single nucleotide polymorphism (SNP) database and identified using statistical methods as useful for the determination of ancestral proportions within individuals or study groups; 3) several hundred AIMs that are demonstrated as useful for the determination of ancestral proportions within individuals or

study groups; and 4) software programs that can be used for the determination of ancestral proportions within individuals or study groups.

[0067] Previously, efforts have been made to control the two sources of population structure, including sampling effects and natural human demography, which were believed to confound efforts to identify markers of genes associated with particular traits. However, as disclosed herein, population structure is reflective of human demography, and markers that correlate with a trait value are useful as reporters of structure that correlate with trait value (rather than markers in LD with phenotypically active loci), and, therefore, provide a valuable tool that enables accurate classification in a cost-effective and practical manner. Alleles associated with a trait due to population structure are not linked to phenotypically active loci, but are merely correlated with trait value because they are enriched for in branches of the human family tree for which the trait value is more common. As disclosed herein, the distribution of trait values among the various branches of the human family tree are such that accurate classification can be obtained only through an appreciation of that structure, rather than a full understanding of the biological mechanism of the trait, and, as a result, markers that were considered false positives when considered with respect to their use for identifying phenotypically active loci, in fact, can enable accurate classification analysis; i.e., they are true positives provided the structure from which they were derived is reflective of human demography rather than sampling effects. The present methods are based on correlation between markers and BGA, where BGA is itself on some level of complexity correlated with a trait value, not linkage or linkage disequilibrium.

[0068] Accordingly, the present invention provides a method of inferring, with a predetermined level of confidence, a trait of an individual. In one embodiment, a method of the invention is performed by contacting a nucleic acid sample of a test individual with hybridizing oligonucleotides that can detect nucleotide occurrences of single nucleotide polymorphisms (SNPs) of a panel of at least about ten AIMs; and identifying, with a predetermined level of confidence, a population structure that correlates with, or is most likely given, the nucleotide occurrences of the AIMs in the individual, wherein the population structure correlates with a trait. The panel of AIMs are selected on their delta value (see

27

below) and, where relevant, based on the particular platform used to perform the method, and are indicative of a population structure correlated with the trait. AIMs are exemplified herein by the polynucleotides set forth as SEQ ID NOS:1 to 331, wherein the SNP position generally is at nucleotide position 50 (but see, e.g., SEQ ID NO:35, nucleotide 56; SEQ ID NO:51, position 48; SEQ ID NO:56, position 26).

[0069] A test individual for whom a trait is to be inferred can be any individual for whom it is desired to infer a trait, and generally is a human. However, the methods of the invention also can be used for inferring traits of other mammals, including, for example, domestic animals such as cats, dogs, or horses; farm animals such as cattle, sheep, pigs, or goats; or other animals. The trait to be examined can be any trait of interest, including, as exemplified herein, proportional ancestry (BGA); hair, skin or iris pigmentation; or drug responsiveness.

[0070] The methods of the invention are particularly useful because they allow for an inference to be made of a desired trait with a predetermined level of confidence. As used herein, reference to a "predetermined level of confidence" means that an inference or estimate of the invention is made using statistical methods that provide a confidence interval to be determined about a mean or a maximum likelihood value. In addition to determining the maximum likelihood value of within-individual or within-sample structure, other similarly likely values can also be determined and these can be combined to define the x-fold likelihood confidence intervals, where x is any number such as 2, 5 or 10. For example, all of the structure results corresponding to a likelihood value 10 times lower than the Maximum Likelihood Value can be plotted or listed to define the 10-fold likelihood confidence interval. As for any statistical test, an assay of the invention is designed such that performance of the test results in a value having a desired confidence level. As disclosed herein, a method of the invention can be performed such that the result has a predetermined level of confidence by varying the number of AIMs examined with respect to a trait. For example, use of a certain panel of ten AIMs will allow an inference to be made as to whether an individual has a particular trait, e.g., responsiveness to LipitorTM, with a certain level of confidence, whereas use of a panel of twenty AIMs, which can, but need not be partially overlapping with the panel of ten AIMs, will allow the same inference to be made, but with a higher level of confidence. Similarly, use of two panels of ten AIMs each can allow an inference to be made

that an individual has, for example, 80% IndoEuropean ancestry and 20% East Asian ancestry (with an error, e.g., of \pm 10%), whereas the use of two panels of twenty AIMs each can allow the same inference, but with an error, e.g., of \pm 5%.

[0071] A sample useful for practicing a method of the invention can be any biological sample of a test individual that contains nucleic acid molecules, including portions of the gene sequences containing AIMs that are to be examined or, wherein the polymorphism of an AIM results in an amino acid change in an encoded polypeptide, any biological sample that contains the encoded polypeptides. As such, the sample can be a cell, tissue or organ sample, or can be a sample of a biological fluid such as semen, saliva, blood, cerebrospinal fluid, and the like.

A nucleic acid sample useful for practicing a method of the invention will depend, [0072] in part, on whether the SNPs to be identified are in coding regions or in non-coding regions. Where one or more SNPs is present in a non-coding region of a gene, the nucleic acid sample generally is a deoxyribonucleic acid (DNA) sample, particularly genomic DNA or an amplification product thereof. However, where the AIM is contained within a transcribed sequence, e.g., rDNA, microsatellite DNA, or heteronuclear ribonucleic acid (RNA), which includes unspliced mRNA precursor RNA molecules including non-coding RNA sequence, an RNA sample can be used and examined directly, or a cDNA or amplification product thereof can be examined according to the present methods. Where one or more SNPs is present in a coding region of a gene, the nucleic acid sample can be DNA or RNA, or products derived therefrom, for example, amplification products. Furthermore, while the methods of the invention are exemplified with respect to a nucleic acid sample, it will be recognized that particular SNPs, when present in coding regions of a gene, can result in polypeptides containing different amino acids at the positions corresponding to the SNPs due to non-degenerate codon changes. As such, in one aspect, the methods of the invention are practiced using a sample containing polypeptides of the subject.

[0073] A method of the invention is performed by contacting the sample and hybridizing oligonucleotides under conditions suitable for detecting the nucleotide occurrences of the AIMs of the individual by the hybridizing oligonucleotides. Further, in aspects of the

methods of the invention, the sample can be contacted with second hybridizing oligonucleotides, for example, to determine a sub-population structure. It should be recognized that the term "second", when used in reference to hybridizing oligonucleotides (or to a panel of AIMs), is used for convenience of discussion so as to allow a clear distinction, e.g., of steps for performing a method. In this respect, it should be further recognized that one or more hybridizing oligonucleotides used, e.g., to determine a population structure, also can be included among the second hybridizing oligonucleotides.

[0074] Conditions suitable for detecting the nucleotide occurrences of AIMs will vary depending on the sequences of the hybridizing oligonucleotides, including their length and complementarity, as well as on the particular assay being used and, for example, whether the assay is being performed as a multiplex assay. The hybridizing oligonucleotides, which are at least 15 nucleotides in length, can contain deoxyribonucleotides or ribonucleotides, which are linked together by a phosphodiester bond, and can be single stranded or double stranded, though they generally are used in a single stranded form. Such hybridizing oligonucleotides can be prepared using methods of chemical synthesis or by enzymatic methods such as by the polymerase chain reaction (PCR).

[0075] The hybridizing oligonucleotides, or other polynucleotides useful in a methods or contained in a kit of the invention also can contain nucleoside or nucleotide analogs, and can have a backbone bond other than a phosphodiester bond, such oligonucleotides providing certain advantages such as having increased stability or more desirable hybridization properties. Nucleotide analogs are well known in the art and commercially available, as are polynucleotides containing such nucleotide analogs (Lin et al., Nucl. Acids Res. 22:5220-5234, 1994; Jellinek et al., Biochemistry 34:11363-11372, 1995; Pagratis et al., Nature Biotechnol. 15:68-73, 1997, each of which is incorporated herein by reference). The covalent bond also can be any of numerous other bonds, including a thiodiester bond, a phosphorothioate bond, a peptide-like bond or any other bond known to those in the art as useful for linking nucleotides to produce synthetic oligonucleotides (see, for example, Tam et al., Nucl. Acids Res. 22:977-986, 1994; Ecker and Crooke, BioTechnology 13:351360, 1995, each of which is incorporated herein by reference). The incorporation of non-naturally occurring nucleotide analogs or bonds linking the nucleotides or analogs can be particularly

useful where the oligonucleotide is to be exposed to an environment that can contain a nucleolytic activity, including, for example, a tissue culture medium or sample comprising a cell extract because the modified oligonucleotides can be less susceptible to degradation.

[0076] Generally, the hybridizing oligonucleotides useful for purposes of the present invention are at least about 15 bases in length, which is sufficient to permit the oligonucleotide to selectively hybridize to a target polynucleotide comprising the AIM, and can be at least about 18 nucleotides or 21 nucleotides or 25 nucleotides or more in length. The term "selective hybridization" or "selectively hybridize" refers to hybridization under moderately stringent or highly stringent physiological conditions, which can distinguish related nucleotide sequences from unrelated nucleotide sequences. In nucleic acid hybridization reactions, the conditions used to achieve a particular level of stringency are known to vary, depending on the nature of the nucleic acids being hybridized, including, for example, the length, degree of complementarity, nucleotide sequence composition (e.g., relative GC:AT content), and nucleic acid type, i.e., whether the oligonucleotide or the target nucleic acid sequence is DNA or RNA. An additional consideration is whether one of the nucleic acids is immobilized, for example, on a filter, bead, chip, or other solid matrix.

[0077] Methods for selecting appropriate stringency conditions can be determined empirically or estimated using various formulas, and are well known in the art (see, for example, Sambrook et al., supra, 1989). An example of progressively higher stringency conditions is as follows: 2X SSC/0.1% SDS at about room temperature (hybridization conditions); 0.2X SSC/0.1% SDS at about room temperature (low stringency conditions); 0.2X SSC/0.1% SDS at about 42°C (moderate stringency conditions); and 0.1X SSC at about 68°C (high stringency conditions). Washing can be carried out using only one of these conditions, for example, high stringency conditions, or each of the conditions can be used, for example, for 10 to 15 minutes each, in the order listed above, repeating any or all of the steps listed. As such, final conditions will vary, depending on the particular hybridization reaction involved, and can be determined empirically. It should be recognized that a variety of conditions can be utilized to provide selective hybridization conditions. For example, when a multiplex assay is to be performed using a plurality of different hybridizing oligonucleotides specific for different AIMs of a panel, the conditions (as well as the AIMs/hybridizing

oligonucleotides) can be selected such that selective hybridization occurs for all of the hybridizing oligonucleotides in the reaction.

In various embodiments, it can be useful to detectably label a polynucleotide or [0078]hybridizing oligonucleotide. Detectable labeling of a polynucleotide is well known in the art and includes, for example, the use of detectable labels such as chemiluminescent labels, radionuclides, enzymes, haptens such as digoxygenin and biotin, fluorophores, and unique oligonucleotide sequences. For example, PCR products can be performed, wherein one primer is biotinylated and the other primer contains digoxygenin. The amplification products can then be bound to a streptavidin plate, washed, reacted with an enzyme-conjugated antibody to digoxygenin, and developed with a chromogenic, fluorogenic, or chemiluminescent substrate for the enzyme. Alternatively, a radioactive method can be used to detect generated amplification products, for example, by including a radiolabeled deoxynucleoside triphosphate into the amplification reaction, then blotting the amplification products onto DEAE paper for detection. In addition, if one primer is biotinylated, then streptavidin-coated scintillation proximity assay plates can be used to measure the PCR products. Additional methods of detection can use a chemiluminescent label, for example, a lanthanide chelate such as used in the DELFIA® assay (Pall Corp.), a fluorescent label, or an electrochemiluminescent label such as ruthenium tris-bipyridyl (ORI-GEN).

[0079] Methods for detecting a nucleotide occurrence at a SNP or DIP position of an AIM can utilize one or more oligonucleotide probes or primers, including, for example, an amplification primer pair, that selectively hybridize to a target polynucleotide spanning the AIM. Oligonucleotide probes useful in practicing a method of the invention can include, for example, an oligonucleotide that is complementary to and spans a portion of the target polynucleotide, including the position of the SNP (or DIP), wherein the presence of a specific nucleotide at the position of the SNP is detected by the presence or absence of selective hybridization of the probe. Such a method can further include contacting the target polynucleotide and hybridized oligonucleotide with an endonuclease, and detecting the presence or absence of a cleavage product of the probe, depending on whether the nucleotide occurrence at the SNP site is complementary to the corresponding nucleotide of the probe. A

32

pair of probes that specifically hybridize upstream and adjacent and downstream and adjacent to the site of the SNP, wherein one of the probes includes a nucleotide complementary to a nucleotide occurrence of the SNP, also can be used in an oligonucleotide ligation assay, wherein the presence or absence of a ligation product is indicative of the nucleotide occurrence at the SNP site. An oligonucleotide also can be useful as a primer, for example, for a primer extension reaction, wherein the product (or absence of a product) of the extension reaction is indicative of the nucleotide occurrence. In addition, a primer pair useful for amplifying a portion of the target polynucleotide including the SNP or DIP site can be useful, wherein the amplification product is examined to determine the nucleotide occurrence at the SNP site or to determine whether there is an insertion or a deletion at the DIP site.

[0080] Numerous methods are known for determining a nucleotide occurrence at a particular position in a polynucleotide (i.e., of a SNP or DIP). Such methods can utilize one or more oligonucleotide probes or primers, including, for example, an amplification primer pair, that selectively hybridize to a target polynucleotide, which contains one or more SNP positions. Hybridizing oligonucleotide useful in practicing a method of the invention can include, for example, an oligonucleotide that is complementary to and spans a portion of the target polynucleotide, including the position of the SNP or DIP (including whether the DIP has a deletion or insertion), wherein the presence of a specific nucleotide at the SNP site or the presence of a deletion or insertion at the DIP site is detected by the presence or absence of selective hybridization of the oligonucleotide probe. Such a method can further include contacting the target polynucleotide and hybridized oligonucleotide with an endonuclease, and detecting the presence or absence of a cleavage product of the probe, depending on whether the nucleotide occurrence at the SNP site is complementary to the corresponding nucleotide of the probe.

[0081] An oligonucleotide ligation assay also can be used to identify a nucleotide occurrence at a SNP site, wherein a pair of probes that selectively hybridize upstream and adjacent to and downstream and adjacent to the site of the SNP, and wherein one of the probes includes a terminal nucleotide complementary to a nucleotide occurrence of the SNP. Where the terminal nucleotide of the probe is complementary to the nucleotide occurrence, selective hybridization includes the terminal nucleotide such that, in the presence of a ligase,

the upstream and downstream oligonucleotides are ligated. As such, the presence or absence of a ligation product is indicative of the nucleotide occurrence at the SNP site.

[0082] A hybridizing oligonucleotide also can be useful as a primer, for example, for a primer extension reaction, wherein the product (or absence of a product) of the extension reaction is indicative of the nucleotide occurrence at a SNP site or an insertion or deletion at a DIP site. In addition, a primer pair useful for amplifying a portion of the target polynucleotide including the SNP or DIP site can be useful, wherein the amplification product is examined to determine the nucleotide occurrence at the SNP site or the presence of a deletion or an insertion at the DIP site. Particularly useful methods include those that are readily adaptable to a high throughput format, to a multiplex format, or to both.

[0083] Conditions that allow generation of an amplification product in a sample in which an amplification reaction is being performed are such that the reaction contains the necessary components for the amplification reaction to occur. Such conditions include, for example, appropriate buffer capacity and pH, salt concentration, metal ion concentration if necessary for the particular polymerase, appropriate temperatures that allow for selective hybridization of the primer or primer pair to the template target polynucleotide, as well as appropriate cycling of temperatures that permit polymerase activity and melting of a primer or primer extension or amplification product from the template or, where relevant, from forming a secondary structure such as a stem-loop structure. Such conditions and methods for selecting such conditions are routine and well known in the art (see, for example, Innis et al., "PCR Strategies" (Academic Press 1995); Ausubel et al., "Short Protocols in Molecular Biology" 4th Edition (John Wiley and Sons, 1999), each of which is incorporated herein by reference).

[0084] A primer extension or amplification product can be detected directly or indirectly and/or can be sequenced using various methods known in the art. Amplification products that span a SNP site can be sequenced using traditional sequence methodologies, including, for example, the dideoxy-mediated chain termination method (Sanger et al., *J. Molec. Biol.* 94:441, 1975; Prober et al. *Science* 238:336-340, 1987) or the chemical degradation method (Maxam et al., *Proc. Natl. Acad. Sci. USA* 74:560, 1977) to determine the nucleotide occurrence at the SNP loci.

34

[0085] The nucleotide occurrence at a SNP site also can be determined using a microsequencing method, wherein the identity of only a single nucleotide is determined at a predetermined site (U.S. Pat. No. 6,294,336). Microsequencing methods include the Genetic Bit Analysis method (WO 92/15712). Additional, primer-guided, nucleotide incorporation procedures for assaying polymorphic sites in DNA have also been described (Komher et al., Nucl. Acids. Res. 17:7779-7784, 1989; Sokolov, Nucl. Acids Res. 18:3671, 1990; Syvanen et al., Genomics 8:684-692, 1990; Prezan et al, Hum. Mutat. 1:159-164, 1992; Nyren et al., Anal. Biochem. 208:171-175, 1993). These methods differ from Genetic Bit™. Analysis in that they all rely on the incorporation of labeled deoxyribonucleotides to discriminate between bases at a polymorphic site. In such a format, the signal is proportional to the number of deoxyribonucleotides incorporated, and polymorphisms that occur in runs of the same nucleotide generate signals that are proportional to the length of the run (Syvanen et al. Amer. J. Hum. Genet. 52:46-59, 1993).

[0086] Another method for determining the nucleotide occurrence at a SNP position is described by Macevicz (U.S. Pat. No. 5,002,867), wherein a nucleic acid sequence is determined via hybridization with multiple mixtures of oligonucleotide probes. In accordance with such a method, the sequence of a target polynucleotide is determined by permitting the target to sequentially hybridize with sets of probes having an invariant nucleotide at one position, and a variant nucleotides at other positions. The nucleotide sequence is determined by hybridizing the target with a set of probes, then determining the number of sites that at least one member of the set is capable of hybridizing to the target (i.e., the number of matches). This procedure is repeated until each member of a sets of probes has been tested. U.S. Pat. No. 6,294,336 provides a solid phase sequencing method for determining the sequence of nucleic acid molecules (either DNA or RNA) by utilizing a primer that selectively binds a polynucleotide target at a site wherein the SNP is the most 3' nucleotide selectively bound to the target.

[0087] The nucleotide occurrence of a SNP in a sample also can be determined using the SNP-ITTM method (Orchid BioSciences, Inc., Princeton, NJ). In general, SNP-ITTM is a 3-step primer extension reaction. In the first step a target polynucleotide is isolated from a

sample by hybridization to a capture primer, which provides a first level of specificity. In a second step the capture primer is extended from a terminating nucleotide trisphosphate at the target SNP site, which provides a second level of specificity. In a third step, the extended nucleotide trisphosphate can be detected using a variety of known formats, including: direct fluorescence, indirect fluorescence, an indirect colorimetric assay, mass spectrometry, fluorescence polarization, etc. Reactions can be processed in 384 well format in an automated format using a SNPstreamTM instrument (Orchid BioSciences, Inc., Princeton, NJ). Phase known data can be generated by inputting phase unknown raw data from the SNPstreamTM instrument into the Stephens and Donnelly's PHASE program.

[0088] Melting curve analysis of SNPs (McSNP® analysis) provides another method for detecting a nucleotide occurrence in an AIM (Akey et al., *supra*, 2001). McSNP® analysis provides the additional advantages that it does not require a step of gel electrophoresis, thus minimizing the time and cost for detecting a SNP, and that it is readily adaptable to high throughput formats, thus allowing examination of one or more panels of AIMs and/or samples in parallel.

[0089] Where the particular nucleotide occurrence of a SNP is such that the nucleotide occurrence results in an amino acid change in an encoded polypeptide, the nucleotide occurrence can be identified indirectly by detecting the particular amino acid in the polypeptide. The method for determining the amino acid will depend, for example, on the structure of the polypeptide or on the position of the amino acid in the polypeptide. Where the polypeptide contains only a single occurrence of an amino acid encoded by the particular SNP, the polypeptide can be examined for the presence or absence of the amino acid. For example, where the amino acid is at or near the amino terminus or the carboxy terminus of the polypeptide, simple sequencing of the terminal amino acids can be performed.

Alternatively, the polypeptide can be treated with one or more enzymes and a peptide fragment containing the amino acid position of interest can be examined, for example, by sequencing the peptide, or by detecting a particular migration of the peptide following electrophoresis. Where the particular amino acid comprises an epitope of the polypeptide, the specific binding, or absence thereof, of an antibody specific for the epitope can be

36

detected. Other methods for detecting a particular amino acid in a polypeptide or peptide fragment thereof are well known and can be selected based, for example, on convenience or availability of equipment such as a mass spectrometer, capillary electrophoresis system, magnetic resonance imaging equipment, and the like.

[0090] In another embodiment, a method of the invention utilizes an antibody, or antigen binding fragment thereof, that specifically binds, for example, to a polypeptide comprising an amino acid encoded by a nucleotide sequence comprising one nucleotide occurrence of a SNP, but not substantially to a polypeptide comprising an different amino acid encoded by the codon comprising the SNP; or that specifically binds, for example, to a polypeptide comprising an amino acid sequence encoded by one form a DIP (e.g., that having the insertion), but not substantially to that encoded by the alternative form (e.g., that having the deletion). As used herein, the term "specific interaction," or "specifically binds" means that two molecules form a complex that is relatively stable under physiologic conditions. The term is used herein to refer to various interactions, including, for example, the interaction of an antibody that binds a target polynucleotide including the SNP site only if the SNP has a specified, but not an alternative, nucleotide occurrence (e.g, an A, but not a T); or the interaction of an antibody that binds a polypeptide that includes one amino acid that is encoded by a codon that includes a SNP site, but not a polypeptide having an alternative amino acid encoded by the codon comprising the SNP.

[0091] A specific interaction can be characterized by a dissociation constant of at least about 1×10^{-6} M, generally at least about 1×10^{-7} M, usually at least about 1×10^{-8} M, and particularly at least about 1×10^{-9} M or 1×10^{-10} M or greater. A specific interaction generally is stable under physiological conditions, including, for example, conditions that occur in a living individual such as a human or other vertebrate or invertebrate, as well as conditions that occur in a cell culture such as used for maintaining mammalian cells or cells from another vertebrate organism or an invertebrate organism. Methods for determining whether two molecules interact specifically are well known and include, for example, equilibrium dialysis, surface plasmon resonance, and the like.

37

[0092] Antibodies useful in a method of the invention include antibodies that specifically bind polynucleotides that encompass an AIM, or that bind polypeptides that include an amino acid encoded by a codon that includes a SNP or that include amino acids due to an insertion at a DIP site. Such antibodies are selected such that they specifically bind a polypeptide that includes a first amino acid encoded by a codon that includes the SNP loci, but do not bind, or bind measurably more weakly to a polypeptide that includes a second amino acid encoded by a codon that includes a different nucleotide occurrence at the SNP.

[0093] The term "antibody" is used broadly herein to refer to immunoglobulin molecules and antigen binding portions of immunoglobulin molecules that specifically bind an antigen. As such, antibodies useful in a method of the invention can be polyclonal, monoclonal, multispecific, human, humanized or chimeric antibodies, single chain antibodies, Fab fragments, F(ab') fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies, and the like, as well as antigen/epitope binding fragments of such antibodies. Antigen binding fragments of antibodies include, but are not limited to, Fab, Fab' and F(ab')2, Fd, single-chain Fv's (scFv), single-chain antibodies, disulfide-linked Fv fragments (sdFv) and fragments comprising either a VL or VH domain. Thus, antigen-binding antibody fragments, including single-chain antibodies, can comprise the variable region(s) alone or in combination with the entirety or a portion of the hinge region, CH1, CH2, and/or CH3 domains. The antibodies can be from any animal origin including birds and mammals, or can be expressed recombinantly, for example, in insect or mammalian host cells or in plants.

[0094] There is much that can be learned today through the use of genetic markers in numerous scientific fields. The use of genetic sequences has become routine for forensics and disease research, but the majority of the benefits from the recently completed human genome project still await discovery. Within the genome exist sequences and patterns of sequences that will prove useful for a variety of purposes including increasing crop yields, extending human life spans, minimizing the suffering caused by drugs and enhancing the quality of our lives through better, more effective and specific treatments. Until now, biomedical research has been conducted on relatively simple terms. Nevertheless, more one

thousand simple Mendelian traits have been mapped by following the transmission of genetic markers in families.

[0095] Many statistical methods are available for studying genetic traits, including traditional family-based linkage analysis, variance component methods, sib-pair linkage, measured genotype, transmission disequilibrium, genomic control, and structure analysis. Some of the genes underlying variation in susceptibility to common diseases (e.g., heart disease, obesity, type 2 diabetes, hypertension, and cancer) eventually will be identified using genetic approaches. However, there are a number of complexities in genetic research on common diseases because many of these conditions are multifactorial (i.e., have several sources of variability in risk) and polygenic (i.e., result due to the actions and interactions among several genes). Additional difficulties in the study of common diseases can derive from the late onset of symptoms and heterogeneity in etiology. Thus, identifying the genes involved in complex diseases remains one of the greatest challenges in the field of human genetics.

[0096] There has been increased interest in association studies as a useful approach to map common disease and drug response genes (Risch and Merikangas, Science 273:1516-1517, 1996; Jorde, Genome Res. 10:1435-1444, 2000; Nordborg and Tavaré, Trends Genet. 18:83-90, 2002). Until the present disclosure, however, the implication of ancestry for identifying these genes has not been fully appreciated. As such, the methods of the invention provide a previously undescribed platform for the identification of genes associated with disease susceptibility and drug responsiveness, as well as for the development of advanced forensic methods. As such, compositions and methods are provided for inferring an individual's response to commonly used medications, which, remarkably, is a function of individual ancestry; the disclosed markers and methods are, to a differing extent for each drug, useful for the inference of such response. In addition, compositions and methods are provided for inferring individual and/or group ancestral proportions from knowledge of the individual's or group's DNA sequences. Further, compositions and methods are provided for using knowledge of ancestry relevant DNA sequences to identify disease susceptibility and drug response genes through the MALD process. Also, compositions and methods are provided for qualifying and normalizing study groups for more traditional methods of

mapping disease genes. Each of these processes requires an accurate knowledge of ancestry, which can be determined using the methods and compositions disclosed herein.

[0097] The populations that will be best suited for linkage disequilibrium (LD) mapping has prompted much discussion and debate (see Wright et al., Nat. Genet. 23:397-404, 1999; Eaves et al., Nat. Genet 25:320-323, 2000; Nordborg and Tavaré, supra, 2002; Kaessmann et al., Amer. J. Hum. Genet. 70:673-685, 2002). The extent of LD is a complex function of a number of genetic and evolutionary factors such as mutation, recombination and gene conversion rates, demographic and selective events, and the age of the mutation itself. Some of these factors affect the whole genome, while others only affect particular genome regions. Additionally, variation in mutation, recombination, and gene conversion rates throughout the genome are expected to create LD differences between genomic regions (see, for example, Taillon-Miller et al., Nat. Genet. 25:324-328, 2000).

It has been proposed that small, isolated and inbred populations will have advantages over other populations, due to the lower heterogeneity and the larger extent of linkage disequilibrium (Wright et al., supra, 1999; Nordborg and Tavaré, supra, 2002; Kaessmann et al., supra, 2002). Other populations well suited for mapping are recently admixed populations (e.g., Hispanics and African Americans), which offer the advantage that LD has been created recently due to the admixture process. Because this LD is recent, it can extend over large chromosomal regions. However, it is also extremely important to control for the genetic structure (inter-individual variation in admixture proportions) present in these populations in order to avoid false positives (Parra et al., supra, 1998; Lautenberger et al., Amer. J. Hum. Genet. 66:969-978, 2000; Pfaff et al., Amer. J. Hum. Genet. 68:198-207, 2001; Nordborg and Tavaré, supra, 2002, each of which is incorporated herein by reference). Interest in admixture mapping has increased in recent years (McKeigue et al., Ann. Hum. Genet. 64:171-186, 2000; Smith et al., J. Invest. Dermatol. 111:119-122, 2001; Collins-Schramm et al., Amer. J. Hum. Genet. 70:737-750, 2002, each of which is incorporated herein by reference). A general description of admixture mapping is provided below, as are some details about a statistical approach developed for admixture mapping and its application to skin pigmentation as a model phenotype.

[0099] Admixture generates allelic associations between all marker loci where allele frequencies are different between the parental populations (Chakraborty and Weiss, Proc. Natl. Acad. Sci., USA 85:9119-9123, 1988). These associations decay with time in a way that is dependent on the genetic distance between them. Thus, disease (or trait) risk alleles that are different between the parental populations can be mapped in admixed populations using special panels of genetic markers showing high frequency differences between the parental populations. These markers, termed AIMs, are characterized by having particular alleles that are more common in one group of populations than in other populations. One measure of the informativeness of such markers is the allele frequency differential, delta (δ) , which is simply the absolute value of the difference of a particular allele between populations (Chakraborty and Weiss, supra, 1988; Dean et al., supra, 1994).

In admixed populations, allelic associations were generated recently and, therefore, [0100]are more easily detected for a given sample size because they extend over longer distances than in non-admixed populations (up to 10-20 centiMorgans (cM) or more). The statistical basis of this approach was first explored by Chakraborty and Weiss (supra, 1988) and subsequently by Stephens, Briscoe and O'Brien, who named the method "mapping by admixture linkage disequilibrium" (MALD; Stephens et al., Amer. J. Hum. Genet. 55:809-824, 1994; Briscoe et al., J. Hered. 85:59-63, 1994). Further, whether one is using a MALD approach or a more traditional LD approach for genetic research, to eliminate associations of the trait with alleles at unlinked loci, it is necessary to control in the analysis for individual ancestry estimated from the marker data. The SNP sequences (markers; AIMs) and methods disclosed herein (BGA test) are a particularly efficient means by which to accomplish this task. An Analysis of Covariance (ANCOVA) test has been employed using the estimate of individual admixture as a conditioning variable to control for the effect of individual ancestry in two ways: 1) leaving out the locus under consideration (ANCOVA/IAE minus marker); and 2) using the complete individual ancestry estimate for the conditioning (ANCOVA/IAE). This method is described in detail herein.

[0101] An alternative approach to exploiting admixture has been developed that, while based on earlier work, has little in common with classical LD mapping, and is more

41

analogous to linkage analysis of an experimental cross (McKeigue, Amer. J. Hum. Genet. 63:241-251, 1998, which is incorporated herein by reference; McKeigue et al., supra, 2000). For this reason, the term "admixture mapping" has been proposed as more appropriate than "mapping by admixture linkage disequilibrium". Instead of testing for allelic associations, according to the present methods, the underlying variation in ancestry is modeled on chromosomes of mixed descent to extract all the information about linkage that is generated by admixture. The methods and markers disclosed are necessary and sufficient to accomplish this process. Advanced statistical methods are utilized to apply this approach in practice, though the underlying principle on which it relies to detect linkage is straightforward. Suppose, for instance, that a locus accounts for some of the variation in pigmentation between West Africans and Europeans. If individuals of mixed descent are classified according to whether they have 0, 1 or 2 alleles of African ancestry at this locus, then in a comparison of these three groups with other factors held constant, the mean pigmentation level will vary with the proportion of alleles at the locus that are of African ancestry. Controlling the analysis for parental admixture eliminates association of the trait with ancestry at unlinked loci and ensures that the comparison is made with other factors held constant.

[0102] To infer the ancestry of the alleles at the locus from the marker genotype, the conditional probability of each allelic state is required given the ancestry of the allele (ancestry specific allele frequencies), e.g., West African or European. There is growing evidence that admixture mapping will be an effective means of gene identification, and it has been reported that, in admixed populations, strong allelic association is observed between linked markers spaced at substantial distances (Parra et al., supra, 1998; Parra et al., Amer. J. Phys. Anthropol. 114:18-29, 2001; McKeigue et al., supra, 2000; Lautenberger et al., supra, 2000; Smith et al., supra, 2001; Wilson and Goldstein, Amer. J. Hum. Genet. 67:926-935, 2000; Pfaff et al., supra, 2001). Given the very high levels of association observed over long genetic distances, it is expected that phenotypes different between parental populations because of some genetic factor will also show associations with linked AIMs. A phenotype that is well suited to apply admixture mapping is skin pigmentation.

[0103] Notwithstanding the power of AIMs for disease gene and forensics analysis, no studies have been conducted to elucidate this power. As disclosed herein, 1) SNPs or deletion/insertion polymorphisms (collectively referred to as AIMs) in the human genome that are of potential use for drug response, disease gene or forensics research were identified; 2) biochemical and genetic test results are provided that demonstrate these AIMs can be useful for disease gene and forensics research; 3) the usefulness of AIMs derived from systematic screens of the human genome in actual drug response, disease gene or forensics research is demonstrated; 4) the usefulness of AIMs derived from systematic screens of the human genome to make an inference as to whether an individual is susceptible to acquire a disease, or to not respond to a drug, is demonstrated; 5) the usefulness of AIMs derived from systematic screens of the human genome to make an inference as to whether a crime scene DNA specimen was derived from an individual of, for example, an 80% European, 10% African and 10% Asian heritage or some other ratio/mix is demonstrated; 6) the usefulness of AIMs derived from systematic screens of the human genome to infer the ancestral proportions of an individual from their DNA (e.g., whether the individual is of 80% European, 10% African and 10% Asian heritage, or some other ratio/mix) is demonstrated; and 7) the usefulness of AIMs derived from systematic screens of the human genome to infer the ancestral proportions of a group of individuals from their DNA (for example, whether the group, which can be a population sample, a family, or a clinically defined group of persons, is of 80% European, 10% African and 10% Asian heritage, or some other ratio/mix) is demonstrated.

[0104] The present results demonstrate that AIMs are useful for the applications described above, and the sequences exemplified herein, as well as additional AIMs identified using the methods disclosed herein, enable these applications. The AIMs and methods of the invention are useful for the study of human diseases, drug response, and physical traits and, therefore, provide exceptional commercial potential. For example, in this dawning era of personalized drug prescription and disease risk assessment, the markers and methods of the invention provide the tools needed to proceed in this fledgling industry. As exemplified herein, an individual's response to a particular medication was dependent on the degree to which that individual exhibited a certain population structure (i.e., was of certain ancestral heritage) in

addition to, but irrespective of, the person's genotype for drug target or xenobiotic metabolism gene sequences. As such, the compositions and methods of the invention provide a means to predict an individual's likelihood to respond to a particular drug.

For example, in screen of genetic markers associated with patient response to the [0105] cholesterol lowering drug, LipitorTM, in terms of low-density lipoprotein (LDL) response, which is an indicator of favorable response, some of the most powerful markers identified for LDL response to Lipitor™ were gene types that are not immediately recognized as relevant for drug response, including, for example, TYR, OCA2, TYRP, FDPS, and HMGCR (see, also, Intl. Publ. No. WO 03/002721 (PCT/US02/20847), and Intl. Publ. No. WO 03/045227 (PCT/US02/38345), each of which is incorporated herein by reference). When combined with markers from genes that are biologically relevant for response, they augment the ability to make accurate inferences of response from the DNA. Each of these markers also is an excellent AIM, indicating that the linkage of the AIMs to drug response is likely a function of ancestral differences in response proclivity (see Example 5). As such, ancestral heritage can be predictive of favorable response to LipitorTM. This association has been observed for almost every type of response (n = 54) to almost every type of drug (n = 23) examined, thus confirming that the inference of drug response can be accomplished, at least in part, through the inference of ancestral proportions. As such, it appears that the genes truly relevant for drug response are a function, at least in part, of individual ancestry, and that the gene sequences relevant for drug response are statistically linked with markers that are informative as to ancestry (i.e., AIMs).

[0106] Screening genomes for the true identity of genes associated with a particular trait such as drug responsiveness is extraordinarily expensive and time consuming. As such, the use of AIMs for making inferences about individual proclivity to drugs provides a significant short-cut for the rapid development of tests that can be used to match patients with those drugs most appropriate for their genetic constitution. Thus, in addition to being useful for the admixture mapping of disease genes, the disclosed methods and exemplified markers provide tools that can direct treatment protocols by clinicians. The identification of AIMs from publicly available human genome data, and the ability to effectively use the AIMs for the

development of patient-drug classification sets, admixture screening panels and forensics tools, was accomplished using the disclosed method, including screening the SNP database (see, for example, world wide web ("www") at URL "nih.ncbi.nlm.gov") for AIMs; screening the AIMs against a multi-ancestral panel of DNA samples to verify those that, indeed, are good AIMs; using the disclosed statistical and software methods for using the AIM sequences to make biologically relevant inferences; and recognizing that an individual's likelihood to respond to a drug or develop a disease can be predicted through a knowledge of their ancestry, which, in turn, is indicated through the individual's AIM sequences.

Prior to the present disclosure, individual ancestry could be estimated using two [0107]independent methods: a Maximum Likelihood approach (Hanis et al, Amer. J. Phys. Anthropol. 70(4):433-441, 1986, which is incorporated herein by reference), and a Bayesian method implemented in the STRUCTURE program (Pritchard et al., Genetics 155:945-959, 2000, which is incorporated herein by reference). While the Maximum Likelihood method and the Bayesian method provide point estimates of proportional ancestry or admixture, there are several deficits in these methods that are addressed by the disclosed methods. For example, using the disclosed algorithm (see Example 6; see, also, Table 12, containing flow chart exemplifying algorithm), 1) the most likely group(s) from which the individual is derived were estimated simultaneously with the estimate of proportional ancestry; 2) multidimensional confidence intervals were computed and projected, thus reducing the complexity for presentation; 3) an approach to estimate the number of ancestors and their admixture proportions at each level in the past (parental, grandparental, great-grandparental, etc.) was developed; and 4) proportional BGA affiliation within individuals for more than two BGA groups at a time was derived, thus providing, for example, improved and more accurate forensics applications, as well as allowing for the development of classifiers for quantitative or continuously distributed traits (i.e., not dichotomous), the trait values of which are at least in part a function of BGA.

[0108] Independent methods for classification of individuals into population groups have been developed (Shriver et al., *supra*, 1997, Frudakis et al., *supra*, 2002, each of which is incorporated herein by reference). The present methods differ from previous classification

methods in that they allow the simultaneous estimation of the best group within which a particular individual would fall, as well as the proportional assignment of the individual to multiple parental groups (Example 6; see, also, Table 12). Thus, where previous methods allowed one to make the statement that a person is much more likely to be African-American than European-American, the present approach allow the same statement, and also provides the proportional ancestry of the individual with confidence intervals (CI); e.g., 25% (95% CI 15-35%); European ancestry; 75% (95% CI 60-80%) African ancestry; and 0% (95% CI 0-6%) Native American ancestry. Further, the confidence intervals can be expressed in multidimensional space to provide a clearer representation of the ancestry measured for the person in question (see below; see, also, Figure 2). Though methods for constructing such a representation were known, the present disclosure is the first to provide for the representations to be presented with quantifiable confidence.

There are clear differences in the patterns of chromosomal segment ancestry [0109] (PCSA) among persons with different ancestral histories (see Figure 1). A series of AIMs across the chromosomes can facilitate the estimation of the most likely parental combinations that lead to the profile of sequences observed in a given person. One example of where estimates of PCSA is important is in the discrimination of persons of Hispanic ancestry from those having primarily European ancestry with some proportion of recent Native American ancestry. Indeed, this is an important determination as the political and legal rights claimed by and provided to these two groups can depend on their ancestry. Hispanic populations such as Mexican-Americans (MA) have approximately 30-40% Native American ancestry, while the balance is European with a minor portion (5% or so) African ancestry. A person who is one quarter Native American will have 25% Native American ancestry and, therefore, will overlap with many MA persons in his level of estimated ancestry. It is expected that PCSA patterns will be significantly different for these two cases and may provide some of the only genetic evidence that would facilitate an accurate definition of the ancestry in such a case. As disclosed herein, PCSA can be used in ancestry studies.

[0110] An important step in these determinations is the phasing of the AIMs along chromosomal segments (see Example 2, Figure 8; see, also Example 5, Figures 12 to 16).

46

Phasing AIMs along the chromosome can be accomplished by several methods, including 1) estimation from the genotypes of the individual, 2) molecular haplotyping (e.g., allele specific PCR combined with genotyping), and 3) single sperm analysis (for female subjects the sperm of a male full sibling would provide the same profile). In addition, the disclosed methods allow simultaneous consideration of the two sex chromosomes (X and Y) and the mtDNA for ancestral inferences. AIMs are found on each of these sources, and can be informative for many of the questions regarding the ancestral proportions of a person and the population(s) from which a particular person is derived. For example, Hispanic/Latino populations have very high (65-100%) frequencies of Native American mtDNA haplogroups, while showing only a minority contribution from Native American populations in autosomal markers. Thus, for example, a person with reputed Native American ancestry on her father's side, with a non-Native American mtDNA haplogroup, is more likely not Hispanic than partially Native American as she may suspect, than were she to have a Native American mtDNA haplogroup.

[0111] Linkage disequilibrium (LD) is increasingly being used as a mapping tool for both fine-scale determination of gene position and for the initial localization of disease genes in special populations. Allelic associations are significantly non-random and correlated with physical distance within small (< 60 kb) genomic regions (see Jorde, Amer. J. Hum. Genet. 66:979-988, 1995; Jorde, Genome Res. 10:1435-1444, 2000, for review), possibly reflecting an underlying "block structure" that characterizes many genomic regions (Reich et al., 2001; Daly et al., 2001). Thus, if disease alleles in a population share a recent common origin, nearby genetic markers with the strongest associations will be closest to the disease-causing locus. This approach has been important in the positional cloning of several simple Mendelian diseases, including the cystic fibrosis gene, the Huntington's disease gene, and the diastrophic dysplasia gene.

[0112] In addition to applications in fine-mapping or positional cloning, LD can be used for initial disease gene mapping in homogeneous populations that have undergone recent increases in size or are genetically inbred. In such populations, disease alleles were probably present in a small number of founders, and recombination has had limited opportunity to

47

randomize associations between these alleles and alleles at linked marker loci. An analysis of allelic associations between affected and unaffected individuals from these populations can thus facilitate the localization of the disease locus. A number of Mendelian diseases have been mapped using this approach: several diseases in the Finish population, Hischsprung's disease in Mennonites, benign recurrent intrahepatic cholestasis in an isolated Dutch fishing community, familial persistent hyperinsulinemic hypoglycemia of infancy in a consanguineous group of Saudi Arabian families, and Bardet-Biedl syndrome in Bedouins.

[0113] There has been much debate as to which populations will be best suited for LD mapping of complex polygenic diseases (see, e.g., Wright et al., supra, 1999; Eaves et al., supra, 2000; Nordborg and Tavaré, supra, 2002; Kaessmann et al., supra, 2002). The extent of LD is a complex function of a number of genetic and evolutionary factors such as mutation, recombination and gene conversion rates, demographic and selective events, and the age of the mutation itself. Some of these factors affect the whole genome while others only affect particular genome regions. Additionally, variation of mutation, recombination and gene conversion rates throughout the genome is expected to create LD differences between genome regions.

LD-based methods, it has been proposed that small, isolated and inbred populations will have advantages over other populations, due to the lower heterogeneity and the larger extent of linkage disequilibrium (see, e.g., Wright et al., supra, 1999; Nordborg and Tavaré, supra, 2002; Kaessmann et al., supra, 2002). On the other side, admixed populations such as Hispanics and African Americans offer the advantage that linkage disequilibrium has been created recently due to the admixture process, and it can extend over large chromosomal regions, although it is extremely important to control for the genetic structure present in these populations in order to avoid false positives (Parra et al., supra, 1998; Lautenberger et al., supra, 2000; Pfaff et al., supra, 2001; Nordborg and Tavaré, supra, 2002). Despite the increased research focus in LD based methods, however, many issues regarding LD in human populations remain largely unexplored. Currently the NHGRI is organizing a systematic project to help develop informational tools for gene identification studies by identifying the

common haplotypes in several populations. This "Haplotype Map Project" (HMP) will likely be a large-scale multicenter effort focused on finding common haplotypes in general population samples. The HMP will likely prove to be an important data resource for identifying AIMs as disclosed herein because several populations will be investigated for haplotype block structure, thus providing additional candidate AIMs and a basic plan for the fine scale LD structure in some of the parental populations.

[0115] Admixture mapping as disclosed herein is complementary to, but distinct from, the HMP. First, the primary focus of the HMP is to understand the fine scale structure of individual genomic regions throughout the genome, whereas the present methods allows an understanding of the LD that results specifically from admixture. The level of LD from admixture is on the order of millions of bases (Mb; megabases) and tens of Mb, while the HMP is focused on the level of 10's to 100's of kilobases (kb), and genomic and population features that affect the results from one project may not be noted in the other. Second, admixture mapping require accurate parental allele frequency estimates. As such, a large number of different African, Native American, European, and Asian populations have been typed (see Table 6, below), while the HMP will likely focus on one or two samples of the major population groups.

[0116] Third, large samples (n = 500) of African-Americans and Hispanics have been typed, thus providing sufficient statistical power to test the coverage of the admixture map and to compare analytical methods. In addition, several representative populations from different regions of the country were typed so that geographical variation in ancestral proportions and admixture dynamics can be examined. Although some admixed populations will likely be included in the HMP, the numbers of individuals and numbers of different population samples being discussed are fewer than those as disclosed herein and, therefore, will not allow the same comparisons. For example, having a sample of 10 for each of 4 ancestral groups is not adequate for the identification of sequences present preferentially in one or some of those groups; as disclosed herein, at least 50 individuals were tested for each of several tens of ancestral groups (not just four) in order to comprehensively identify these markers.

49

[0117] Fourth, the focus of current population variation efforts (e.g., the SNP Consortium allele frequency project) and, very likely, the HMP has been on East Asian, African, and European samples to the exclusion of Native American populations for a number of complex reasons. The exclusion of these populations, however, results in a deficit in an understanding of the genetics of the fastest growing group of US resident populations, i.e., Hispanics, who have a significant level of Native American ancestry (20% to 40%). With the markers and methods disclosed herein, the disease genetics of Hispanic populations can be examined. Similarly, several diverse Native American populations may represent important parental populations for the numerous distinct groups often grouped together as Hispanic.

[0118] The population-based association methods disclosed herein provide several advantages over traditional linkage studies. Localizing disease genes by traditional genetic linkage methods relies on the use of related persons, either extended multigenerational families or pairs of related individuals. These approaches are effective and very powerful when investigating diseases caused by single genes. However, polygenic and multifactorial diseases like Type 2 diabetes, hypertension, and prostate cancer result from the interaction of several genes and multiple environmental influences, and are more difficult to study using traditional methods. The identification of genes contributing to susceptibility to common disease is complicated by heterogeneity. The source of the genetic heterogeneity determines which mapping methods are most likely to work for gene identification. Two primary types of genetic heterogeneity are locus heterogeneity, wherein more than one locus is affecting a genetic trait, and allelic heterogeneity, wherein within a particular causative locus there are multiple alleles that are important in altering the phenotype. Traditional linkage analysis using extended families is generally insensitive to allelic heterogeneity, but can be adversely affected by locus heterogeneity. Alternatively, LD based methods are generally adversely affected by allele heterogeneity, but less affected by locus heterogeneity.

[0119] Provided there is little allelic heterogeneity, association-based approaches like measured genotype and transmission disequilibrium test (TDT) may be more sensitive than family-based LOD score or sib-pair methods. Risch and Merikangas (*supra*, 1996) compared the number of individuals needed for sib-pair studies and TDT studies, and showed that the

50

number of individuals needed to detect linkage is much smaller for TDT than for sib-pair studies. This is especially true when the disease locus has a small effect. For example, for a locus with risk ratio of 2.0 and a gene frequency of 50%, 2500 sib-pairs or 340 case/parents for TDT would be required. There are some examples in which the demonstration of association using Haplotype Relative Risk (HRR) or case/control design, or linkage with TDT, has preceded the demonstration of linkage with sib-pairs. A classic example is the association between the insulin gene and IDDM, which was demonstrated in cases and controls, then confirmed using TDT, but often not observed in sib-pair linkage studies (reviewed in Spielman et al., Amer. J. Hum. Genet. 28:317-331, 1993). Yaouanq et al. (Science, 1997) reported very significant (p < 10⁻⁹) evidence for linkage between the HLA and multiple sclerosis using TDT in a series of 157 French families (99 simplex and 58 multiplex). When the 58 multiplex families were analyzed alone, p values of 0.0001 and 0.03 resulted for TDT and sib-pair methods, respectively.

[0120] Although association studies based on candidate genes have relatively higher power to detect disease genes than linkage analysis in families (Risch and Merikangas, supra, 1996), thoroughly testing all the genes in a genome with over 40,000 genes is currently not practical. The Haplotype Map Project may succeed in creating the informational resources necessary to perform gene identification based on linkage disequilibrium. However, even if the block structure models of the human genome can be explained by four haplotypes in each gene, the minimum number of SNPs and DIPs would then be 80,000 and the actual number likely higher. Although genotyping technologies are advancing rapidly, typing this number of markers in a large number of research subjects is not yet practical. Additionally, there are some important assumptions that are implicit in plans to identify genes using LD in large populations. One important difficulty using linkage disequilibrium in genome-wide screening is that LD decays exponentially with the recombination fraction between the marker and the disease locus and with the age of the disease-causing mutations. For older mutations that predispose to diseases, LD becomes very weak even between the disease allele and alleles at relatively closely spaced marker loci.

[0121] LD mapping has been useful in mapping of rare genetic diseases such as cystic fibrosis and diseases in special populations like the Finns and Bedouins, populations that have been subject to significant population bottlenecks, inbreeding, or founder effects. In these situations, LD exists because the variant allele is relatively young, as in the case of cystic fibrosis, or the population has reduced genetic variability, which elevates the LD throughout the genome. A leading model for the genetics of common disease stipulates predisposing alleles at a number of loci which, when present in particular combinations. increase an individuals risk (Greenberg, Amer. J. Hum. Genet. 52:135-143, 1993; Lander and Schork, Science 265:2037-2048, 1994; Risch and Merikangas, supra, 1996). If the disease is common, then for this model, the predisposing alleles also are expected to be at relatively high frequencies. However, assuming the neutral model, the frequency of an allele in a population is on average related to the age of the allele such that more frequent alleles are older than rare alleles. This fact poses a problem for the application of LD-based methods to identify common disease genes in populations that are not isolated or inbred since, in homogeneous populations, the LD is inversely related to the age of the allele and risk alleles for common disease are expected on average to be relatively old.

[0122] The application of the compositions and methods of the present invention for admixture mapping allows for a more precise and reliable mapping of complex traits. Admixture mapping takes advantage of the LD created when previously isolated populations admix, and can circumvent these problems in mapping complex traits. It was first recognized that admixed populations could be useful in determining genetic linkage by Chakraborty and Weiss (supra, 1988). When genetically divergent populations hybridize, non-random allelic associations result among loci that have significant allele frequency differentials, even among unlinked loci. This LD quickly decays when the genetic loci in question are not located close together on the same chromosome.

[0123] LD decays as a function of the recombination rate (θ) between the two markers and the number of generations (n) since their hybridization, and can be represented as $D_n = (1-\theta)^n$ D_0 , where D_n is the linkage disequilibrium n generations after hybridization and D_0 is the initial linkage disequilibrium (Chakraborty and Weiss, *supra*, 1988). Given this exponential

52

relationship between the decrease in LD and genetic distance, it is possible to discriminate between LD in an admixed population (if the time since admixture is short) that remains high because markers are close together, genetically linked, and background linkage disequilibrium among unlinked loci. For example, after 10 generations, the linkage disequilibrium at unlinked loci is reduced to 0.1% of the initial level, while at loci 10 cM and 1 cM apart, the disequilibrium due to true linkage will still be 34.9% and 90.4%, respectively, of the initial level. The critical parameters for effective detection of linkage in an admixed population identified are the frequency differential (δ) between the parental populations and the number of generations since hybridization. Linkage by association analysis in admixed populations worked efficiently if δ was large (not less than 0.2) and the number of generations since admixture small (on the order of 10 generations; Chakraborty and Weiss, supra, 1988).

[0124] Stephens et al. (supra, 1994) and Briscoe et al. (supra, 1994) studied this approach using computer simulations (MALD) and detailed practical considerations for study design. Using simple models of the type of admixture that has occurred in the Americas, they suggested that using sample sizes of 200-300 patients, typed for 200-300 evenly spaced markers, each having $\delta > 0.3$, one would have > 95% chance of locating the causative gene. A consistent result from the several models studied was a primary dependence of the power of MALD on the allele frequency differentials of the markers used. If δ is small, the initial LD will be small and difficult to distinguish from the background noise.

[0125] Stephens et al. (supra, 1994) suggested using loci where $\delta > 0.4$ between the admixed parental populations for effective admixture mapping. They also demonstrate that admixture mapping is most effective in populations that hybridized between 4 and 20 generations ago, and that incremental admixture (the slow introgression of one population into another; also known as the continuous gene flow model) affects the power of admixture mapping, but not critically, provided there has been no new introgression from the parental populations in the past three generations. The disclosed admixture mapping technique can identify the location of disease susceptibility genes by the analysis of admixed populations composed of parental populations where there is a large difference in the frequency of the

53

susceptible genotype. As such, applications of admixture mapping include the study of Type 2 diabetes susceptibility in Pacific Island populations, hypertension obesity, and prostate cancer in African Americans, and Type 2 diabetes, obesity and gallbladder disease in Hispanic populations.

[0126] McKeigue developed an approach to exploit admixture in mapping genes that builds on earlier work (McKeigue, supra, 1997; McKeigue, supra, 1998, McKeigue et al., supra, 2000). Although the approach is powered by the LD that is generated by admixture, it is more analogous to linkage analysis of an experimental cross. For this reason, the term "admixture mapping" was proposed. Instead of testing for allelic associations, one can model the underlying variation in ancestry on chromosomes of mixed descent to extract all the information about linkage that is generated by admixture.

[0127] As discussed above, advanced statistical methods are required to apply this approach in practice. Conditioning on parental admixture eliminates association of the trait with ancestry at unlinked loci and ensures that the comparison is made with other factors held constant. In non-statistical language, a comparison is made in each individual of the proportion of alleles at the marker locus that are of a particular descent with the expected proportion given the admixture of that individual's parents. One simple way to do this is to use the Analysis of Covariance (ANCOVA) test (see Tables 2 and 3), though this simpler approach does not use all of the information available. As such, Bayesian methods also have been used (see Tables 2 and 3).

[0128] To infer the ancestry of the alleles at the locus from the marker genotype, the ancestry-specific allele frequencies are required; i.e., the conditional probability of each allelic state given the ancestry of the allele (West African or European, in this example). The total population of alleles at any locus in the admixed population can be considered to be made up of two subpopulations - alleles of African ancestry and alleles of European ancestry. As long as the ancestry-specific allele frequencies are correctly specified for the admixed population under study, Bayes' theorem can be applied to invert these conditional probabilities and calculate the posterior distribution of ancestry at the locus (0, 1 or 2 alleles of African ancestry) for each individual under study. If the information conveyed by typing a

WO 2004/016768

single marker is not sufficient to assign the ancestry of each allele at the marker locus to one of the two founding populations, markers can be combined in a multipoint analysis to estimate ancestry at adjacent loci.

[0129] Simulation studies showed that, with enough markers, a high proportion of information about ancestry at each locus can be extracted even though no single marker is fully informative for ancestry (McKeigue, supra, 1998). Based on these simulations, panels of markers with Fst >0.4 at an average spacing of 2-3 cM for a total of 1,000 AIMs can be constructed as disclosed herein. It should be recognized that the panel of 1,000 AIMs for a particular population (e.g., an African-American group that is primarily West African and European) will often overlap with panels for other groups. In other words, it is often the case that AIMs selected for one level of distinction (e.g., African/European) are also informative for other distinctions (e.g., Native American/European). Table 1 lists an initially identified panel that includes of 32 AIMs (SEQ ID NOS:332 to 363; see, also, Example 1). Using a cutoff of d > 0.3, only four of these markers are restricted in informativeness to one of the three comparisons (African/European; African/Native American; Native American/European); the rest are informative for two of the comparisons, and one marker is informative for all three comparisons. In a further study, a panel of 71 AIMs was identified (SEQ ID NOS:1 to 71; Table 6) that are informative as to IndoEuropean, sub-Saharan African, Native American, and East Indian (see Example 2).

[0130] There is growing evidence that admixture mapping will be an effective means of gene identification. At least three independent groups have reported strong admixture linkage disequilibrium (ALD) between linked markers spaced at substantial distances (see, e.g., Parra et al., supra, 1998 and 2001; Pfaff et al., supra, 2001; McKeigue et al., supra, 2000). Given the very high levels of association that have been observed over long genetic distances, it is expected that phenotypes dramatically different between parental populations because of some genetic difference will also show associations with linked AIMs. However, as promising as that MALD approach appears, until the present disclosure, no systematic screen has been reported identifying SNP based versions of the AIMs required. McKeigue and others have identified panels of STR AIMs for use with this approach, but the use of

55

STRs for this purpose is problematic because of the allelic complexity of STRs and the massive databases required in order to accurately estimate allele frequencies. Even small errors or faulty assumptions on the frequencies of unobserved alleles can amplify to cripple the statistical power of a study.

[0131] Heterogeneity within the parental populations can have a confounding effect on admixture mapping studies. In the case of African-American populations, the process of admixture that took place in the New World involved a heterogeneous group of populations mainly from West-Central Africa and Europe, as well as some Native American populations. Regarding the European genetic contribution, the most important source populations came from Great Britain, Ireland, Germany and Italy. In spite of the diverse geographical areas of origin of the parental European populations, it is important to indicate the relative homogeneity of European populations from the genetic point of view (see for example Cavalli-Sforza et al., supra, 1994).

[0132] With respect to the African contribution, it is well known that the African continent contains a tremendous amount of genetic diversity. However, only a subset of the African genetic diversity contributed to the formation of African-American populations. The majority of enslaved Africans came from West-Central Africa, approximately from Senegal in the North, to Angola in the South (Curtin, In The Atlantic Slave Trade; Madison, University of Wisconsin Press 1969); other areas of Africa were not affected by the slave trade. Of the four main linguistic families present in Africa, Niger-Congo Kordofanian, Nilo-Saharan, Afro-Asiatic and Khoisan (Greenberg, supra, 1963), the majority of enslaved Africans forcefully brought to the New World were members of the Niger-Congo family. This widespread family encompasses West African languages (spoken by peoples from Senegal to Nigeria) and Bantu languages (dominant in Central and Southern Africa). The Bantu languages were dispersed throughout Africa by a "recent" expansion that took place about 3,000 years ago, and probably originated in West Africa (Nigeria and Cameroon; see Excoffier et al., Yearbook Phys. Anthropol. 30:151-194, 1987 and Cavalli-Sforza et al., supra, 1994). This recent origin is reflected in the linguistic and genetic homogeneity of the Bantu (Excoffier et al., supra, 1987, Weber et al., supra, 2000). Thus, the available

historical, linguistic and genetic evidence indicate that only a subset of the diversity found in sub-Saharan Africa has contributed to the African-American gene pool, and that potential problems of heterogeneity are much less than if the diversity of the whole continent of Africa were represented in contemporary African-American populations. Unfortunately, the extent of the heterogeneity present in West and Central Africa remains largely unknown due to the lack of available information for the populations of this area.

[0133] Since the extent of heterogeneity within European populations and within West and Central Africa remains largely unknown, potential effects of heterogeneity need to be addressed, particularly when considering an admixture mapping approach. There are two levels at which heterogeneity can affect an admixture mapping effort. First, heterogeneity can lead to erroneous estimates of the parental frequencies for the markers used in the map, thus biasing the estimate of admixture. Given that the goal of admixture mapping is to infer linkage conditioning on parental admixture, it is important to avoid misspecification of the ancestry-specific allele frequencies, because this could affect the final outcome of the analysis. Second, heterogeneity can affect the number of loci for the phenotype being studied.

[0134] The effect of heterogeneity in biasing the estimates of the genetic contributions to an admixed population can be reduced by selecting markers showing homogeneity within the main parental populations (Europeans and Africans). In this way, the problem of contribution of different geographical areas to the parental populations is minimized, reducing the bias in admixture estimates. This strategy was implemented in previous admixture studies (Parra et al., supra, 1998, 2001; Pfaff et al., supra, 2001), wherein potentially informative markers in different European and African populations were systematically analyzed. As an example, currently, to test for heterogeneity within Africa, each potentially informative marker was genotyped in samples from five African populations, two from Nigeria, two from Sierra Leone and one from Central African republic, and the markers showing significant heterogeneity were excluded from the analysis (see below). All of these samples came from areas that were affected by the slave trade. If desired, a sample from Angola, which is a region that was the source of around 40% of enslaved Africans, can

57

be incorporated, thus providing another sample of African parental populations. In addition to this strategy, it is important to note that there are statistical methods to test for misspecification of parental frequencies (see, e.g., McKeigue et al., *supra*, 2000).

[0135] With respect to the potential problem of heterogeneity in the phenotypes being studied, it is expected that heterogeneity due to the presence of multiple genes (locus heterogeneity) affecting a phenotype will reduce the power of admixture mapping to detect significant genotypic effects, as it does with any other mapping method. Heterogeneity also can be due to multiple functional alleles within a particular gene (allelic heterogeneity). One example is MC1R, where approximately six relatively common variants lead to red hair, freckles, and pale skin among Native Europeans and their descendant populations. Within Europeans these variants are on different haplotype backgrounds, thus decreasing the power to detect an effect of the MC1R gene in association studies relative to the case where a single mutation had occurred and risen to high frequencies. However, in an admixed population (e.g., European/African), these variants will all be in allelic association with markers informative for ancestry (e.g., the MC1R marker, see Table 1) and, since they all have the effect of lightening the skin, their information will be compounded making the identification of MC1R by admixture mapping no different with six functional variants than were there only one functional variant unique to Europeans. So long as the effects of functional variants within a particular parental population are in the same direction (for example, in lowering the risk of disease), allelic heterogeneity will not be a serious problem in admixture mapping.

[0136] The majority (80-90%) of genetic variation among human individuals is interindividual; only 10-20% of the variation is due to population differences (e.g., Nei, supra, 1987; Cavalli-Sforza et al., supra, 1994, Deka et al., supra, 1995). Most populations share alleles and those alleles that are most frequent in one population generally are also frequent in others. There are very few classical (blood group, serum protein, and immunological) or DNA genetic markers which are either population-specific or have large frequency differentials among geographically and ethnically defined populations (Roychodhury and Nei, supra, 1988; Cavalli-Sforza et al., supra, 1994). Despite this apparent lack of unique genetic markers, there are marked physical and physiological differences among human

58

populations that presumably reflect long-term adaptation to unique ecological conditions, random genetic drift, and sex selection. In contemporary populations, these differences are evident both in morphological differences between ethnic groups and in differences in susceptibility and resistance to disease.

[0137] The most useful unique alleles for admixture and mapping studies are those that also have large differences in allele frequency among populations (Reed, supra, 1973; Chakraborty et al., supra, 1992; Stephens et al., supra, 1994). The fact that they are totally absent from all other populations does simplify some of the statistical computations, and can facilitate more confident parental allele frequency estimates, but is not the primary reason for their utility. The designation population-specific alleles (PSAs) was initially used to describe genetic markers with large allele frequency differentials between populations (Shriver et al., supra, 1997; Parra et al., supra, 1998), but these markers are now referred to by the more correct and descriptive term, Ancestry Informative Markers (AIMs). For a biallelic marker, the frequency differential (δ) is equal to p_x - p_y , which is equal to q_y - q_x , where p_x and p_y are the frequencies of one allele in populations X and Y and qx and qy are the frequencies of the other. Median δ levels among major ethnic groups range between 15% and 20%, and the vast majority (> 95%) of arbitrarily identified biallelic genetic markers have δ < 50% (Dean et al., supra, 1994, which is incorporated herein by reference). Statistical estimates of power in an admixture mapping study based on using markers with an Fst > 0.4 were previously presented (McKeigue et al., supra, 2000). With 1,000 such markers evenly spaced across the genome, it was demonstrated that it was possible to have a statistical power of 80% to identify a disease gene that explains a 2 fold relative risk between the parental populations.

[0138] AIMs, and their use according to a method of the invention are demonstrated in Examples 1 to 6 (below). In addition, allele frequency data for markers informative for admixture mapping in African-American and Hispanic populations have been reported (Smith et al., supra, 2001; Collins-Schramm et al., supra, 2002). In order to apply the methods of the invention to an analysis of disease predisposition or drug responsiveness, estimation of admixture proportions and admixture dynamic is important. Controlling for genetic structure in admixed populations requires knowledge of the ancestral proportions and

the genetic structure of these populations. Reliable estimates of admixture proportions can allow the informed identification of populations to consider. Since the admixture LD that is created during hybridization is dependent on the level of admixture, sampling should focus generally on those areas of the country where there has been more admixture.

[0139] In addition to knowing the ancestral proportions, it is important to understand the level of population structure present in the populations under consideration. A homogeneous population is one in which there is no assortive mating, a panmictic population in which families are formed more or less by random combination and without regard to DNA genotypes. In most large cosmopolitan populations, homogeneity is expected and found. If, however, there exists stratification within the population such that individuals do not mate at random, the population will not be homogeneous. Admixture is one of the possible mechanisms introducing genetic structure in a population, and taking into account this genetic structure facilitates admixture mapping.

The effect of genetic structure is considered at two levels. First, parental [0140] populations are evaluated to determine whether they show heterogeneity in the allele frequencies of the selected AIMs; heterogeneity can affect the estimate of admixture proportions, as discussed above. Several methods that can detect the presence of genetic structure. These methods can be grouped in two main categories, termed genomic control (GC) methods (Devlin and Roeder, supra, 1999), and structured association (SA) methods (Pritchard and Donelly, supra, 2001). Both methods require genotyping of a panel of unlinked markers to estimate and correct for the effect of genetic structure, which, as discussed above, may have been due to sampling effects, or due to real demographic strata in the sampled population. The SA method (Pritchard et al., supra, 2000; Pritchard and Donelly, supra, 2001) was used to test for genetic structure in the parental populations. This method is based on using the genotypic information provided by the unlinked markers to infer population structure, and has been implemented in a software program available from Jonathan Pritchard. In addition, to test for the presence of structure, the program estimates individual ancestry proportions, and, for the present studies, this Bayesian method was used

60

to complement the Maximum Likelihood Estimate method. These two methods produce estimates of individual ancestry that are highly correlated.

[0141] The second source of genetic structure in admixed populations is due to the admixture process itself, in which newly created linkage disequilibrium is introduced in the admixed population. AIMS, such as those exemplified herein, are particularly sensitive indicators of population structure that is related to ancestral proportions. To evaluate the presence of population structure, samples are tested for the non-random association of alleles both within a locus (Hardy-Weinberg disequilibrium) and among loci (gametic disequilibrium), and the distribution of individual ancestry estimates also is examined (see, Pfaff et al., supra, 2001; Parra et al., supra, 2001).

[0142] The history of African Americans can be traced back to 1619, when the first Africans arrived at the British colonies (Jamestown), although as early as 1526 the presence of African slaves was reported in Spanish expeditions to what would become the United States (South Carolina, Georgia, Florida and New Mexico). Institutional slavery began very soon after, but it was not until the beginning of the 18th century that the importation of slaves reached increased rates, in parallel with the demand for workers to cultivate the tobacco, indigo, and rice plantations in the southern colonies; peaks occurred in the decade from 1790-1800 and the first years of the 19th century. In 1808, slave trade became illegal but continued at a low rate for several more decades. Different estimates have been offered on the total number of slaves brought into the United States with generally accepted numbers ranging between 380,000 and 570,000.

[0143] Although it is difficult to precisely determine the ethnic origins of the African slaves, information from shipping lists has provided an approximate picture of their geographic provenance. The slave trade affected a very wide area of Western and Western-Central Africa, mainly the coastline between the present day countries of Senegal in the North and Angola in the South. The most important regions were Senegambia (Gambia and Senegal), Sierra Leone (Guinea and Sierra Leone), Windward Coast (Ivory Coast and Liberia), Gold Coast (Ghana), Bight of Benin (From the Volta river to the Benin river), Bight of Biafra (East of Benin river to Gabon), and Angola (Southwest Africa, including part

61

of Gabon, Congo and Angola). Curtin (supra, 1969) has offered, based on data on the English trade of the 18th century (the peak of the Atlantic slave trade), estimates of the proportional contribution by areas, showing that Angola and Bight of Biafra were the regions contributing the highest numbers of slaves imported into the North American mainland (around 25% each). However, there were significant differences in ethnic origin depending on the port of entry in the United States, and the figures for the Colonies of Virginia and South Carolina differed considerably.

[0144] The history of African Americans has been marked not only by the forced migration from Africa, but also by the admixture with the other ethnic groups they met when they arrived in North America, including with Europeans and Native Americans. However, few historical records address the issue of admixture. Additionally, there have been important factors that, in the time since the abolition of slavery until the present, have configured the present African-American population. Of special interest is the pattern of migration of African Americans within the United States over the past 150 years. In this sense, the redistribution of African Americans in the Southern States during the 19th century, and the Great Migration from the rural South to the urban areas in the North beginning after World War I are of particular relevance, and have had an enormous impact in defining the present distribution of the African-American population in the US (Johnson and Campbell, In Black Migration in American: A Social Demographic History; Duke University Press, Durham NC 1981).

[0145] With respect to Hispanics, the term "Hispanic" was coined mainly for governmental demographic purposes, and is generally employed to identify persons of Latin American origin or descent, living in the United States. Although this definition lumps together people with very different historical, cultural and linguistic backgrounds, this classification has been widely used. Even though Central America, the Caribbean, and South America have been for centuries under the domination of the Iberian imperial powers (Spain and Portugal), they have had quite different regional histories, both before and after the Colonial period. Populations from four continents, North and South America, Europe, and Africa, have contributed to the formation of contemporary Hispanic populations. The

anthropological background of the main three Hispanic groups currently living in the United States - Mexican Americans, Puerto Ricans and Cuban Americans, which together makeup more than 80% of the total US Hispanic population - is considered here.

[0146] Mexican Americans show the highest Amerindian contribution of the three aforementioned groups. Soon after the Spanish conquest of Mexico, at the beginning of the 16th century, intermixture of the Spanish men with Amerindian women resulted in an increasingly important mixed population (Mestizos), and this racial mixing continued through the three centuries of Spanish domination in "New Spain", configuring both biologically and culturally the Mexican population. The majority of estimates have indicated an Amerindian component in Mexican Americans ranging between 30% and 40% (Hanis et al., supra, 1986; Long et al., 1991; Hanis et al., Diabetes Care 14:618-627, 1991; Merriwether et al., Amer. J. Phys. Anthrop. 102:153-159, 1997). It is interesting to point out, as well, that some studies have shown differences in the amount of Amerindian ancestry depending on socioeconomic status (Chakraborty et al., Genet. Epidemiol. 3:435-454, 1986; Mitchell et al., Ethnicity and Disease 3:22-31, 1992). There was also a substantial African presence in the Mexico territory during the Spanish rule. Curtin (supra, 1969) has estimated the total number of Africans imported into Mexico during the entire period of Slave trade to be around 200,000. Their contribution to the Mexican gene pool, however, has been estimated to be much lower than the European and Amerindian contribution, ranging from zero to 10% (see, e.g., Hanis et al., supra, 1991).

In the Caribbean Colonies (Cuba and Puerto Rico), the situation was very different [0147] from the Mainland. The Native American population was far smaller, and was decimated by slavery and disease very soon after the first contact with the Europeans. Nevertheless, the rate of admixture during the initial phases of the colonization was high enough to result in an appreciable genetic contribution (about 18%) from the Arawaks and Caribs, the original inhabitants of the Hispanic Caribbean (Hanis et al., supra, 1991). Another distinctive feature of this region is a significant African influence, which is also reflected in many aspects of the present societies of countries like Cuba, Puerto Rico, and the Dominican Republic. African slaves were imported to work in the sugar plantations in large numbers, even outnumbering

the population of European origin (Kanellos and Perez, In Chronology of Hispanic-American history: from pre-Columbian times to the present; New York, Gale Research 1995). Accordingly, the percentage of African genetic contribution in contemporary Cubans (20%) and Puerto Ricans (37%) is significantly higher than in other Hispanic populations (Hanis et al., supra, 1991).

[0148] It is clear that race is a complex concept and, in general usage, reflects both a cultural and biological feature of a person or group of people. Given the fact that physical differences between populations are often accompanied by cultural differences, it has been difficult to separate these two elements. There has been a movement in several fields of science to oversimplify the issue declaring that race is merely a social construct. While this often can be true, depending on what aspect of variation between people is being considered, it can be false for many particular instances of differences between the populations of the world. One clear example of a biological difference is skin color. Culture or environment has almost no effect on the level of pigmentation in a person's skin. Yet there are dramatic differences across populations. Pigmentation is, of course, only skin deep and is quite simple in light of the complex environments in which we live and how these affect individual and group quality of life.

[0149] The human species is relatively young and, as a species, most likely originated in east Africa 100,000 years ago, and diverged as groups to settle the globe (Cavalli-Sforza and Cavalli-Sforza, In *The Great Human Diasporas*. *The History of Diversity and Evolution* (Perseus Books, Cambridge MA 1995). During these migrations, and in the time since, there has been some degree of independent evolution of the populations that settled the various continents of the world. The simplest evidence of this evolution is seen in the differences in allele frequencies at genetic markers. Generally, alleles that are found in one population are also found in all populations, and the alleles that are the most common in one population also are common in others. These similarities between populations highlight the recent common origin of all populations. However, there are examples of genetic markers that are different between populations and, as disclosed herein, these markers, AIMs, can be used to estimate the Ancestral origins of a person or population.

64

[0150] The present invention provides methods of estimating proportional ancestry of at least two ancestral groups of a test individual and, in particular, provides a confidence level with respect to the proportional ancestry. A method of the invention can be performed by contacting a sample, which includes nucleic acid molecules of the test individual, with hybridizing oligonucleotides that can detect nucleotide occurrences of SNPs of a panel of at least about ten AIMs that are indicative of BGA for each ancestral group examined, wherein the contacting is under conditions suitable for detecting the nucleotide occurrences of the AIMs of the test individual by the hybridizing oligonucleotides; and identifying, with a predetermined level of confidence, a population structure that correlates with the nucleotide occurrences of the AIMs of each of the ancestral groups examined, wherein the population structure is indicative of proportional ancestry.

[0151] The term "biogeographical ancestry" or "BGA" is used herein to describe the biological or genetic component of race. BGA is a simple and objective description of the ancestral origins of a person, in terms of the major population groups (e.g., Native American, East Asian, Indo-European, and sub-Saharan African). BGA estimates can represent the mixed nature of many people and populations today. In many countries, including the United States, there has been extensive mixing among populations that initially had been separate. The term "admixture" is used herein to refer to such population mixing. In this respect, BGA estimates can be understood as individual admixture proportions, which take the form of a series of percentages that add to 100%. For example, a person can have 75% Indo-European, 15% African, and 10% Native American ancestry, or can have 100% Indo-European ancestry, or the like.

[0152] The proportional ancestry estimated according to a method of the invention can be a proportion of any ancestral group, including, for example, a proportion of sub-Saharan African, Native American, IndoEuropean, East Asian, Middle Eastern, or Pacific Islander ancestral group, and generally is a combination of two or more of such ancestral groups. Thus, the proportional ancestry of a test individual can include proportional affiliation among the sub-Saharan African and IndoEuropean ancestral groups (e.g., 80% sub-Saharan African and 20% IndoEuropean; or 60% sub-Saharan African, 20% IndoEuropean, and 20% of a third

65

ancestral group); or can include proportional affiliation among the Native American and IndoEuropean ancestral groups; East Asian and Native American ancestral groups; IndoEuropean and East Asian ancestral groups; and the like.

[0153] A panel of AIMs useful for estimating proportional ancestry of an individual can include AIMs as set forth in SEQ ID NOS:1 to 331, for example, AIMs as set forth in SEQ ID NOS:1 to 71, which can be useful for determining proportional ancestries including IndoEuropean, sub-Saharan African, East Asian, and Native American; or AIMs as set forth in SEQ ID NOS:7, 21, 23, 27, 45, 54, 59, 63, and 72 to 152, which can be useful for determining proportional ancestry of East Asians and sub-Saharan Africans; or in SEO ID NOS:3, 8, 9, 11, 12, 33, 40, 59, 63, and 153 to 239, which can be useful for determining proportional ancestry of East Asians and IndoEuropeans; or in SEQ ID NOS:1, 8, 11, 21, 24, 40, 172, and 240 to 331, which can be useful for determining proportional ancestry of IndoEuropeans and sub-Saharan Africans;.

An estimate can be made, for example, of an individual's proportional ancestry [0154] with respect to three ancestral groups. In this method, identifying a population structure within an individual that correlates with the nucleotide occurrences of the AIMs of the test individual can be practiced by performing a likelihood determination for affiliation with each of a sub-Saharan African ancestral group, a Native American ancestral group, an IndoEuropean ancestral group, and an East Asian ancestral group; thereafter selecting three ancestral groups having a greatest likelihood value for the individual; determining a likelihood of all possible proportional affiliations among the three ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with the nucleotide occurrences of the AIMs of the test individual is identified; and identifying a single proportional combination of maximum likelihood. Alternatively, identifying a population structure that correlates with the nucleotide occurrences of the AIMs can be practiced by performing six two-way (binary) comparisons comprising likelihood determinations for affiliation of each group compared to each other group; thereafter selecting three ancestral groups having a greatest likelihood value across all comparisons; determining a likelihood of all possible proportional affiliations among the three ancestral

66

groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with the nucleotide occurrences of the AIMs of the test individual is identified; and identifying a single proportional combination of maximum likelihood. Such a methodology works as well for individuals of three-way admixture as individuals that are 100% affiliated with a single group.

[0155] An estimate of an individual's proportional ancestry that includes proportions of three ancestral groups also can be made by performing three three-way comparisons among the groups; determining a likelihood of all possible proportional affiliations among the three ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with the nucleotide occurrences of the AIMs of the test individual is identified; and identifying a single proportional combination of maximum likelihood. An advantage of the present methods is that a graphical representation of the comparison of the three ancestral groups can be generated, wherein the graphical representation comprises a triangle with each ancestral group independently represented by a vertex of the triangle, and wherein the maximum likelihood value of proportional affiliation for an individual comprises a point within the triangle (see Figures 2 and 3). If desired, the graphical representation can further include a confidence contour that indicates a level of confidence associated with estimating the proportional ancestry.

[0156] An estimate of an individual's proportional ancestry also can be made where the proportional ancestry includes proportions of four ancestral groups. In various aspects of this method, identifying a population structure that correlates with the nucleotide occurrences of the AIMs of the test individual is practiced by performing six two-way comparisons, or by performing three three-way comparisons, or by performing one four-way comparison among the groups; determining a likelihood of all possible proportional affiliations among the four ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with the nucleotide occurrences of the AIMs of the test individual is identified; and identifying a single proportional combination of maximum likelihood. If desired, the method can further include generating a graphical representation of the comparison of the three ancestral groups, wherein the graphical representation comprises

a pyramid with each ancestral group independently represented by a vertex of the pyramid, and wherein the maximum likelihood value of proportional affiliation for an individual comprises a point within the pyramid. If desired, the graphical representation can further include a confidence contour comprising a sphere around the point, wherein the sphere indicates a level of confidence associated with estimating the proportional ancestry.

[0157] As disclosed herein, such methods are useful, for example, as a forensic tool. The present methods provide substantially greater information for forensics because, using a DNA sample obtained at a crime scene, the methods can provide an investigator with prospective information as to the likelihood of an individuals ancestry, as well as hair, skin and eye pigmentation. In comparison, present DNA methods only allow provide retrospective information because they require that a DNA sample from a crime scene be compared with DNA samples contained in a database or taken from specific individuals. Thus, while the latter methods can provide confirmation that a suspect is likely the perpetrator of a crime, they provide no useful information until the suspect is apprehended, except in cases where the suspect's DNA sample already has been entered into a database.

[0158] The methods of estimating proportional ancestry of a test individual as disclosed herein also provide a tool that can supplement genealogical information, which generally is based on relationships established using geopolitical information (see Example 3). For example, the present methods provide information that can be used to generate an ancestral map of the world, wherein locations of populations having a proportional ancestry corresponding to the proportional ancestry of the test individual are indicated on the ancestral map. As such, the method can further include overlaying the ancestral map with a genealogical map, wherein the genealogical map indicates locations of populations having geopolitical relevance with respect to the test individual, and statistically combining the information of the ancestral map and genealogical map to obtain a most likely estimate of family history of the test individual.

[0159] Identifying a population structure that correlates with the nucleotide occurrences of the AIMs, according to a method of the invention, can be performed by comparing the nucleotide occurrences of the AIMs of the test individual with known proportional ancestries

corresponding to nucleotide occurrences of AIMs indicative of BGA. The known proportional ancestries corresponding to nucleotide occurrences of AIMs indicative of BGA can be contained in a table or other list, and the nucleotide occurrences of the test individual can be compared to the table or list visually, or can be contained database, and the comparison can be made electronically, for example, using a computer. A particularly useful application of a method of the invention involves associating known proportional ancestries corresponding to nucleotide occurrences of AIMs indicative of BGA of individuals, with a photograph of a person from whom the known proportional ancestry was determined, thus providing a means to further infer physical characteristics of a test individual. In one aspect, the photograph is a digital photograph, which comprises digital information that can be contained in a database that can further contain a plurality of such digital information of digital photographs, each of which is associated with a known proportional ancestry corresponding to nucleotide occurrences of AIMs indicative of BGA of the person in the photographs.

[0160] A method of the invention can further include identifying a photograph of a person having a proportional ancestry corresponding to the proportional ancestry of the test individual. Such identifying can be done by manually looking through one or more files of photographs, wherein the photographs are organized, for example, according to the nucleotide occurrences of AIMs of the person in the photograph. Identifying the photograph also can be performed by scanning a database comprising a plurality of files, each file containing digital information corresponding to a digital photograph of a person having a known proportional ancestry, and identifying at least one photograph of a person having nucleotide occurrences of AIMs indicative of BGA that correspond to the nucleotide occurrences of AIMs indicative of BGA of the test individual.

[0161] According to the present invention, BGA can be determined using any of several variations of the disclosed BGA test, including three BGA tests referred to as the ANCESTRYbyDNATM 1.0 test, the ANCESTRYbyDNATM 2.0 test, and the ANCESTRYbyDNATM 3.0 test (DNAPrint genomics, Inc.; Sarasota FL), which utilize selected panel of Ancestry Informative Markers (AIMs) that have been characterized in a

69

large number of well-defined population samples. The AIMs are selected on the basis of a showing of substantial differences in frequency between population groups and, as such, provide information as to the origin of a particular person whose ancestry is otherwise unknown. For example, the Duffy Null allele (FY*0) is very common (approaching fixation or an allele frequency of 100%) in all sub-Saharan African populations, but is not found outside of Africa. Thus, a person with this allele is very likely to have some level of African ancestry. Upon analysis of AIMs in a DNA sample from a person of unknown origin, a likelihood (or probability) can be determined that the person is derived from particular parental populations by calculating all of the possible mixes of parental populations. The population (or combination of populations) where the likelihood is the highest is taken as the best estimate of the ancestral proportions of the person; confidence intervals on these point estimates of ancestral proportions are also calculated.

[0162] An objective assessment of the biological component of human ancestry provides important knowledge about the person whose DNA is examined. For example, an analysis of the biological component of ancestry can elucidate health disparities by identifying, for example, genetic contributions to the higher rates of hypertension and diabetes in African Americans, or the higher rates of dementia in European Americans. Estimates of BGA also can help connect individuals separated by adoption or some other event with their ancestral populations. And even if a person is not particularly motivated to reconnect with ancestors, he or she can uncover the past of their family, for example, to verify family legends or identify forgotten roots. Because the disclosed method is based on an analysis of DNA, it provides a personal demographics tool, which, unlike a census, can provide highly accurate demographics data.

[0163] There are several commercially available tests that analyze mitochondrial DNA (mtDNA) or Y chromosome markers, and have been promoted as a means of learning one's ancestral origins. Although these tests can provide information regarding the provenance of some of a person's ancestors, the tests are very limited. For example, one generation ago a person has two ancestors, one mother and one father; five generations ago, a person has 32 ancestors; while 10 generations ago, a person has 1024 ancestors. Ten generations is roughly

70

250 years and well within the time frame of genealogical interest, especially when considering, for example, the settlement of North America. Because the mtDNA and Y chromosome tests only look at a small portion of the genome (the matrilineal and patrilineal lineages, respectively), they can only provide information relating to a very small proportion of a person's ancestors. The BGA test of the invention utilizes sequences throughout a person's genome and, therefore, can provide information about a greater number of ancestors.

[0164] Accordingly, the present invention provides a method of estimating, with a predetermined level of confidence, proportional ancestry of at least two ancestral groups of a test individual. Such a method, referred to as a "biogeographical ancestry test" or "BGA test", can be performed, for example, by contacting a sample, which includes nucleic acid molecules of the test individual, with hybridizing oligonucleotides that can detect nucleotide occurrences of SNPs of a panel of at least about ten AIMs that are indicative of BGA for each ancestral group examined, wherein the contacting is under conditions suitable for detecting the nucleotide occurrences of the AIMs of the test individual by the hybridizing oligonucleotides; and identifying, with a predetermined level of confidence, a population structure that correlates with the nucleotide occurrences of the AIMs of each of the ancestral groups examined, wherein the population structure is indicative of proportional ancestry.

[0165] As used herein, the term "proportional ancestry" refers to the percent contribution of each (if more than one) ancestral group to which an individual belongs. The proportional ancestry estimated according to a method of the invention can be a proportion of any ancestral group, including, for example, a proportion of sub-Saharan African, Native American, IndoEuropean, East Asian, Middle Eastern, or Pacific Islander ancestral group, and generally is a combination of two or more of such ancestral groups. Thus, the proportional ancestry of a test individual can include proportions of sub-Saharan African and IndoEuropean ancestral groups (e.g., 80% sub-Saharan African and 20% IndoEuropean; or 60% sub-Saharan African, 20% IndoEuropean, and 20% of a third ancestral group); or can include proportions of Native American and IndoEuropean ancestral groups; East Asian and Native American ancestral groups; IndoEuropean and East Asian ancestral groups; and the like. Similarly, the proportional ancestry can include proportions of Native American, East

Asian, and IndoEuropean ancestral groups; sub-Saharan African, Native American, and IndoEuropean ancestral groups; sub-Saharan African, Native American, and East Asian ancestral groups; and the like.

[0166] A panel of AIMs useful for estimating proportional ancestry of an individual can include AIMs as set forth in SEQ ID NOS:1 to 331, for example, AIMs as set forth in SEQ ID NOS:1 to 71, which can be useful for determining proportional ancestries including IndoEuropean, sub-Saharan African, East Asian, and Native American. For example, the AIMs as set forth in SEQ ID NOS:7, 21, 23, 27, 45, 54, 59, 63, and 72 to 152 can be useful for determining proportional ancestry of East Asians and sub-Saharan Africans; the AIMs as set forth in SEQ ID NOS:3, 8, 9, 11, 12, 33, 40, 59, 63, and 153 to 239 can be useful for determining proportional ancestry of East Asians and IndoEuropeans; and the AIMs as set forth in SEQ ID NOS:1, 8, 11, 21, 24, 40, 172, and 240 to 331 can be useful for determining proportional ancestry of IndoEuropeans and sub-Saharan Africans.

[0167] The ANCESTRYbyDNATM 1.0 test (DNAPrint genomics, Inc.) is a first version of the BGA test that was specifically designed to provide information on the proportions of ancestry at the continental level. As such, the ANCESTRYbyDNATM 1.0 test allowed information to be obtained as to levels of Native American, European, and African ancestry, as three component groups. The ANCESTRYbyDNATM 2.0 test, in comparison, provides information on the proportions of ancestry at the continental level for most continents, including Native American, Indo-European (includes European, Middle Eastern and South Asian groups such as Indians), African, and East Asian (which includes Pacific Islanders, and can distinguish ancestries within Asia and the Pacific Rim. The ANCESTRYbyDNATM 3.0 test can further define the levels of ancestry within continents, for example, by distinguishing Japanese from Chinese, or Northern European from Middle Eastern, thus providing greater insight into where within a particular continent a person's ancestors were derived.

[0168] For the ANCESTRYbyDNATM 2.0 test, a logical grouping into four BGA delineations was made, wherein South Asian, Middle Eastern and European are grouped into a single group called IndoEuropean (see Example 2). This grouping was based on anthropological evidence and cultural connections between these groups (e.g., their languages

are derived from a common base). The results disclosed herein demonstrate that these groups are far more similar to one another in genetic sequence content than to other groups. The ANCESTRYbyDNATM 2.0 test also performs more accurately when Pacific Islanders are grouped with East Asians. As such, the four groupings used in the ANCESTRYbyDNATM 2.0 test include 1) Native American (i.e., those who migrated to inhabit South and North America); 2) IndoEuropean (Europeans, Middle Easterners and South Asians such as Indians; 3) East Asians (Japanese, Chinese, Koreans, Pacific Islanders); and 4) Africans (sub-Saharan). The ANCESTRYbyDNATM 3.0 test can further distinguish between South Asian and European, and between Pacific Islander and East Asian, thus providing 6 proportions (Native American, European, African, South Asian, East Asian and Pacific Islander), although the confidence intervals are larger than those obtained with the ANCESTRYbyDNATM 2.0 test. Further improvement to the tests are provided, wherein the confidence intervals are reduced. Confidence intervals around a point estimate can be reduced, thus increasing the accuracy of the test, by analyzing a complementary panel, thereby improving the confidence intervals by about 50%.

[0169] The algorithm used to determine the ancestral proportions was developed based on the idea that it is possible to use certain statistical methods to make an inference of the proportionality of ancestry in an individual sample based on their sequence (see Example 6; see, also, Table 12). The method of making this inference using the present algorithm is similar to those of others, wherein, if the frequency of an allele in a population is known, and this frequency is significantly different from population to population, a "Maximum Likelihood Estimation" (MLE) can be used to determine the probability that a person with the allele belongs to one of the groups. Expanded to include multiple alleles from multiple genetic loci and multiple populations, the process is the same. By way of simplification, Bayes' theorem states that the probability of an event given a circumstance (called a posterior probability) is a function of the frequency of the circumstance given the event (a conditional probability) and the frequency of the event itself (the prior probability). By determining the probability of the event given the circumstance for a wide range of possible events, that with the highest probability can be selected, thus obtaining the MLE for the probability.

73

[0170] In the present algorithm, the event is a proportionality of ancestry, and the circumstance is the genotype of the individual. If the minor allele frequency for 10 SNPs in 2 populations of human beings is known, and the sequence of a person at each of the 10 SNPs is known, a simple binary classification into one of the two groups can be made by choosing that for which the conditional probability is highest. This would offer little improvement over current methods for determining the BGA from a DNA sample. What is provided by the present invention is the ability to obtain the proportionality of ancestry for more complex and realistic scenarios of ancestry. There are many possible combinations such as 99%, African, 1% European, 0 Native American, 0% East Asian; or 98% African, 1% European, 1% Native American, 0% East Asian; and the like. The posterior probabilities for each of the thousands of possibilities are not the same for any particular individual, given his or her multilocus genotype (i.e., genotypes of many AIMs), and, in fact, there is one that has the highest posterior probability or likelihood for each genotype. It is this combination that the present algorithm selects (i.e., the MLE).

[0171]Previous methods have been limited in that the confidence of the estimate was not known. The present algorithm addresses this limitation by plotting the MLE graphically, including plotting the confidence regions around the MLE such that a level of confidence can be ascertained (see Figures 2 and 3). Further, the algorithm (i.e., the software code) that performs the MLE calculation operates in an unusually efficient manner. The triangle plot provided by the algorithm is an original method to graphically represent the MLE calculations and their confidence intervals. To read a triangle plot (see below), a perpendicular line is dropped from each vertex (triangle point) of the triangle to the opposite edge (base) of the triangle (see Figure 2A). In this figure, the circle represents the MLE, and a line has been dropped from the Native American (NAM) vertex to the line below; the line serves as a scale for the percentage of Native American ancestry, from 0% at the base to 100% at the vertex (or tip). Projecting the circle on this line can be analogized to holding a flashlight to the right of the triangle at the same level as the circle and observing the shadow the circle makes on the line. Where this shadow falls on the line indicates the percentage of Native American ancestry. In this example, the individual is about 15% Native American, as indicated by the hash mark on the line.

The results provided using the disclosed method provide a statistical estimate of [0172] BGA admixture for an individual (the Maximum Likelihood Estimate (MLE)), which is indicated as a point on a triangle plot to represent the proportions of the most relevant three groups for the individual. While the MLE is the most likely estimate, the true value for the individual can be a different set of proportions. A triangle plot with calculated and plotted estimates that are 2 times, 5 times and 10 times less likely than the MLE is exemplified. The first contour around the MLE delimits the space within which the estimates are up to 2 times less likely, with those positions near the line reflecting values close to 2 and those near the MLE closer to 1; the second contour around the MLE delimits the space within which the estimates are 5 times less likely in the same graded fashion proceeding from the first contour line to the second contour line. The third contour delimits the space within which the estimates are from 5 fold (near the second contour line) up to 10 times less likely (near the third contour line). The greater the number of DNA positions read, the closer these contour lines approach the MLE point. On the triangle plot, the likelihood (probability) that the true value is represented by a different point than the MLE increases until the MLE is met, where the probability is maximum (i.e., the Maximum Likelihood Estimate; MLE). The test can be performed so that the contour lines are very close to the MLE by sequencing a very large collection of markers. However, to keep the test affordable and efficient, the survey can be limited to a desired number of markers (e.g., 10, 15, 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or more) that is sufficient to determine the most likely proportions with good confidence. In this respect, a variety of different panels of 100 SNP markers have been examined, a panel of 71 AIMs has been used in a number of studies, and a panel of 175 AIMs is being examined such that very confidence is achieved.

[0173] The BGA test of the invention has been validated by determining the frequency of DNA sequence variants in various human populations. In addition, the test has been evaluated using a large number of people from a wide range of ancestral groups, and the estimates have corresponded well to what is known from anthropological and historical data. For example, Hispanics are known to have arisen as an ethnic group from the blending of colonial Europeans with Native Americans, and the hundreds of Hispanics examined using the BGA test aligned with these two groups almost exclusively. As another example, though

75

Nigerians plot as of almost pure African BGA, African Americans plot more as a mixture between this group and Europeans, which is what would be expected from knowledge about the admixture between Africans and Europeans in the United States.

[0174] The method also was validated through pedigree challenge (see Example 1); i.e., when the BGA is determined from a mother and father, that of their children should plot somewhere between the two. Numerous family pedigrees have been examined using the test, and the ancestral proportions of offspring have always plotted between those of the child's parents. When the MLE estimates are tested objectively (blindly), they prove to be excellent estimates of ancestral proportions. For example, the data for a European American man, whose mother is European mix and father is mostly Greek, showed the man to be of 85% European ancestry, but also of 15% Native American ancestry (Example 1). In fact, his paternal great-grandmother was full-blood Cherokee, thus confirming the result of the test (based on the laws of genetics, the man would be predicted to have a 12% Native American Ancestry if his great-grandmother was 100% Native American and none of his other relatives were of Native American ancestry). Further, the man's wife is Mexican, and she was determined to be of mostly Native American, but with some Native American and African heritage. This was also expected based on what is known from anthropological origin of Hispanics, who derived from the union of Spanish explorers with Native Americans in Colonial Caribbean and Latin America. Each of the three children of the man and woman plotted roughly half way between both parents, as expected. None of the children showed any Asian or Pacific Islander ancestry, which would have been impossible (assuming an accurate test) because none of the parents showed any significant Asian or Pacific Islander heritage, and none of the children were found to have more African ancestry than their mother, which would also be impossible given the fact that the father has virtually none. Thus, the results of the children were consistent with those of the parents, and the MLE values were accurate estimates when tested against what was known from biographical data.

[0175] The genotypes (nucleotide letters) determined to date are quite accurate. Because the latest genetic reading equipment available is used, an accuracy greater than 99% accuracy is routinely achieved for each site. If an accurate value was not obtained for a particular site

in a particular sample, an "FL" is indicated, instead of the genotype letters for that site. Having a few FL's generally does not prevent a good ancestry estimate. A sample can produce an FL for a site because, for example, a small region of the chromosome around this site is missing or is of different sequence character than for most (this result is not uncommon given the highly variable nature of the chromosomal positions we measure); or because not enough DNA was obtained from the buccal swab used to collect a DNA sample.

[0176] The genome was scanned for a useful panel of BGA AIMs and the best 71 AIMs were identified using the maximum likelihood algorithm to measure BGA admixture proportions were selected (Table 6). Using these AIMS, majority BGA affiliations were measurable in a manner consistent with self-held notions on BGA, and BGA admixture proportions were measurable with significantly improved precision, accuracy and reliability compared to previously described methods for the inference of race (see Example 2; see, also, Example 1, using 32 marker test). This test can be used during study design to help reduce or eliminate the insidious effects that cryptic or micro population structure imposes. The test also can be useful for forensic scientists who currently use imprecise and sometimes inaccurate means by which to infer race from crime scene DNA.

[0177] The present invention also provides articles of manufacture, including one or a plurality of photographs, each photograph being of a person having a known proportional ancestry corresponding to a population structure comprising nucleotide occurrences of AIMs indicative of BGA, the known proportional ancestry being associated with the photograph in the article. An article of manufacture of the invention (i.e., a photograph and the proportional ancestry information) can be contained in one or more files (e.g., the photograph and information in one file, or the photograph in one file and the information in a second file, which is or can be linked to the photograph). If desired, more than one photograph of an individual having a known proportional ancestry can be contained in the same or a linked file, for example, photographs containing different profiles of the individual or photographs of the individual at various ages.

[0178] Similarly, a plurality of the articles (i.e., photographs and proportional ancestry information) can be contained in a file, for example, a file containing a plurality of

77

photographs of different persons, wherein the some or all of the persons have the same or different known proportional ancestries that correspond to a population structure comprising nucleotide occurrences of AIMs indicative of BGA. Such a plurality of articles also can be contained in different files, including, for example, a plurality of files, each containing one photograph and information regarding the known proportional ancestry of the individual in the photograph, or each containing two or more photographs of different individuals, each of which contains the same known proportional ancestry, or each containing two or more photographs of different individuals, some or all of which have a different proportional ancestry as compared to another individual whose photograph is contained in the file. Accordingly, a plurality of such articles is provided, as is a plurality of files, each file of which can contain one or more articles, i.e., photographs, which can be of one or more persons having the same or different known proportional ancestries that correspond to a population structure comprising nucleotide occurrences of AIMs indicative of BGA; and the plurality of files can contain files, each of which contains one or more photographs of one or more persons, and when containing one or more photographs of two or more different persons, the different persons can have the same or different known proportional ancestries.

[0179] The article of manufacture, i.e., the photograph of a person having a known proportional ancestry corresponding to a population structure comprising nucleotide occurrences of AIMs indicative of BGA can be a digital photograph, which comprises digital information, including for the photographic image and any other information that may be relevant or desired (e.g., the age, name, or contact information of the subject in the photograph, or the subject's answer on a questionnaire as to what the subject believes his or her ancestry to be). Such digital information of one or more digital photographs can be contained in a database thus facilitating searching of the photographs and/or known proportional ancestry information using electronic means. As such, the present invention further provides a plurality of the articles of manufactures, including at least two digital photographs, each of which comprises digital information. Where the digital information for one or a plurality of the articles is contained in a database, it can comprise any medium suitable for containing such a database, including, for example, computer hardware or software, a magnetic tape, or a computer disc such as floppy disc, CD, or DVD. As such, the

78

database can be accessed through a computer, which can contain the database therein, can accept a medium containing the database, or can access the database through a wired or wireless network, e.g., an intranet or internet.

The present invention also provides kits useful for practicing a method of the [0180]invention. Such kits can contain, for example, a plurality of hybridizing oligonucleotides, each of which has a length of at least fifteen contiguous nucleotides of a polynucleotide as set forth in SEQ ID NOS:1 to 331 (or a polynucleotide complementary thereto), the plurality including at least five (e.g., 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, etc.) of such oligonucleotides, each based on different polynucleotides as set forth in SEQ ID NOS:1 to 331. In one embodiment, the hybridizing oligonucleotides that include at least fifteen contiguous nucleotides of at least five polynucleotides as set forth in SEQ ID NOS:1 to 71, or polynucleotides complementary to any of SEQ ID NOS:1 to 71. In another embodiment, the hybridizing oligonucleotides are specific for at least ten AIMs as set forth in SEQ ID NOS:1 to 71. A kit of the invention also can contain at least two panels of such hybridizing oligonucleotide, including, for example, a panel of at least five (e.g., 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, etc.) hybridizing oligonucleotides specific for AIMs as set forth in SEQ ID NOS:7, 21, 23, 27, 45, 54, 59, 63, and 72 to 152; or a panel of at least five hybridizing oligonucleotides specific for an AIM as set forth in SEQ ID NOS:3, 8, 9, 11, 12, 33, 40, 59, 63, and 153 to 239; or a panel of at least five hybridizing oligonucleotides specific for AIMs as set forth in SEQ ID NOS:1, 8, 11, 21, 24, 40, 172, and 240 to 331; or two or more of such panels and/or a panel of at least five hybridizing oligonucleotides specific for AIMs as set forth in SEQ ID NOS:1 to 71.

[0181] The hybridizing polynucleotides of a kit of the invention can include probes, which are useful for detecting a particular AIM, including a particular nucleotide occurrence at the SNP position of the AIM; can include primers, including primers useful for a primer extension reaction and primer pairs useful for a nucleic acid amplification reaction; or can include combinations of such probes and primers. A hybridizing oligonucleotide of the plurality can, but need not, include a nucleotide corresponding to nucleotide position of the SNP or DIP of an AIM, e.g., nucleotide 50 of an AIM as set forth in any of SEQ ID NOS:1

79

to 55 and 57 to 331 or nucleotide 26 of SEQ ID NO:56, or to a nucleotide sequence complementary thereto, such a hybridizing oligonucleotide being useful as a probe to identify the presence or absence of a particular nucleotide occurrence at the SNP position of the AIM.

[0182] A kit of the invention also can contain at least one pair of hybridizing oligonucleotides useful for detecting the nucleotide occurrence at the SNP position or the presence or absence of a nucleotide sequence the DIP position of an AIM. For example, a pair of hybridizing oligonucleotides can include one oligonucleotide that hybridizes upstream and adjacent to the SNP position of an AIM and a second oligonucleotide that hybridizes downstream of and adjacent to the SNP position of the AIM, wherein one or the other of the pair further contains a nucleotide complementary to a nucleotide occurrence suspected of being at the SNP position of the AIM (i.e., one of the polymorphic nucleotides), such a pair of hybridizing oligonucleotides being useful in an oligonucleotide ligation assay. In another example, a pair of hybridizing oligonucleotides can include an amplification primer pair, including a forward primer and a reverse primer, such a pair of hybridizing oligonucleotides being useful for amplifying a portion of polynucleotide that includes the SNP or DIP position of the AIM.

[0183] A kit of the invention can further contain additional reagents useful for practicing a method of the invention. As such, the kit can contain one or more polynucleotides comprising an AIM, including, for example, a polynucleotide containing an AIM for which a hybridizing oligonucleotide or pair of hybridizing oligonucleotides of the kit is designed to detect, such polynucleotide(s) being useful as controls. Further, hybridizing oligonucleotides of the kit can be detectably labeled, or the kit can contain reagents useful for detectably labeling one or more of the hybridizing oligonucleotides of the kit, including different detectable labels that can be used to differentially label the hybridizing oligonucleotides; such a kit can further include reagents for linking the label to hybridizing oligonucleotides, or for detecting the labeled oligonucleotide, or the like. A kit of the invention also can contain, for example, a polymerase, particularly where hybridizing oligonucleotides of the kit include primers or amplification primer pairs; or a ligase, where the kit contains hybridizing oligonucleotides useful for an oligonucleotide ligation assay. In addition, the kit can contain

appropriate buffers, deoxyribonucleotide triphosphates, etc., depending, for example, on the particular hybridizing oligonucleotides contained in the kit and the purpose for which the kit is being provided.

[0184] The following examples are intended to illustrate but not limit the invention.

EXAMPLE 1

DETERMINATION OF BIOGEOGRAPHICAL ANCESTRY USING ANCESTRY INFORMATIVE MARKERS

[0185] This Example demonstrates that a panel of 32 Ancestry Informative Markers (AIMs) allows an estimate of the genetic contribution from populations of African, European and Native American ancestry.

[0186] The AIMs used in the exemplified study include single nucleotide polymorphisms (SNPs), deletion/insertion polymorphisms (DIPs) and Alu sequences (see Example 2 for identification of AIMs). Markers showing differences between the parental populations greater than 30% were selected (Table 1; see, also, SEQ ID NOS:332-363). Informative genetic markers were identified by testing each candidate marker in a panel of European (Spanish, and German), African (from Nigeria, Sierra Leone, and Central African Republic), and Native American populations (Mayan and South Western Native Americans) to confirm the usefulness of the marker for admixture estimation.

TABLE 1
Ancestry Informative Marker Panel

MARKER LOCATION	Mb	AF/EU	AF/NA	EU/NA
MID 575 (356*) 1p34.3	~42	0.130	0.417	0.546
MID 187 (357) 1p32	~50.2	0.370	0.440	0.070
FY-NULL (339) 1q23.2	~181	0.999	0.999	0.000
AT3 (359) 1q25.1	~196	0.575	0.777	0.202
F13B (338) 1q31.3	~220	0.641	0.674	0.033
TSC1102055 1q32.1	~234.5	0.441	0.303	0.744
(343)				
WI-11392 (NS**) 1q42.2	~269.5	0.444	0.256	0.188
WI-16857 (345) 2p16.1	~56.2	0.536	0.548	0.012
WI-11153 (346) 3p12.1	~95.0	0.652	0.022	0.629
GC*1F (NS) 4q13.3	~75.7	0.697	0.530	0.166
GC*1S (NS) 4q13.3	~75.7	0.538	0.478	0.060
MID-52 (NS) 4q24	~110.7	0.186	0.500	0.687
SGC30610 (354) 5q11.2	~61.5	0.146	0.281	0.427
SGC30055 (355) 5q22.1	~124.7	0.457	0.675	0.218
WI-17163 (347) 5q33.1	~173.9	0.120	0.641	0.521
WI-9231 (348) 7p22.3	~1.2	0.017	0.387	0.370
WI-4019 (349) 7q21.3	~100	0.124	0.173	0.296
CYP3A4 (NS) 7q22.1	~101.9	0.761	0.755	0.006
LPL (340) 8p21.3	~22.3	0.479	0.521	0.042
CRH (NS) 8q13.2	~73.2	0.609	0.655	0.046
WI-11909 (350) 9q21.31	~81.0	0.075	0.587	0.663
D11S429 (337) 11q13.3	~70.4	0.429	0.054	0.376
TYR (344) 11q14.3	~95.4	0.444	0.055	0.389
DRD2-Taq I "D" 11q23.2 (336)	~125.0	0.535	0.046	0.582
DRD2-Bel I (335) 11q23.2	~125.0	0.080	0.565	0.485
APOA1 (360) 11q23.3	~128.9	0.505	0.555	0.050

82

GNB3 (332)	12p13.31	~7.2	0.463	0.430	0.033
RB1 (361)	13q14.2	~47.4	0.611	0.711	0.100
OCA2 (342)	15q12	~24.0	0.631	0.369	0.263
WI-14319 (351)	15q14 .	~30.0	0.185	0.310	0.494
CYP19 (334)	15q21.2	~47.6	0.045	0.379	0.423
PV92 (362)	16q23.3	~96.5	0.073	0.551	0.624
MC1R314 (341)	16q24.3	~103.8	0.350	0.441	0.090
WI-14867 (352)	17p13.2	~3.5	0.448	0.404	0.045
WI-7423 (353)	17p13.1	~8.2	0.476	0.074	0.402
Sb19.3 (363)	19p13.11	~27.0	0.488	0.236	0.253
CKM (333)	19q13.2	~55.8	0.150	0.694	0.545
MID 154 (358)	20q11.23	~34	0.444	0.368	0.076
MID 93 (NS)	22q13.2	~38.6	0.554	0.179	0.733

Shown are the marker name and chromosomal band, approximate location of the marker on the chromosome in megabases (Mb), and difference in frequency between African and European populations (AF/EU), African and Native Americans (AF/NA) and European and Native Americans (EU/NA). Differences greater than 30% are marked in bold letters (see, also, Shriver et al., *supra*, 2003, which is incorporated herein by reference)

[0187] The publicly available human genome sequence database and polymorphism database were screened in order to identify SNPs that met the criteria for being a good AIM. Allele frequencies are available for many of the SNPs in the public databases for three populations – Africans, Europeans and Asians. Since these frequencies are obtained from small samples they are not always accurate. The main criteria for selection herein was the delta value that derived from using these frequencies, which is a statistical measure of the difference in minor allele frequency between various populations of human beings. For example, a C or a G polymorphism at a particular place in the human genome, where the C is present mainly in individuals of European descent and the G present mainly in individuals of Native American descent, would have a high delta value and, therefore, qualify as a good AIM. Similarly, an A or a C polymorphism at a particular place in the human genome, where

^{*} Numbers in parentheses are SEQ ID NO: for AIMs; NS - sequences not shown.

83

the A is present mainly in individuals of African descent and the C present mainly in individuals of Asian descent would have a large frequency differential between these groups and, therefore, a high delta value, thus qualifying as a good AIM. A list of such "candidate AIMs was compiled, ranked from largest delta value to smallest delta value for each of the possible pair-wise population comparisons, and screened, one at a time, against a panel of "parental" samples. Parental samples are samples from regions of the world that are relatively homogeneous, for example, Niger or Congo for sub-Saharan Africans, Southern Mexico for Native Americans, China for East Asians, and Europe for Europeans.

[0188] About half of the candidate AIMs proved to be not very useful because their actual delta values were not as high as expected from the public database allele frequencies (some were not even SNPs, or could not be assayed using the present platform). Sequences that were validated as true AIMs, such as those exemplified herein, were useful for admixture mapping, making inferences of individual ancestry proportions, and making inferences of population group admixture proportions, as well as for screening genomes in order to identify markers with alleles that correlated with certain human traits through their ancestry informativeness. Even though each candidate AIM was initially selected from the public database based on crude population structure differences (i.e., continental populations), many of them were found to carry information on finer levels of structure because the separation of subgroups of humans from larger groups throughout human evolution has provided a fertile opportunity for genetic drift, founder effects, and natural selection to operate in either fixing or eliminating their sequences.

[0189] Sequences are shown in the Sequence Listing from 5' to 3' (left to right), and, for SEQ ID NOS:1 to 331, with the SNP generally, but not always, at nucleotide position 50 from the 5' terminus (except for SEQ ID NO:56, position 26). The polymorphism is indicated with an IUB symbol, wherein S=C/G, Y=C/T, R=A/G, K=G/T, W=A/T, etc. As such, the disclosed sequences (SEQ ID NOS:1 to 331) provide information as to the target being examined (i.e., the polymorphism) as well as information for preparing primers and amplification primer pairs, and hybridization probes, for sampling the SNP (i.e., determining

the genotype of a sample). Further, the disclosed sequences can be used, if desired, to scan public databases to identify additional upstream and downstream nucleotide sequences.

[0190] This panel of markers was extremely powerful for estimating with precision admixture proportions in population samples (standard error typically between 1% and 5%). In addition, the AIMs provided reasonable estimates of individual ancestry, and suggested that an equivalent precision can be obtained using more markers (confirmed in Example 2). Two independent methods, a Maximum Likelihood Estimate (MLE) method (Chakraborty et al., supra, 1986) and a Bayesian method using the program STRUCTURE (Pritchard et al., supra, 2000) estimating individual ancestry, were used; the values obtained by both methods were highly correlated ($R^2 = 0.9836$) when estimates of individual ancestry were compared in terms of percent African genetic contribution in a sample of African Americans from Washington DC. These markers are excellent for determining whether there is population structure in a sample from an admixed population. This ability is important in terms of admixture mapping applications because, as discussed below, the process of admixture can produce significant structure in a population, and consequently a high number of false positive results (positive associations caused by genetic structure, not by physical linkage of a marker with a disease causative gene), increasing significantly the risk of misinterpreting mapping results.

[0191] The present study confirms that AIMs can be identified using the disclosed methods, and provides a panel of 32 AIMs that can be applied towards an ultimate goal of compiling a panel of approximately 1,000 AIMs spanning the entire human genome. Candidate AIMs were obtained by screening SNP allele frequency data generated through The SNP Consortium (TSC). Six sites, including the Sanger Centre, Celera Genomics, Washington University, Orchid Biosciences, Motorola, and Whitehead Institute, have generated, as of 2003, allele frequencies on 60,000 SNPs located throughout the genome using a central collection of 42 individuals from each of 3 populations (African-American, European-American, and Asian-American). This database, which is freely available to researchers (see, e.g., using hypertext transfer protocol ("http"), at URL "snp.cshl.org"), has

been used to provide the present results, thus demonstrating the usefulness of the resource to compile a genome-wide panel of AIMs.

[0192] The present study focused on the accuracy of the SNP database and the number of candidate SNPs present therein. With respect to the accuracy of the database, each group involved in the SNP consortium has taken a different approach to generating data. As such, initial concerns regarding how the data can be combined was addressed. Because the genotyping approaches were different for each group, it was necessary to address the question of ascertainment biases that might differentially affect the data of particular groups. For example, most of the groups produced their allele frequencies after sequencing a subset of the TSC diversity panels, then scoring these markers in the larger groups of 42 individuals from the 3 populations. The Washington University group has taken an approach whereby pooled sequencing throughout regions was performed, and the allele frequencies calculated for variable positions discovered during this effort. The Orchid group has not used sequencing but, instead, started with loci from the TSC SNP database that are known to be polymorphic. Given such differences, a systematic characterization was made as the extent, if any, that different biases may have affected the results.

[0193] One approach for systematically characterizing such potential bias was to compare the allele frequencies for loci that were genotyped by more than one group. Although there was dispersion around the 45° line, as expected, there was general agreement in the frequency data obtained by the different groups (R²=0.8762), indicating that the extent of the allele frequency bias introduced by the differing genotyping and ascertainment strategies was limited. The next step in testing the accuracy of these data was to separate the data by site and perform pair-wise comparisons, which would allow the identification of particular sites that have allele frequency estimates that deviate more when compared to other sites.

[0194] With respect to the number of candidate AIMs, it also was important to determine how many of the 60,000 SNPs characterized by the TSC would be useful for admixture mapping. Since it can be useful ultimately to compile a panel of about 1,000 markers showing large frequency differences between the relevant population groups (Fst>0.4), it was important to evaluate which percentage of the available markers have the desired

characteristics. Candidate AIMs were based on the recommendation of McKeigue et al. (supra, 2000). The markers with information available for African, Asian and European populations, the cumulative proportion of markers in each Fst category (0-1, in 0.05 intervals) and the total number of candidate AIMs for each possible comparison. The distribution of pairwise Fst from the TSC allele frequency project was as follows: Asian-European (556 candidate AIMs/25,110 total SNPs; average Fst = 0.0720); Asian-African (1026 candidate AIMs/25,578 total SNPs; average Fst = 0.0886); and European-African (1306 candidate AIMs/30,103 total SNPs; average Fst = 0.0861). As such, the screen revealed that about 2-5% of the markers can be useful for admixture mapping.

[0195] The geographical pattern of admixture in the US admixed populations, particularly African Americans and Hispanics, was the subject of an initial examination. The admixture proportions of more than 18 African-American populations were characterized, and a map was generated showing an estimate of the European genetic contribution to African Americans from several different geographic areas in the United States. The European admixture ranged between 3.5% in the Gullah of South Carolina to 22.5% in New Orleans (e.g., 18.8% in Chicago; and 16.4% in Houston). Most of these estimates were obtained using an initial panel of 10 informative AIMs. The observed distribution was interpreted in terms of well known historical and demographic events that have played an important role in African American history (see Parra et al., supra, 1998, Parra et al., supra, 2001). These data allow the application of admixture mapping to identify genes involved in complex diseases. It is expected that admixture mapping will be more suitable in populations showing a high degree of admixture and, therefore, populations such as the Gullah (3.5%) and Jamaicans (6.6%), in which European genetic contribution has been very limited, may not be suitable for this kind of analysis.

[0196] In preliminary studies using mitochondrial DNA (mtDNA), it was observed that African Americans have a detectable, although low, Native American genetic contribution, in accordance with the self-reported Native American ancestry often mentioned by African-American individuals. Having identified 30 AIMs informative for African/Native American and 19 AIMs for European/Native American contrasts (see Table 1; see, also, Shriver et al.,

87

supra, 2003), the presence of Native American admixture in three African-American populations was examined using nuclear DNA markers. In accordance with the mtDNA estimates (which only provide "maternal" contribution information), evidence of a low Native American genetic contribution was detected in each of the African-American samples (Washington DC, 6%; Afro-Caribbeans from London, 5%; and Bogalusa, Louisiana, 6%).

[0197] Regarding admixture in Hispanics, the relative European, Native American and African contribution in a sample of Spanish-Americans from San Luis Valley CO was estimated. A 59% European admixture, 35% Native American admixture, and 6% African admixture was observed in this sample, in good agreement with estimates previously described for populations of Mexican ancestry (Chakraborty et al., supra, 1986; Hanis et al., supra, 1991; Tseng et al., Amer. J. Phys. Anthropol. 106:361-71.1998; Collins-Schram et al., supra, 2002). As shown in Example 2, further characterization of admixture in additional samples from Mexico and in two samples from Hispanics of Puerto Rican ancestry (New York and Puerto Rico) has been performed.

A sample of individuals of European ancestry (N=199) currently living in State [0198] College PA also were analyzed. The genetic contribution in this sample was predominantly of European origin (91%), with evidence of some African (3%) and Native American (6%) influence. These results are summarized in Figure 4, using a triangular plot, which clearly reveals the differences in average admixture levels between European Americans, Spanish Americans, and African Americans. The triangular plot shown in Figure 4 represents the average admixture estimate in a particular sample; the underlying distributions of individual ancestry are complex, with different individuals showing widely dispersed values of African, European and Native American ancestry (not shown). In African Americans, most individuals showed predominantly African genetic contribution, but some persons showed relatively high European contribution and also, to a lesser extent, Native American ancestry. European Americans clustered more tightly near the pole corresponding to high European contribution, with few persons showing evidence of Native American and African ancestry. Spanish Americans showed the highest dispersion of individual ancestry, as expected given the high admixture level observed in this sample.

88

[0199] Notably, individuals showed the whole range of European and Native American ancestry (from 100% European to 100% Native American), and a relatively lower African genetic contribution also was evident in some individuals. Some of the variance observed in individual ancestry was likely due to the stochastic error due to the limited number of markers used to infer ancestry. Thus, while the 20-32 markers used in the exemplified test detected individual ancestry, the standard error of the estimates was fairly high; increasing the number of AIMs is expected to increase the precision of the individual ancestry estimates (see Example 2). The other component of the variation in individual ancestry was due to true differences in ancestry between individuals. The remarkable correlation in individual ancestry values obtained by two totally independent methods, ML and STRUCTURE (discussed above), indicates that this panel of markers can capture the underlying individual ancestry patterns characteristic of these populations. As disclosed below, controlling for variations in individual ancestry allows the avoidance of false positive results.

[0200] The effect of admixture dynamics in population structure and the extent of linkage disequilibrium (LD) was examined. The importance of the admixture model (hybrid isolation model vs. continuous gene flow model) in terms of the population structure and LD created in the admixture process was previously described (Pfaff et al., supra, 2001), and two methods to quantify the level of population structure in admixed populations were presented. Population structure is a key aspect of admixture mapping, as well as of any genetic association study in an admixed population. This issue has been explored in an African-American, a Spanish-American and a European-American sample using more informative markers than previously available.

[0201] The presence of structure was evaluated in two different ways. First, the observed number of significant associations was compared between unlinked markers with the number expected at the 5% significance level, and second, the average correlation of individual ancestry was estimated using two subsets of genetic markers. In agreement with previously reported data, the African-American population from Washington DC showed a significant genetic structure, as reflected by a much higher number of significant associations between unlinked markers than expected by chance (10.5% vs. 5%, Figure 5A). Very strong

associations were observed between markers located as far apart as 24 Mb (AT3-F13B, G=15.21, p<0.0001), providing clear evidence that these significant associations are caused by the admixture process. The alleles that were associated were always combinations of the alleles that are frequent in African populations, with the higher the difference in frequency, the more frequently associations were observed between markers: FY, showing the highest difference in frequency between African and Europeans, was significantly associated with 9 unlinked markers. Thus, the admixture process in this African-American population, showing 17% European ancestry, created a strong association between markers showing high frequency differences between African and European populations. Although these associations are significant both when the markers are linked and when they are unlinked, linked markers tended to show higher G values than unlinked markers, indicating that the association due to true linkage can be distinguished from the association due to genetic structure, as previously demonstrated (McKeigue et al., supra, 2000). Interestingly, significant associations were observed between markers showing high frequency differences between European and African populations, but not between markers showing high frequency differences between African and Native American, or European and Native American populations. This result was likely due to the low Native American ancestry observed in this sample (6%), which was insufficient to create detectable associations due to the admixture process in such a small sample.

[0202] Another line of evidence demonstrating the high level of genetic structure present in this African-American sample was the significant correlation between independent estimates of individual ancestry using different subsets of the genetic markers. The average correlation over 100 random selections of independent subsets of markers was r=0.40, p<0.0001 (Figure 5B). The pattern of genetic structure and LD observed in Washington DC African Americans and in other African-American samples analyzed with a more limited set of markers (Jackson MI, and the Low Country counties, South Carolina, Pfaff et al., *supra*, 2001) indicated that the best model to describe the admixture process in these populations was the continuous gene flow model. Additional support for this model came from the strong correlation between D₀ (the initial expected association between markers originated by the admixture process) and D_t (the current association between markers). As shown using

computer simulations (Pfaff et al., supra, 2001), in populations following the continuous gene flow model, positive correlations between D_0 and D_t are expected and, in fact, this result was observed in African-American populations. In populations following the hybrid isolation model, significant correlations between D_0 and D_t are not expected.

The Spanish-American sample from San Luis Valley that was analyzed showed [0203] less genetic structure than any of the African-American populations. The number of observed significant associations between unlinked markers was only slightly higher than expected at a 5% significance level (7.3% vs. 5%, Figure 5A). This result was interesting considering that the Spanish-American population was considerably more admixed than the African Americans, and under the same model of admixture dynamics, would be expected to show considerably more structure. The correlation of individual ancestry estimates based on independent markers, although significant, was much lower than the values observed in the African-American populations (r=0.11, p<0.0001, Figure 5B). Also, there was no correlation of Do and Dt in the San Luis Valley sample, in contrast to the results observed in African Americans. These results demonstrate that admixture dynamics (the way in which the population was formed and has evolved) have been different in the African-American populations and the San Luis Valley population, with the former more closely resembling the continuous gene flow model than the latter. Of course, other Hispanic populations can show different patterns of admixture dynamics than that observed in San Luis Valley.

[0204] As expected from the lower admixture levels observed in European Americans, there was no evidence of genetic structure due to admixture in this sample from State College PA (Figures 5A and 5B). The number of significant associations between unlinked markers was similar to the value expected by chance, and there was no correlation between individual ancestry estimates of independent subsets of markers (p=0.149, NS).

[0205] These results demonstrate that the use of selected genetic markers (AIMs) allows an analysis of the dynamics of the admixture process and the effect of this admixture process on the pattern of LD in admixed populations. In admixed populations that have had an admixture process similar to the hybrid isolation model (initial admixture followed by independent evolution of the admixed population without further genetic contribution of the

parental populations), few false positive results are expected (recalling, that the falseness is relevant for a "gene hunter" searching for genes that cause traits through LD or linkage, not for those seeking to develop classification tools). In admixed populations that more closely resemble the continuous gene flow model (continuous genetic contribution each generation from one of the parental populations to the admixed population), the LD is expected to extend much longer distances and problems with false positive results will arise. Fortunately for the gene hunter, the information conveyed by the AIMs can control for genetic structure and minimize false positives. An example is provided below demonstrating how such control can be achieved using appropriate statistical methods and skin pigmentation as a model phenotype.

[0206] Skin pigmentation and individual ancestry was examined in an African-American sample and a Spanish-American sample. As previously demonstrated, the genetic structure created by admixture can be effectively controlled, and association due to linkage can be distinguished from spurious association due to genetic structure using appropriate statistical tests (McKeigue et al., supra, 2000). In the present study, the same methods were applied in a study of skin pigmentation in two admixed samples (African Americans from Washington DC and Spanish Americans from San Luis Valley). Information on skin pigmentation was collected for each individual in both studies, and the subjects were genotyped for a panel of AIMs and individual ancestry proportions were calculated using the maximum likelihood method (Chakraborty et al., supra, 1986). Individual ancestry (% African or % Native American) was plotted against melanin index (African) or skin reflectance (Native American) for each individual. Several of the AIMs showing high differences in frequency between the parental populations were also candidate genes for pigmentation.

[0207] In the African-American sample, a strong and highly significant correlation (R²=0.1879, p<0.0001) was observed between individual ancestry and the melanin index, which measures the melanin content of the skin. Individuals with darker skin had, on average, higher levels of African ancestry. The individual ancestry estimates were based on 21 markers, and, therefore, subject to a relatively high variance, thus explaining at least some of the dispersion observed in the graph. An interesting feature of these results was the

. 92

evident decrease in variance observed in moving from the right (more African ancestry) to the left (more European ancestry). This result is consistent with the higher level of variability in skin color that is found in African populations as compared to Europeans. The high correlation observed between individual ancestry and skin pigmentation can be due to the population structure typical of African-American populations (as discussed above), and related to the limited number of genes that were used for determining the parental population differences contained within this relationship.

[0208] A similar plot was prepared for the San Luis Valley sample. Individual ancestry estimates using 15 Native American/European AIMs were plotted against pigmentation level as measured by the percent of light reflected through the PHOTOVOLT 670 Green filter. Because skin pigmentation was measured in different ways (absorbance vs. reflectance) in these two studies, the trends observed when graphed are reversed. In the Spanish-American sample, the correlation between individual ancestry and skin color also was significant (R²=0.0481, p<0.001), but lower than in the African-American sample, possibly due to the reduced genetic structure present in this sample.

[0209] Tests for differences in the average pigment levels by genotype for the AIMs typed in the African-American population sample discussed above were performed. The panel of AIMs included three candidate gene markers, OCA2, TYR, and MC1R. The analysis was performed in three alternative ways: first with no consideration of the individual ancestry estimates (ANOVA); second after conditioning to control for the effect of individual ancestry leaving out the locus under consideration (ANCOVA/IAE minus marker); and third using the complete individual ancestry estimate for the conditioning (ANCOVA/IAE). As shown in Table 2, eight of twenty-one (38%) of the markers showed significant differences (p < 0.05) among the three genotypes, including two of the four candidate gene markers (OCA2 and TYR). When using an alpha level of 0.05, only 5% of the markers tested were expected to yield significant results. As such, the finding of 38% significant difference indicates that population structure is related to both ancestry and pigmentation (Pfaff et al., supra, 2001, Parra et al., supra, 2001).

[0210] One way to remove the effects of population structure is to test for differences conditioning on the individual ancestry estimates (IAE). When the complete IAE was used to condition (ANCOVA/IAE), only one locus showed significant average differences among genotypes, OCA2, the human P gene. When a less conservative conditioning approach was taken, in which the locus under consideration was left out of the individual ancestry estimate (ANCOVA/IAE minus marker), there were four significant results: OCA2, TYR, FY, and SGC30055.

[0211] A Bayesian full probability model for admixture and marker genotypes was also set up (McKeigue et al., supra, 2000). Score tests for linkage were based on testing for an independent association of pigmentation with number of alleles of European ancestry at each locus, one at a time, in a regression model that includes individual ancestry (estimated from marker data). The 1-sided probabilities for the score tests are shown in Table 2, where three loci showed evidence of linkage to skin pigmentation at an alpha level of 0.05 {OCA2} (p = 0.005), AT3 (p = 0.027), and TYR (p = 0.033)}. To confirm these results, other markers informative for ancestry in OCA2 were identified and will be analyzed by the score test method. The concordance between these ANOVA results and the Bayesian admixture mapping results was encouraging, and both methods will benefit from the addition of new unlinked AIMs, which will increase the precision of the individual ancestry estimates.

[0212] The Spanish-American sample from the San Luis Valley CO, also was analyzed for linkage and association using the Bayesian and ANOVA methods (Table 3). This analysis included 442 individuals who were typed for 15 marker loci informative for ancestry (2 SNPs in the DRD2 gene treated as one locus). The CYP19E2 marker (located near MYO5A, a pigmentation candidate gene) showed strong evidence for linkage with the ethnic difference in skin pigmentation. However, this result should be interpreted with caution because, unless several closely linked markers informative for ancestry are used, the test for linkage is not robust to misspecification of ancestry-specific allele frequencies. SNPs around the MYO5A gene can be analyzed to confirm these preliminary results.

TABLE 2 Testing for an effect of single-locus genotypes on pigmentation in an African-American sample

Marker ¹	DELTA ²	ANOVA ³	ANCOVA ⁴	ANCOVA ⁵	Bayesian score
	AF vs		IAE minus	IAE	test probability ⁶
	EU		marker		
FY-null	0.997	0.000*	0.004*	0.396	0.106
LPL	0.479	0.000*	0.130	0.501	0.167
WI-14867	0.448	0.061	0.561	0.253	0.100
AT3	0.575	0.001*	0.090	0.479	0.027*
Sb19.3	0.488	0.497	0.784	0.993	0.402
APOA1	0.505	0.507	0.718	0.317	0.488
D11S429	0.429	0.578	0.833	0.442	0.139
RB1	0.611	0.743	0.535	0.128	0.102
OCA2	0.631	0.000*	0.001*	0.019*	0.005*
MC1R	0.428	0.601	0.482	0.590	0.486
WI-16857	0.536	0.745	0.789	0.311	0.137
WI-11153	0.652	0.026*	0.225	0.698	0.223
SGC30055	0.457	0.001*	0.048*	0.291	0.061
WI-7423	0.476	0.193	0.544	0.169	0.058
WI-11392	0.444	0.032*	0.200	0.507	0.257
MID 154	0.444	0.260	0.439	0.728	0.250
MID 187	0.370	0.121	0.302	0.296	0.450
DRD2 TAQ.D	0.553	0.226	0.273	0.247	0.471
GNB3	0.463	0.984	0.530	0.166	0.065
TYR-192	0.449	0.003*	0.014*	0.117	0.033*
GC	0.697	0.213	0.750	0.604	0.190

¹ Marker indicates the Ancestry Informative marker used in the test. Markers shown in bold and italics are candidate genes for pigmentation (viz. OCA2, MC1R, TYR).

² "DELTA" (δ) is the allele frequency difference between African and European populations.

³ Analysis of variance significance level where sex is the only covariate.

⁴ Significance level for a one-way ANCOVA analysis using individual ancestry estimates (M) where the tested locus was excluded as the covariate.

Same as three except the M is based on all 21 markers. Bayesian Admixture Mapping 1-sided probability.

TABLE 3 Testing for an effect of single-locus genotypes on pigmentation in a Spanish-American sample

Marker ¹	DELTA ² NA vs EU	ANOVA ³	ANCOVA ⁴ IAE minus marker	ANCOVA ⁵ IAE	Bayesian score test p-value ⁶
MID-575	0.546	0.240	0.383	0.248	0.93
TSC1102055	0.744	0.027*	0.023*	0.366	0.39
WI-11153	0.628	0.012*	0.036*	0.406	0.12
SGC-30610	0.427	0.093	0.133	0.565	0.13
WI-17163	0.521	0.192	0.268	0.967	0.17
WI-4019	0.296	0.875	0.762	0.319	0.93
WI-11909	0.663	0.026*	0.020*	0.146	0.13
TYR-192	0.417	0.006*	0.012*	0.129	0.07
DRD2Bcl	0.485	0.254	0.517	0.827	0.35
DRD2TaqD	0.582	0.418	0.461	0.550	0.35
D11S429	0.376	0.235	0.511	0.545	0.65
WI-14319	0.494	0.036*	0.054	0.061	0.30
CYP19E2	0.423	0.000*	0.000*	0.002*	0.001*
PV92	0.624	0.183	0.276	0.591	0.50
WI-7423	0.402	0.426	0.318	0.309	0.64
CKMM	0.545	0.257	0.569	0.579	0.55

¹ Marker indicates the Ancestry Informative marker used in the test. Markers shown in bold and italics are in or near candidate genes for pigmentation (viz. TYR-192 and CYP19E2 near MYO5A).

² "DELTA" (δ) is the allele frequency difference between Native American and European parental populations.

³ Analysis of variance significance level, where sex is the only covariate.

⁴ Significance level for a one-way ANCOVA analysis using individual ancestry estimates (M) where the tested locus was excluded as the covariate.

⁵ Same as three except the M is based on all 15 markers.

⁶ Bayesian Admixture Mapping 1-sided probability.

PAIRWISE POPULATION COMPARISON OF SNP DISTINCTION

[0213] Table 4 shows the results of genotyping and statistical analysis that demonstrate several different but important points (the sequences for each of the AIMs in Table 4 can be found by reference to Table 6, using the marker number.

TABLE 4
Pair-wise population comparisons of SNP distinction

AF-	AF-	AF-	AF-	AF-	AF-	CT-	CT-	CT-	CT-	CT-	EA-	EA-	EA	EA-	SA-	SA-	SA-	ME	ME :	PI-	
CT	EA	SA	ME]	PI .	ΑI	EA	SA	ME	PΙ	ΑI	SA	ME	-PI	ΑI	ME	ΡI	ΑI	-PI	-AI	ΑI	marker
0.4	0:6	0.5	0.4	0.7	0.3	0.3										0.2	0.3	0.3	0.2	0.4	959
		0.5							0,3					0.2		0.3	0.1	0.3	0.1	0.4	961
0.1			0.1						0.2					0.2		0.2		0.2		0.2	962
0.3			0.4				0		-	0.4								0.1			963
0.9		0.8									0.5					0.7	0.4	0.6	0.3	0.3	964
0.3			0.2			0.4					0.3			0.5	0			0.2	0.3	0.5	969
0.2			0.2			0	0	-	-	0.1	-	•	_	0.1	0	0	0.1	-	0.1	0.1	970
		0.6							0.2					0.1		0		0.2		0	971
0.5	CHRIST		0.5	~					0.4				0.1	_				0.3		0	972
	0.8	1	0.3	15.6		استنحقن			0.3	•		P***********								0	973
0.1	1111131111	1	47	1 7 TULL +	400.00	0.6			0.6	•		0.6	•					0.6		0.1	975
	0.7	•	0.5				0		0.5			0.3								0	977
0.5				-		0.1	0		0.4					0.1				0.3			978
11111925	-		0.4						0.5						0.1			0.3			979
0.6		0.5	t	1 3-1 -	P-0-1-1	•				0.1		0.2				0.1		0.2			980
0.2 0.1			0.5	0.2					0.4			0.4				_		_		0.1	986
0.1		_	0.1			0.2					0.1			0.1		0.2		0.2	-	0.2	993
0.1	_			0.1	0.1		0.2		0.2 0.2				0.1	0.1				0.1 0.2			1000
0			0.1			-		_		0.1	-		0.2		0.1			0.2		0.2 0.1	1001 1003
C				0.1	0.1				0.1							0.1		0.2	0	0.1	1003
C	-	_	0		0.1	_	_		0.1			_	0.1	-		0.1		0.1	-	0.1	1015
C		_	0	0	0.1	-	-	_	0.1		-	_				0.1			0	0.1	1013
0.7			0:7	_	_		_		0.2	CANDOTT .	_	H0.6									1022
0.5	विकास स्थान	THE RESERVE	0.5	Per 1771	25 12 12 12				0.2	-	_		-					0.3			1033
0.6	1	0.7		,	1111	1.			0.5						4	*********	•	0.6			1034
	0,5		0.2	,		i			0.2								•	0.1			1035
0.1		•		•		0.1												0.2		0	1037
0.2	2 (0.1	0	0	0	0.2	0.1		0.2	0.2		0			0.1			0	0	0	1038
0.1		0.1	0	0.1	0.1	0.2	0.2											0.1	0.1	0	1039
(0.7	0.3	0.2			P	•		0.5	0.2	2 0.4	0.5	7	Linesian 4	•					0.3	1040
0.6	3 (0.1	0.6	0.2	0.3	0.7	0.6	0.1	0.5	0.4		0.6								0.1	1041
0.5		.0.6	0.6	0.6	0.2	0.3	0.1	0.2	0.1	0.2				0.5			0.4		0.4		1042
0.3	0.5	0.3	0.4	0.6	0.4	0.2	0	• (0.3	0.1	0.3	0.2	. 0	0.2	0.1	0:3	0.1	0.2	0	0.2	1043
0.3	0.8	0.3	0.3	0.7	0.5	0.5	0	• (0.4	0.2	0.5	0.5	0.1	0.3	0	0.4	0.2	0.4	0.2	0.2	1044

AF- AF- AF- AF- AF- CT- CT-	CT- CT-CT-EA- EA- EA EA-SA-SA-SA-ME ME PI-
CT EA SA ME PI AI EA SA I	ME PI AI SA ME -PI AI ME PI AI -PI -AI AI marke
0.3 0.7 0.1 0.3 0.6 0.6 0.3 0.3	0.1 0.3 0.2 0.6 0.4 0 0.1 0.2 0.6 0.5 0.4 0.3 0.1 1047
0.2 0.5 0.2 0.1 0 5 0.1 0.4 0	
07 08 09 09 08 1 0.1 0.2	ATTENDED TO THE TOTAL TO
0.6 0.1 0.5 0.7 0.0.3 0.5 0.1	0.1 0.6 0.4 0.4 0.6 0.1 0.1 0.2 0.5 0.3 0.7 0.4 0.2 1050
0.3 0.6 0.4 0.4 0.6 0.6 0.2 0	
0.1 0.3 0.3 0.3 0 0.2 0.2 0.2	0.3 0 0.1 0 0.1 0.2 0.1 0.1 0.2 0.1 0.3 0.1 0.2 1052
0.3 0.2 0.2 0.3 0.1 0.3 0.1 0	
0.5 ### 0.3 0.4 0.3 0.3 0.5 0.2	0.2 0.3 0.3 0.3 0.4 0.3 0.3 0.1 0.1 0.1 0.1 0.1 0 1055
0.4 0.7 0.4 0.3 0.8 0.8 0.4 0	1000
0.2 0 0.1 0.1 0 0 0.2 0.1	
0 0.1 0 0.1 0.1 0.1 0.1 0	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
0.4 0.7 0.3 0.5 0.7 0.4 0.3 0.1	
0.5 0.2 0.3 0.4 0.1 0.8 0.3 0.2	Carried .
0.6 0.7 0.4 0.4 0.8 0.8 0.1 0.2	
0.2 0.2 0.2 0.2 0.2 0.2 0 0	
0 0 0 0 0 0 0	AND THE PROPERTY OF THE PARTY O
0.1 0.3 0.6 0.5 0.2 0 0.1 0.5	REFERENCE TO THE PARTY OF THE P
0.3 0 0.1 0.2 0 0.1 0.3 0.2	101.
0. 0 0.4 0.5 0 0 0 0.4	10.2
0.1 0.1 0.1 0.1 0.1 0.1 0 0	
0.4 0.1 0.4 0.6 0.5 0.7 0.3 0	120.20.
0.7 0.8 007 0.7 0.8 0.8 0.1 0	
0.5 0.5 0.6 0.4 0.5 0.3 0.1 0.1	
0.3 0.7 0.4 0.3 0.8 0.7 0.4 0	* *** *** *** *** *** *** *** *** ***
0.3 0.1 0.1 ### 0.3 0.1 0.2 0.3 0.3 0.6 0.2 0.2 0.5 0.4 0.3 0.1	
0.3 0.7 0.4 0.3 0.7 0.6 0.5 0.2	
0.3 0.5 0.3 0.2 0.5 0.4 0.2 0.1	
0.5 0.8 0.6 0.3 0.7 0.9 0.4 0.2	THE PARTY OF THE P
0 0.5 0.1 0.1 0.3 0.2 0.5 0.1	
105 07 07 08 07 07 0.2 0.3 0.1	
0.7 0.1 0.6 0.7 0.1 0.4 0.6 0.2	
0.7 0.1 0.5 0.5 0.2 0.6 0.6 0.2	
ERRORG O.1 O.DRIVING O.2 PORTION U.2	0.2 0.3 0.1 0.4 0.4 0.1 0.3 0 0.3 0.1 0.3 0.1 0.4 1008
43 36 45 45 38 39 39 16	7 41 32 30 36 10 21 5 27 24 37 27 23

"DELTA" (8) values for select AIMs from the list claimed are shown.

The AIM unique identifier is shown in the last column.

Cells having numbers in bold (not shaded) indicate good δ values; cells that are shaded with numbers in bold represent extremely high δ values.

AF-African, CT-European, EA- East Asian, SA-South Asian, ME-middle eastern, PI-Pacific Islander, AI-Native American.

98

[0214] First, Table 4 was derived from screening several hundred candidate AIMs electronically selected from the public databases, thus demonstrating that only a minority of the candidate AIMs from the public databases are real AIMs. As discussed above, the public SNP databases were electronically screened to find good candidate AIMs (since frequency data is provided for three "racial" groups, though the level of admixture for these groups is not known). Second, 384 individuals of various continental and BGA origins were genotyped at each of these sites: 70 African samples collected from Nigeria and Congo, 65 European samples collected from Northern Europe, 70 East Asian samples collected from recent immigrants to San Francisco, CA; 35 Middle Eastern samples collected from Turkey, 35 South Asian samples collected from India and 25 Pacific Islander samples collected from the Philippines and US Samoa).

[0215] A sampling of the data for about 70 AIMs that passed the screening process from 175 candidate AIMs screened is shown in Table 4. The delta (δ) value is a measure of how well the sequence for a polymorphism enables one to predict membership to one or the other group; i.e., how distinct the two populations are with regard to the sequence at this polymorphism. δ values are shown for 69 of the 175 AIMs; the other 105 had δ values of 0 for each pair-wise population comparison and, therefore, were not true AIMs. AIM 1068 in Table 4 is representative of types of failure (zeros across all pair wise comparisons – some AIMs with zeros across the population pairs are present in Table 4 because they are informative for populations not shown in this particular table). This result confirms that most of the candidate AIMs culled from the public database are not true AIMs and highlights the value of the present invention.

[0216] A relatively large investment in genotyping and analysis required to identify which candidate AIMs are true AIMs. While this process can be bypassed, e.g., one could simply genotype samples at 100 candidate AIMs and extract the data from those that prove to really be AIMs, genotyping is an expensive procedure and, as such, the waste incurred would make the test economically impractical. To develop an economical and practical test for ancestral proportions, the test must query large numbers of true AIMs. Many publicly available

candidate AIMs are not true AIMs due, for example, to low sample sizes, such that allele frequencies in the public databases are simply not very reliable, though they do offer some information. While it is expected that the frequency of true (i.e., validated) and well-characterized (i.e., population specific frequencies known with certainty) AIMs in a random collection of SNPs would be about 5%, the frequency of true AIMs in a culled set of candidate AIMs is about 50% and, after proceeding as disclosed herein, the frequency of true AIMs in a collection of SNPs is 100%.

Second, the results of Table 4 demonstrate that some of the AIMs are good for the [0217]resolution of Africans vs. Europeans, other AIMs are good for the resolution of Native Americans vs. Africans, etc. Though selected based on European/African/Asian allele frequency differentials, some AIMs provide good distinction of other groups such as Pacific Islanders, South Asians and Middle Easterners. This type of information can only be learned by genotyping in larger samples, and a test for ancestral proportions must go through this step in order to be accurate (for example, if the test only worked in the 3-dimensions allowed by data that is publicly available - European, African and Asian - the results obtained, for example, for a Hispanic would be ambiguous). The panel of SNPs in Table 4 provides a well balanced mix of AIMs with resolution power for each of the possible pair-wise comparisons for 7 population groups, and this panel would constitute a good test for ancestry proportions. Data for south Asians, Middle Easterners and Pacific Islanders do not exist in the public databases and, therefore, was generated for these studies. In comparison, one attempting to develop a test for ancestral proportions in 7-dimensions by simply sequencing at candidate AIMs haphazardly selected from the public SNP databases (i.e., without selection through data production) would need to compile a battery of thousands of SNPs to obtain a panel such as that in Table 4 because certain pairs of populations are difficult to resolve (e.g., South Asians and Europeans, which constitute a larger IndoEuropean group united by a common language base).

[0218] The results obtained using the panel of AIMs shown in Table 4 is shown in Table 5. The presently disclosed algorithm (see Example 6, Table 12) was used to calculate

100

the proportions for a group of 96 individuals who reside in the southeastern United States, and who describe themselves to be Caucasian.

TABLE 5

Ancestral proportions for a large number of self-reported Caucasians

	%	%	%	SELF	М	F	MGM	MGF	PGM	PGF	СО
EUR	97NAM	0 EAS	∜ 3	ca	ca	ca	ca	ca	ca	ca	us
EUR	89 EAS	11 NAM	0	ca	ca	ca	ca	ca	ca	ca	us
EUR	100 NAM	0 EAS	0,	ca	ca	ca	cc	ca	ca	ca	us
EUR	76 NAM	24 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR.	80 NAM	20 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR'	100 EAS	ONAM	0	ca	ca	ca	ca	ca	ca	ca	us
EUR	92 NAM	0,AFR	8	ca	ca	ca	ca	ca	ca	ca	us
EUR	95 EAS	5 NAM	∵:0	ca	ca	ca	ca	ca	ca	ca	brazil
	MAN 88	, 12 EAS	; 0 ;	ca	ca	ca	ca	ca	ca	ca	us
EUR ¹	100 NAM	0 EAS	` O	ca	ca	ca	ca	ca	ai_ca	ca	us
EUR	76 AFR	24 NAM	.0	ca_ai	ca	ca_ai	ca	ca	ai	ca	us
	100 NAM	0 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
*	100 NAM	0 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
	51 NAM	48 EAS	- 1	ca	ca	ca	ca	ca	ca	ca	us
	100 NAM	1.	į: 0 .	ca	ca	ca	ca	ca	ca	ca	us
EUR!	100 NAM	0 EAS	۰., ٥	ca	ca	ca	ca	ca	ca	ca	us
* 1	100 EAS	0 NAM	0	ca	ca	, ca	ca	ca	ca	ca	us
EUR	54 NAM	46 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR	95 NAM	5EAS	0	ca	ca	ca	_, ca	ca	ca	ca	us
	100 NAM	0 EAS	. 0	ca	ca	ca	ca	ca	ai_ca	ca	us
EUR	89 NAM	11 EAS	. 0,	ca	ca	ca	ca	ca	ca	ca	us
EUR	100 NAM	0 EAS	0	ca	ca_me	ca	ai_ca	ai_me	NULL	NULL	us
EUR	88 NAM	12 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR	100 NAM	0 EAS	0	ai_ca	ca	ai_ca	ca	ca	ai	ca ·	us
EUR	100 EAS	. ONAM	, 0	ca	ca	ca	ca	ca	ca	ca	us
EUR	76 NAM		0	ca .	ca	ca	ca	ca	ca	ca	us
EUR	· !		11	ca	hi	ca	hi	hi	ca	ca	us
	100 NAM	0 EAS	0	ca	ca	NULL	ca	ca	ai	ai	us
EUR	67:NAM	33 EAS	0	ca	ca	ca	ai_ca	ca	ca	ca	us
EUR	72 NAM	28 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR		27 EAS	0	hi	hi	hi	hi	hi	hi	hi	us
EUR	77 NAM	23 EAS	, 0	ca	ca	ca	ca	ca	ca	ca	us
EUR	•	34 NAM	0	NULL	ai_ca	ca	ca	ai-ca	ca	ca	us
EUR	99 NAM	, 1 EAS	, 0	ca	ca	ca	ca	ca	ca	ca	us
EUR		33 EAS	0.	ca	ca	ca	ca	ca	ai_ca	ca	us
EUR	100 EAS	O NAM	0	ca	ca	ca	ca	ca	ca	ca	us
EUR	100 NAM	DEAS	0	ca	ca	ca	ca	ca	ca	ca	us

	~~~				_					
%	%	%	SELF	М	F	MGM	MGF	PGM	PGF	CO
EUR 88 NAM	12 EAS	0	ca	ca	ca	00	oó.			puerto
EUR 100 NAM	0 EAS	0	ca	ca		ca	ca	ca	ca	rico
EUR, 90 EAS		0			ai_ca	ca	ca	ai_ca	ca	. us
EUR 78 NAM		. 0	ca	ca	ca	ca	ca	ca	ca	us
EUR: 100 NAM	0 EAS		ca	ca	ca	ca	ca	ca	ai_ca	us
EUR 76 NAM	,	<u>0</u> 10	ca ca	ca	ca	ca	ca	ca	ca	us
EUR 76 NAM	24 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR 100 EAS		- Q		ca	ca	ca	ca	ca	ca	us
	& 3'EAS	. 0	ca	ca	ca	ca	ca	ca	ca	us
EUR 91 NAM	9 EAS	. Ų. ∴:Ö	ca ca	ca NULL	ca NULL	ca NULL	ca	ca	ca	us
EUR 94 NAM	2EAS	4	ca				NULL	NULL	NULL	us
EUR 100 NAM	·.	0	ca	ca	ca	ca	ai	ca	ca	us
EUR 100 NAM		0		ca	ca	ca	ca	ca	ca	us
EUR 97 EAS	3 NAM	1 O	ca	ca	ca	ca	ca	ca	ca	us
EUR 100 NAM	OEAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR 100 EAS	0 LAS	, 0	ca	ca	ca ai	ca ai	ca	ca NULL	ca	us
EUR 100 DAS	0 EAS	.0	ai_ca	ai_ca			ai_ca		NULL	us
EUR 60 NAM	40 EAS	0	ca ca	ca ca	ca ca_hi	ca	ca	ca hi	ca hi	us
EUR 84 AFR	16 EAS	9				ca	ca			us
EUR 100 NAM		. 0	ca	ca	ca	ca	ca	ca	ca	us
EUR 99 NAM	1'EAS	: _'	ca	ca	ca	ca	ca	ca	ca	us
EUR 89 NAM	11 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR 67 NAM	32 EAS	-1	ca	ca	ca	ca	ca	ca	ca	us
NAM 55 EUR	*** A * * * *	3**	me	me	me	me	me	me	me	us
EUR 84 EAS	OAFR	16	ca	ca	ca	ca	ca	ca	ca	us
EUR 74 NAM		13	ca	ca	ca	ca	ca	ca NULL	ca NULL	us
EUR 88 NAM	12 EAS	. IS	ca ca	ca	ca	ca	ca	ca	ca	us
EUR 100 NAM	0EAS	. 0	-	ca	ca	ca	ca			us
EUR 98 NAM		2	ca	ca	ca	ca	ca	ca	ca	us
EUR 100 NAM		0	ca	ca	ca	ca	ca	ca	ca	us
EUR 70 NAM	30 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR 86EAS	14 NAM	0	ca ca	ca	ca	ca ca	ca ca	me ca	ca ca	us
EUR 65 NAM	16 EAS	:19	ca	ca ca	ca	_	ca	ca	ca	us
EUR 100 NAM		0.	ca	ca	ca ca	ai_ca ca	ca	ca	ca	us us
EUR 100 NAM	ONAM	0	ca	ca	ca	ca	ca	ca	ca	us
•	15 EAS		ai_ca	ai_ca	ai_ca	ai	ca	ca	ca	us
EUR 100 NAM			NULL	ca ca	ai_ca	ca	ca	NULL	NULL	us
EUR 72 NAM	• •	0	ca	ca	ca	ca	ca	ca	ai	us
	11 EAS		aa_ca					aa_ca		
EUR 84EAS	10 NAM		ca_ca	aa	aa ca	aa ca	aa_ca ca	ca_ca	aa_ca ca	us us
EUR 95 NAM	5EAS	.0	ca	ca			ca	ca	ca	us us
EUR 98 NAM		0	ca	ca	ca	ca	ca	ca	ca	
EUR 97 EAS	3 NAM		ca	ca	ca	ca	ca	ca	ca	us us
EUR 62 EAS				ca hi	ca ca	hi	hi	ca	ca	us us
UN U4'NU	I U I VANIVI	13	- Jul 111	111	va	111	, , , ,			u J

102

	%	%	%	SELF	M	F	MGM	MGF	PGM	PGF	CO
EUR	70 EAS	12 NAM	. 18	hi	hi	hi	hi	hi	hi	hi	puerto rico
EUR	74 NAM	26 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR	94 NAM	0 AFR	6	ca	ca	ca	ca	ca	ca	ca	NULL puerto
EUR:	65 EAS	11;AFR	.24	hi	hi	hi	hi	ca	hi	hi	rico
EUR	75 ¹ NAM	25 EAS	0	ca	ca	ca	ca	ca	ca	ca	us
EUR	91 NAM	9EAS	. 0	ca	ca	ca	ca	ca	ca	ca	us
EUR	100 NAM	0 EAS	. ,0	ca	ca	ca	ca	ca	ca	ca	us
EUR	71 NAM	29 EAS	0	ai_ca	ai_ca	ca	ca	ai_ca	ca	ca	us
EUR	72 NAM	28 EAS	.0	ca	ca	ca	ca	ca	ca	ca	us
EAS	56 NAM	44 EUR	0	ca	ca	ca	ca	ca	ca	ca	us

The percentage of ancestry is given after each group: EUR – IndoEuropean, NAM - Native American, EAS – East Asian/Pacific Islander, AFR – African. The self-reported race for each individual (SELF), their mother (M), father (F), maternal grandmother (MGM), maternal grandfather (MGF), paternal grandmother (PGM), paternal grandfather (PGF) and the country of their birth is shown.

[0219] The results of Table 5 demonstrate that most people who describe themselves to be Caucasian are, indeed, of majority IndoEuropean ancestry as determined using the BGA test, i.e., using the panel of markers in Table 4 and the algorithm (Example 6, Table 12). About 40% of these Caucasians were measured as 100% European ancestry, with no admixture, whereas 60% showed detectable admixture. By way of critical evaluation, the proportions obtained can be compared with self-reported admixture. For individuals of Table 5 that claim all of their parents and grandparents are unmixed Caucasians ("ca" across all columns), the rate at which 90% or greater European ancestry is found is higher (55%) for persons who report no mixture in their pedigree than for persons who report mixture in their pedigree (35%). The fact that half of those reported no mixture in their pedigree illustrates how little most people know about their BGA, at least in anthropological as compared to geopolitical terms.

[0220] The public databases employed a small number of samples for each of the three groups (African-Americans, Europeans and Asians). Thus, from the public databases, the actual allele frequencies for the claimed SNPs is uncertain and was only determined with

103

accuracy from the present work. Further, the use of African-Americans as a parental group is faulty since, as disclosed herein, they are an admixed population (between Africans and Europeans). The best way to find SNP markers that are useful for the present methods is to genotype a number of samples from the major BGA groups of the world for all of those of apparently different minor allele frequency between at least two of the groups, calculate the  $\delta$  values, and rank them.

#### FAMILY PEDIGREE

[0221] The BGA method for the use of SNPs (AIMs) to determine ancestral proportions was applied to the examination of a family pedigree of a father, mother and their 3 children (see Figures 6 and 7). The man was mostly European and his wife is Mexican, so, if the test is accurate, their three children should plot somewhere between the man and his wife and this is exactly what was observed. Three of the father's grandparents on the father's side of the family are relatively pure Greek/European, but one was almost pure Cherokee. All four of his grandparents on his mother's side were European mix. The plots for the mother (Figure 6B) and father (Figure 6A) are shown. By drawing lines from the vertices that bisect the triangle edge opposite that vertex, where the vertex represents 100% and the triangle edge 0%, it can be seen that the father is about 85% European, 11% Native American, 4% African (he was of no detectable East Asian, South Asian or Pacific Islander ancestry). The 11% Native American seems to have come from his paternal grandparent. The percent that would be predicted, knowing that 7/8 of his great-grandparents were European mix and 1/8 were Native American, is 12% (1/8), which is in good agreement with the data produced from the test. The mother is from Mexico and is of Hispanic descent. As discussed previously, Hispanics are an admixed population with European and Native American contribution. It can be seen here that the mother is of 11% European, 76% Native American and 13% African descent (there was no detectable East Asian, South Asian or Pacific Islander ancestry).

[0222] Because the three children each received one chromosome from each of their mother and father, they should each plot somewhere in between him and his wife. Using the method the children plot as expected (Figure 7). It is clear from these results that the point estimate of the children is between that of the parents, as would be expected. From these

results it seems clear that Child #1 (Figure 7A; 80% European, 18% Native American, 2% African) has ancestral proportions more similar to his father than his mother, whereas Child #2 (Figure 7B; 61% European, 31% Native American, 8% African) and Child #3 (Figure 7C; 54% European, 37% Native American, 9% African) are of ancestral proportions about half way between their two parents.

[0223] Although each child receives a chromosome from the mother and one from the father, the children are different because the mother has chromosomes of mostly Native American descent, though some have European and African flavor as well, and the father has chromosomes of mostly European flavor though some are of Native American flavor as well. When a child is conceived, he or she receive a chromosome from each parent, but which of the two the child receives from the mother is random (i.e., "independent assortment"). Since some of the mother's chromosome pairs have a member of the pair with European flavor, some of the children will receive the "European flavored chromosome" and other children will not. From the present study, it is clear that Child #1 (Figure 7A) received more of his mother's European flavored chromosomes than did the other two children. Thus, while each child is a 50/50 blend of maternal and paternal chromosomes, their ancestral proportions are unique and random functions of those of their parents.

#### **EXAMPLE 2**

#### FOUR WAY ADMIXTURE ESTIMATE OF ANCESTRY

[0224] This example demonstrates that a four way admixture BGA test provides the same results obtained using three 3 way BGA tests.

[0225] As indicated above, BioGeographical Ancestry (BGA) is the heritable component of race. Because socio-cultural and geo-political metrics for measuring human race are human, not natural, constructs, their use in genetics research makes it difficult to control for population genetic structure, and may obscure important correlations between BGA and human biology. This example provides methods and compositions to accurately measure genetic structure within individuals. The human genome was mined for candidate Ancestry Informative Markers (AIMs), which were validated on an ultra-high throughput genotyping platform and used to establish parental population allele frequencies. Using 71 of the most

informative AIMs (Table 6), which cover most of the chromosomes, and coalescing the human population to four main continental population groups (sub-Saharan African, East Asian, IndoEuropean and Native American), the MLE method was used to determine individual BGA admixture proportions and their associated confidence intervals. As disclosed herein, self-reported population affiliations correlated almost perfectly with the majority BGA population affiliation determined for a sample of 2,024 international samples. BGA admixture results were surprisingly frequent, and when observed, were generally not inconsistent with anthropological and geopolitical history. The admixture proportions produced tracked in family pedigrees in a manner consistent with the law of independent assortment, and simulation revealed that the markers relevant for resolving the group affiliations functioned independently within the confines of our algorithm. Because a large number of high  $\delta$  value markers were used, the test was surprisingly robust; reasonable levels of simulated allele frequency errors that could be caused by biased parental sampling had no significant impact on the BGA proportions determined. These results demonstrate that BGA admixture can be reliably determined from a DNA sample.

#### Sample Collection:

[0226] Parental Samples - For establishing the parental group allele frequencies, 100 relatively homogeneous descendents of four crudely defined human population groups were genotyped. These four groups corresponded to a coalescence of a simplified human pedigree back in time to a point where populations were relatively isolated to the sub-Saharan African (sub-Saharan Africans), Europe and the Middle Eastern (IndoEuropeans), North/South American (Native Americans) and East Asian (East Asians) continental regions. In essence, the extremes of the existing populations has been taken in terms of physical characteristics and known human migration patterns, and the human pedigree simplified by assuming that, because the rest of the population exhibits physical features along a continuum defined by these extremes, all of humanity arose from radiation within and admixture between these four main continental groups.

[0227] Collection efforts were focused on individuals residing in each region and of relatively homogeneous affiliation with descendents of each group in terms of self-described

106

"race"; each subject exhibited a strong physical appearance associated with descendents of each group and reported homogeneous affiliation with that group. There exists no de-facto final arbiter of BGA affiliation, or race, but collecting without regard to ethnicity might introduce systematic errors in frequency estimation for a given group if systematic admixture is a function of ethnicity. Where possible, an attempt was made to collect from as wide a variety of ethnicities as possible within each parental group, expecting admixture within each parental sample to balance out. Though it would be better to collect from individuals of known homogeneous affiliation, if sampling was not biased in terms of admixture or other population structure, the admixture extant to samples that are practical to collect from would tend to reduce the power of the test rather than introduce systematic bias. The existence of Hardy-Weinberg Equilibrium for each marker within each BGA group was relied on as an indication that a reasonably good sample was obtained. The sub-Saharan African samples were collected in Nigeria and Congo, Africa; the European samples were collected from various locales in the United States; the East Asian samples were collected from Japan and China; and the Native American samples were collected from "Nativos" inhabiting a remote region of southern Mexico. All samples were collected under IRB guidelines for the purposes of genetic studies of human population variation.

[0228] Experimental Samples - After reading and signing an approved IRB consent form, subjects completed a biographical questionnaire and provided either a buccal swab or 4 ml of blood. On the questionnaire, the subjects described themselves, their mother, father and maternal and paternal grandparents as belonging to the "African", "American Indian", "Asian", "Caucasian", "Hispanic" or "Other" group, with the option of reporting "Don't Know" for each family member. For some of the subjects, digital photographs were taken; explicit permission was received from those subjects whose photographs were presented. DNA was extracted from circulating lymphocytes or buccal swabs using commercial kits (Qiagen), and a primer extension protocol employing a 25K SNPstream ultra-high throughput (UHT) genotyping system was used (Orchid Biosciences).

107

### Estimating BioGeographical Ancestry (BGA)

[0229] A software program was written based on the algorithm of Hanis et al. (supra, 1986) for using multilocus AIM genotypes to determine the Maximum Likelihood Estimate (MLE) of individual BGA admixture (Example 6; see, also, Table 12). The delta ( $\delta$ ) value is an expression of the ancestry informativeness of the marker (Dean et al., supra, 1994). For a biallelic marker, the frequency differential ( $\delta$ ) is equal to  $p_x - p_y$ , which is equal to  $q_y - q_x$ , where  $p_x$  and  $p_y$  are the frequencies of one allele in populations X and Y and  $q_x$  and  $q_y$  are the frequencies of the other. To test the departures from independence in allelic state within and between loci, the MLD exact test was used (Zaykin et al., Genetica 96:169-78, 1995).

[0230] The collection of 71 AIMs used for in this Example was selected to maximize the cumulative δ value within, and minimize differences in the cumulative δ value between each of the six possible pairs of the four dimensional (sub-Saharan African, Native American, IndoEuropean and East Asian) problem. The algorithm inverts the population specific allele frequencies to obtain a likelihood estimate of proportional affiliation corresponding to a multilocus genotype using three groups at a time (mainly for computational convenience and because a 4-dimensional admixture is likely to be relatively rare). For example, the likelihood of 100% IndoEuropean, 0% Native American, 0% East Asian is calculated, then the likelihood of 99% IndoEuropean, 1% Native American, 0% East Asian proportions are considered, then the process is repeated for all possible IndoEuropean, Native American and East Asian proportions. The likelihood of maximum value is selected as the Maximum Likelihood Estimate (MLE).

[0231] When plotting a single MLE on a triangle plot, the space within which the likelihood is within 2-fold, 5-fold and 10-fold of the MLE is delimited (these intervals are not plotted when multiple MLEs are shown in a single triangle plot). For calculating the MLE using all four of the BGA groups together, the procedure was executed in the same exact manner; instead of 3 possible 3-way BGA combinations, only one 4-way BGA combination is possible. All of the MLEs described in this Example were calculated using the 3-way calculation scheme. Alternative versions of this type of test are possible, for example, using

108

different AIMs and different parental groups corresponding to different coalescences of the human pedigree, which would be expected to provide results meaningful with respect to a different anthropological time scale than those provided here.

At the time the database was screened, the SNP Consortium (TSC) had contributed [0232] data on approximately 27,000 SNPs where frequencies were available on three populations (African American, European American, and East Asian). This database was screened for candidate AIMs, i.e., SNPs of δ >0.40 between any two of the four continental population groups (see Example 1; see, also, Shriver et al., supra, 1997). Parental samples of sub-Saharan Africans (AA), IndoEuropeans (IE), East Asians (EA) and Native Americans (NA) were screened for each of the 200 candidate AIMs with the largest  $\delta$  values and, of these, 71 were validated as true AIMs (i.e., true SNPs), with minor allele frequency greater than 1% and of  $\delta > 0.40$  for at least one of the group pairs. The 71 AIMS are shown as SEQ ID NOS:1 to 71; the top 100 candidate AIMs for the group pairs were as follows: EA by AA (SEQ ID NOS: SEQ ID NOS:7, 21, 23, 27, 45, 54, 59, 63, and 72 to 152); EA by IE (SEQ ID NOS:3, 8, 9, 11, 12, 33, 40, 59, 63, and 153 to 239); and IE by AA (SEO ID NOS:1, 8, 11, 21, 24, 40, 172, and 240 to 331). It should be noted that some AIMs identified by one pairwise comparison also can be AIMs for a second pairwise comparison (e.g., SEQ ID NO:59 was identified as an AIM for EA by AA and EA by IU comparisons), although such AIMs are an exception. In addition, many of the 71 AIMs are not in the list of the top 100 candidate AIMs shown for any of the pairs (but were in the top 200 candidate AIMs); candidate AIMs were not used, for example, because they did not genotype well due to the SNP type of amplification parameters used for the exemplified platform, or for other reasons as disclosed herein.

109

TABLE 6  $\label{eq:pair-wise} \mbox{ Pair-wise $\delta$ values for the $71$ AIMs used in the BGA test}$ 

AF-	AF-	AF-	CT-	CT-			
CT				AI	EA-AI	AIM	SEQ ID NO:
13	5 0.214					958	31
	0.207		0.54	0.11	0.44	960	65
0.564	0.549 0.364	0.53	0.01	ი ი4	0.02	961	52
0.286	0.364	0.08	0.08	0.36	0.44	963	39
0.84	0.364 0.286 0.464	0.36	0.56	0.48	0.08		70
0:327	0.464	0.01	0.14	0,33	0.47	966	37
_i .0.321	0.097	0.49	0.42	0.17	0.47	969	63
0.163	0.171	0.06	0.01	0.11			40
0.33	0.7	0.57	0.37	0.24	0.13	971	64
0.497	0.186	0.16	0.31	0.34	0.03	972	1 .
0.339	0.779	0.62	0.44	0.28	0.16 0.13	973	22
0.372	0.725	0.85	0.35	0.48	0.13	977	57
0.041	0.122	0.18	0.08	0.14	0.06	978	3
0.549	0.268	0.19	0.28	0.36	0.07	979	34
0.606	5:0.671	0.52	0.07	0.08	0.15	980	21
0.102	0.771	0.61	0.67	0.51	0.16	993	54
0.623	3 ⁶ 0.071	0.09	0.55	0.53	0.06 0.07 0.15 0.16 0.02	1000	18
0.037	V.1	0.14	ี บ.บอ	0.1	0.04	1012	24
0.63	0.265	0.06	0.37	0.57	0.2	1022	20
0.674	0.025	0.14	0.65	0.82	0.17	1029	13
0.523	0.714	0.87	0.19	0.35	0.16	1033	41
0.634	0,295	0.6	0.34	0.03	0.31	1034	46
0.172	0.539	0.63	0.37	0.46	0.09	1035	25
4E-04	1 0.711	0.17	0.71	0.17	0.31 0.09 0.54 0.32 0.16	1040	29
0.65	0.031	0.29	0.68	0.36	0.32	1041	5
0.314	0.535	0.38	0.22	0.06	0.16	1043	35
$\pm 0.334$	1-0.807	0.52	0.47	0.19	0.29	1044	16
0.343	0.686	0.55	0.34	0.21	0.13	1047	61
0.168	0.537	0.08	0.37	0.08	0.45	1048	19
0.686	5 0.8	0.96	0.11	0.27	0.16	1049	26
0.606	9.0.114	- 0.25	0.49	0.35	0.14	1050	48
0.348	0.586	0.58	0.24	0.23	0.01	1051	42
0.263	0.171	0.26	0.09	0.52	0.01 0.43 0.27	1053	62
						1055	67
0.36	0.742	0.76	0.38	0.4	0.01	1056	44
0.153	0.022	0.03	0.18	0.18		1057	
	0.043				0	1058	59

AF-	AF-	AF-	CT-	CT-			
CT		AI	EA	AI	EA-AI		SEQ ID NO:
0.392	0.725	0.39	0.33			1060	27
	0.829			0.14	0.24		23
0.456	0.193	0.78	0.26		0.58	1064	10
0.157	0.157	0.16	0		0	1066	7
0.043	0.043	0.04	0	0	0	1068	11
	0.013			0.44	0.1	1071	53
	0.071			0	0	1073	8
0.704	0.809	0.81	0.1			1075	69
0.464	0,535	0.34	0.07	0.13	0.2	1076	71
0:348	0.728	0.74	0.38	0.39	0.01	1077	
0.261	<i>*</i> 0.736	0.61	0.47	0.35	0.13		
$\pm 0.343$	0.536	0.41	0.19	0.06	0.13	1082	36
0.454	0.821	0.87	0'.37	0.42	0.05	1083	
0.036	0.45	0.17	0.49	0.21	0.28	1084	51
0.74	$\frac{0.117}{}$	0.35	0.62	0.39	0.23	1087	28
0.985			0.01			1111	30
0.851	0.252					976	68
0.495	0.093	0.15	0.59	0.64	0.06	1113	4
0.133	0.2	0,67	0.07	0.54	0.47	1116	50
0.548					0.09		
0.567	0.65	0.64	0.08	0.07	0.01	1120	60
0.368	0.379	0.47	0.75	0.84	0.09	1121	33
0.575	0,664	0.59	0.09	0.02	0.07	1122	58
0.177	0.086	0.3	0.26	0.48	0.21	1124	17
0.416	0.089	0.02	0.33	0.39	0.07	1128	2
0.108	0.449	0.51	0.56	0.62	0.07	1036	9
0.149	0.1	0.1	0.25	0.25	0	1130	55
0.473	0.634	0.61	0.16	0.14	0.02	1136	38
0.892	0.667	0.22	0.23	0.68	0.45	1137	56
0.076	0.279	0.69	0.2	0.61	0.41	1138	66
0.612	.: 0.66	0.62	0.05	. 0.01	0.04	1139	32
0.435	0.6	0.29	0.17	0.14	0.31	1140	
0.487	0.564	0.61	0.08			1141	49
0.155	0.571	0.31	0.73	0.46	0.26	1146	15

54 50 50 45 41 24

AF- sub-Saharan African, CT – Indo European, EA – East Asian, AI – Native American. The AIM unique identifier is shown (AIM) as well as the GENBANK accession number provided by the authors upon submission of the SNP sequence to the NCBI:dbSNP database. The number of AIMs with  $\delta$ >0.40 for each pair-wise comparison is shown at the bottom of the list.

[0233] The 71 AIMs used in the exemplified panel were spread throughout 21 of the 23 autosomal chromosomes (Figure 8), with the average chromosome containing 3 AIMs (see Table 6). Each had alleles in Hardy-Weinberg equilibrium, both overall with all four BGA groups considered together, and within each BGA group, and none were found to be in linkage disequilibrium with one another. The software program used individual genotypes for these AIMs with a maximum likelihood algorithm (see Example 6, Table 12; see, also, Example 1). The use of the 71 markers with this algorithm provides another example of the "BGA test".

[0234] The BGA test was used to calculate BGA admixture proportions for the parental Native American, African and Indo European samples used in the construction of the test. After calculating the admixture proportions for each sample, they were plotted in a triangle plot to allow for the relative proportions of a 3-way mixture to be represented in two dimensions. Because these were the same samples that comprised the parental groups and from which the population allele frequencies were derived, they were expected to exhibit relatively homogeneous BGA (i.e., of low admixture) and, in fact, the sub-Saharan Africans, Native Americans and European parental samples all registered with relatively homogeneous BGA (i.e., they plotted towards the appropriate vertices of a BGA triangle).

[0235] The BGA test was next used to determine BGA proportions for 1,186 individuals of self-reported race (43 African Americans, 1,120 Caucasians, and 23 Hispanics). 306 of the individuals (26%) showed homogeneous BGA (100% for any one group). 101 of the 1,186 (8.5%) harbored >5% BGA affiliation for three groups, indicating that, for these individuals, a modification of the software to perform 4-group calculations might be more appropriate, and the vast majority of the samples were characterized by 2-way admixture. Significantly greater European admixture was identified in African Americans relative to that of Nigerians (visualized as a dispersal of points away from the sub-Saharan African vertex). In contrast, the IndoEuropean samples plotted as neatly in the IndoEuropean vertex as had the parental samples, though low levels of Native American or East Asian admixture were not uncommon roughly two-thirds of subjects harbored detectible, though generally low levels of such

admixture. The Hispanic subjects plotted in an even distribution along the Native American/IndoEuropean axis, consistent with the knowledge Hispanics arose from the blending of colonial Europeans and resident Native Americans about 500 years ago.

[0236] In order to determine whether, and to what extent, the major proportions obtained with the BGA test corroborated with self reported race, BGA admixture proportions were calculated for 2,048 individuals of self-reported race, and blindly (in a computational sense) compared the majority BGA determined from the test against each individual's self-reported majority race. A very strong concordance was observed between the major BGA group determined with the test and the self-reported majority race (Table 7). Using the test, 1252/1252 self-described European-Americans (U.S. born Caucasians) registered with majority IndoEuropean BGA. 191 of 201 self-described African Americans showed majority sub-Saharan BGA, with the remaining 11 showing sub-Saharan BGA as the minor affiliation with IndoEuropean as the majority affiliation. Hispanics showed roughly equal distribution in majority BGA between IndoEuropean and Native American, consistent with the results observed in the triangle plot and the anthropological history of this group.

TABLE 7
Comparison of Majority BioGeographical Ancestry with Self-Reported Race

Self-Reported Race	European	African	Native American	East Asian
European-American (US white)	1252	0	0	0
Africans from Nigeria	0	217	0	0
African-American (US black)	11	190	0	0
Natives ("Nativos") from Guerrero, Mexico	1*	0	72	0
Hispanics born in US, of Mexican heritage	101"	0	90	0
Chinese	0	0	0	10
Japanese	0	0_	0	10
Other general Asian from US	1***	0	0	33
South Asians from India	36	0	0	0.

^{*}Minor proportion was Native American

^{**}Significant Native American as second type.

^{***}Minor proportion was East Asian

113

[0237] Even when unexpected results were obtained, such as the finding that one individual from Southern Mexico was majority European, the results were more or less concordant in that the expected affiliation was the minor affiliation, rather than being absent altogether, and the major affiliation (Indo European) made sense in light of the history of the region (colonized by the Spanish hundreds of years ago). In this particular case, the level of Native American ancestry was only slightly less than 50%. Not one gross error was observed where, for example, a self-reported European-American was classified as majority East Asian. Though the 11 self described African-Americans of majority IndoEuropean BGA would appear to be a gross error, the results were similarly concordant — each of these African American samples exhibited roughly equal IndoEuropean/African proportions, suggesting admixture, consistent with the historical tapestry of the region from which the samples were taken, rather than test error.

To conduct a truly blind test (as opposed to a test that is blind in a computational [0238] sense), the San Diego Police Department Crime Lab (SDPD) and the National Center for Forensic Science at the University of Central Florida (UCF) each submitted ten buccal swabs of numerically encoded identity for BGA tests. The BGA test was performed and the results returned to SDPD and UCF, each of which independently evaluated their results and revealed the self-reported population affiliation of the sample. The major percentages determined from the BGA admixture proportions test were not inconsistent with the self-reported population affiliations (see Table 8). Several of the samples were from individuals affiliated with groups that are logically considered to be admixed - e.g., Filipinos (SDPD2, SDPD3, Table 8), African American or Carribean (SDPD5, SDPD6, UCF7, UCF8, Table 8), Mexican Americans (Hispanic, SDPD8, SDPD10, Table 8) and Puerto Rican (UCF6, Table 8); significant admixture was detected for each of these samples. Moreover, the type of admixture detected was reasonable with respect to the anthropological history of the affiliated population. For example, people of sub-Saharan African, Native American and IndoEuropean descent live in Puerto Rico, whereas East Asians are relatively rare, and the test results for the Puerto Rican individual tested showed IndoEuropean and Native American, not East Asian admixture.

[0239] In addition to determining the MLE of admixture, the software program was designed to survey the probability space, and define that space within which the likelihood of proportional affiliation was 2-fold, 5-fold, and 10-fold less likely to be the correct answer than the MLE (confidence contours, which are plotted on the triangle plot as rings around the MLE). One way to test the accuracy of the maximum likelihood algorithm for determining MLE and confidence contours was to observe whether and how these values change when certain of the AIM markers are eliminated from the analysis by replacing the genotypes for each with "failure" readings. For example, if for a given sample genotype, all of the genotypes for markers of high δ value for the African/East Asian distinction were replaced with "failure" or "no data", an accurate test would be expected to show warped confidence contours only in this dimension of the triangle plot. Accordingly, the BGA test was used to plot one sample of majority East Asian BGA with its associated confidence intervals (Figure 9A), then all 24 of the markers in the test were eliminated with informative  $\delta$  for Native American vs. East Asian BGA, and the MLE and confidence estimates were recalculated with the remaining AIMs (Figure 9B). Upon recalculation with the missing AIMs, the confidence rings were dramatically skewed from the East Asian towards the direction of the Native American BGA vertex (Figure 9B), indicating, as expected, that the lack of AIMS with good δ values for East Asian/Native American distinction produced an estimate for which the confidence along the East Asian/Native American axis was not high. Presumably because the sample typed as of majority East Asian, and AIMs for the distinction of East Asian/IndoEuropean and East Asian/African affiliation were left unmolested, the MLE shift itself was minimal; most of the uncertainty along the East Asian/Native American axis was apparent in the shift of the contours. Similar experiments with other samples and AIMs produced similar results.

115

TABLE 8

Blind Challenge of BGA Test by the San Diego Police Department and the Center for Forensic Science at the University of Central Florida

ID#		<u>%</u>	Admix Ratio	<u>%</u>		%	Self Reported Race
SDPD1	EUROPEAN	98	EAST-ASIAN	0	NATIVE-AMERICAN	4	European-American
SDPD2	EAST-ASIAN	:53	EUROPEAN	47	NATIVE-AMERICAN	0	Filipino
SDPD3	EAST-ASIAN	61	NATIVE-AMERICAN	28	EUROPEAN	11	Filipino
SDPD4	EUROPEAN	100	EAST-ASIAN	0	NATIVE-AMERICAN	0	European-American
SDPD5	AFRICAN	69	EUROPEAN	31	NATIVE-AMERICAN	0:	Caribbean
SDPD6	AFRICAN	67	EUROPEAN	33	EAST-ASIAN	0	African American
SDPD7	EUROPEAN	99	EAST-ASIAN		NATIVE-AMERICAN	0	European-American
SDPD8	NATIVE-AMERICAN	57	EUROPEAN	43	EAST-ASIAN	0	Mexican American
SDPD9	EAST-ASIAN	86	NATIVE-AMERICAN	0	EUROPEAN	14	Filipino
SDPD10	NATIVE-AMERICAN	36	EAST-ASIAN	28	EUROPEAN	36	Mexican American
UCF1	EUROPEAN	90:	EAST-ASIAN	0	NATIVE-AMERICAN	10	Ukraine/Italy
UCF2	EAST-ASIAN	98,	NATIVE-AMERICAN	2	EUROPEAN	0:	Chinese
UCF3	EUROPEAN	.:88	EAST-ASIAN	3	NATIVE-AMERICAN	9	Ukraine
UCF4	EUROPEAN	100	EAST-ASIAN	O	NATIVE-AMERICAN	0	European-American
UCF5	EUROPEAN	87	EAST-ASIAN	0;	NATIVE-AMERICAN	13.	Greek
UCF6	EUROPEAN	62"	NATIVE-AMERICAN	38	EAST-ASIAN	0	Puerto Rican
UCF7	AFRICAN	83	EUROPEAN	17	EAST-ASIAN	0	African American
UCF8	EUROPEAN	69	AFRICAN	31	NATIVE-AMERICAN	0	Jamaican
UCF9	EUROPEAN	∵84	EAST-ASIAN	12	NATIVE-AMERICAN	4:	Finland

2 NATIVE-AMERICAN 0

[0240] In order to determine the reproducibility and consistency of the BGA admixture proportion determinations, five samples were genotyped and analyzed on separate occasions. One sample was chosen of self-reported majority affiliation with the European American, African American, Hispanic and Asian groups, and a fifth sample was chosen from the parental Native American group. With the exception of failed loci, the genotypes at each marker in each individual were 100% consistent between runs, indicating that the AIMs genotype reliably. A 1-3% variation in BGA admixture proportions was observed from run to run; simulation studies showed that the variation was attributable to these genotyping failures. This result indicates that the failed loci did not pose a significant barrier to reproducibility of the BGA admixture determination for an individual, either in terms of majority BGA or admixture levels. From these simulations, it was determined that the BGA

UCF10

**EUROPEAN** 

98

**EAST-ASIAN** 

116

test tolerates about ten locus failures if the samples are of least binary admixture, and for samples of no admixture, larger numbers of failed loci are tolerable (i.e., change in admixture percentages were less than 5%).

In order to determine if the BGA admixture proportions determined with the test [0241] were sensible given the rules of family inheritance, proportions for three generations from several family pedigrees were calculated. A typical result is shown in Figure 10, which depicts the ratios obtained for a family of confirmed paternity (using STR tests) with substantial European/Native American admixture. The first generation individuals were self-reported European-Americans that were determined with the BGA test to harbor significant Native American admixture, which was passed to their son and daughter in different proportions that were not inconsistent with the law of independent assortment. The son's spouse was a Hispanic native of Mexico, and was determined to be of 26% European/74% Native American admixture. Each of their offspring harbor roughly intermediate levels of Native American and IndoEuropean admixture between their parents, again not inconsistent with the law of independent assortment. One of the sons typed as of a small percentage of East Asian ancestry, but the level (4%) was close to the reliable limit established as discussed above (about 3%). Other pedigrees tested (n=8) showed similarly concordant results, indicating that, in terms of majority BGA and admixture levels, the BGA test results were sensible within the context of family pedigrees.

[0242] Because the BGA test relies on parental population allele frequencies, the extent to which the admixture proportions produced by the test were influenced by parental allele sampling bias was investigated. A pair of populations was selected, as were the AIMs relevant for resolving affiliation between these two populations (Table 6 - selected those with the highest  $\delta$  values for this pair of groups), then the allele frequency was adjusted for each of these AIMs in one of the groups such that the  $\delta$  value for the AIM was reduced by 20% (with respect to these two groups). In effect, the power of the test was intentionally degraded by 20% for the resolution of affiliation between a specific pair of groups; this degraded test is referred to as the Ancestry 2.1EA/EU BGA test, where the EA/EU refers to degradation in the distinction between the East Asian (EA) and IndoEuropean (EU) groups. 31 samples

117

were randomly selected, the genotypes were run against the Ancestry 2.1EA/EU BGA test in exactly the same manner as for the original (ANCESTRYbyDNATM 2.0) test, and the results were compared with those obtained with the ANCESTRYbyDNATM 2.0 test.

[0243] If the results from the ANCESTRYbyDNATM 2.0 test were highly sensitive to parental allele frequency error caused by sampling bias, which might be expected to be on the order of a few percent at the most, the 20% change introduced for the Ancestry 2.1EA/EU AIMs should result in admixture proportions substantially different from those of ANCESTRYbyDNATM 2.0. Since the number of IndoEuropean/East Asian mixes observed was significantly greater than other types of mixes, such as African/Asian mixes or Native American/African mixes, the European/East Asian pair was selected for the first test – Ancestry 2.1EA/EU - and the BGA group pair for which the number of AIMs and cumulative δ value is the lowest is Native American/East Asian, the δ values for this pair was altered in the second test – Ancestry 2.1NA/EA. The average change observed in admixture proportions between ANCESTRYbyDNATM 2.0 and Ancestry 2.1EA/EU was 1.4% (Standard Deviation 2.44%). For the Native American/East Asian pair, the average change between ANCESTRYbyDNATM 2.0 and Ancestry 2.1NA/EA was 1% (Standard Deviation 2.3%).

[0244] Socio-cultural or self-held notions of race are not likely to be as tightly linked to human biology as BGA, which is the heritable component of race. Because it is subjective, imprecise and sometimes inaccurate, the use of self-identified race for the inference of BGA, as is currently practiced, obscures how and why human biology is related to human anthropology. Furthermore, the rigid binning of patients into prefabricated racial groups is a practice of generalization that is wholly unsatisfying because many individuals can trace their origins to multiple populations through the process of admixture. A repeatable, testable anthropological approach to define BGA can provide a means to draw connections between BGA and heritable diseases, whether through straight correlation and/or better study designs, or more subtle means such as through gene mapping methods that rely on the admixture process, such as MALD.

118

[0245] As disclosed herein, a 71 marker test allowed a determination of BGA proportions and their confidence intervals. The test enabled a determination of the relative proportionality of BGA within individuals, thus distinguishing the BGA test from other tests previously used for inferring ancestry from DNA. In terms of the majority BGA affiliation, more than 2200 tests were performed and no result was obtained that was inconsistent with self-held notions of race. Previous tests of ancestry have been accurate only to the upper 90% range (Shriver et al., supra, 1997; see, also, Frudakis et al., supra, 2003, which is incorporated herein by reference). The enhanced performance observed with the BGA test can be because CODIS and other STRs that have been commonly used for inferring ancestry from DNA were not selected for their δ values, but were selected for their polymorphic complexity in the world population. For the BGA test disclosed herein, the entire genome was systematically scanned and the best AIMs for this purpose were selected. In addition, most efforts to infer ancestry from DNA using STRs or Alu sequences have attempted to categorize or bin samples into single "racial" groups. For individuals of extensive admixture, such as a 50/50 mix, such a method would seem to produce a "wrong" answer as many times as a "right" answer. In contrast, with the BGA test, ancestry is determined in terms of proportional affiliation, thus ameliorating this problem.

[0246] The BGA test is distinguishable from other tests in that it employs SNPs that cover most of the chromosomes. Pan-chromosomal coverage using the BGA test provides a substantial advantage over tests using CODIS STRs, which only cover a fraction of the chromosomes. In addition, the BGA method appears to be the first that quantifies the confidence limits for its answers. The BGA test as exemplified is largely heuristic, and divides the world into four main anthropological groups that fall largely along continental lines. Though the geographical divisions are respectful of the anthropological history of human migrations, the use of four groups is indeed a simplification of a very complex situation and can be considered to be arbitrary. Further, the problem of determining proportional affiliation has been simplified by calculating the most likely 3-way (rather than 4-way) combination, because individuals of 4-dimensional BGA are thought to be rare, and because it is more convenient in a computational sense. However, while more complex tests may be able to capture more of the extant detail of anthropological history, even a crude 4

population test provides data of meaningful and historical content, provided the results are interpreted strictly with respect to these divisions and to the parental samples used in the construction of the test.

[0247] The choice of specific parental groups and of dividing the world into certain groups simply provides a point in coalescent time by which to scale the inferences provided by the test. In fact, the difference between the answer provided by a test based on 4 world population groups and a more complex test based on 25 would be one of anthropological time scale, not "accuracy". For example, most Hispanics born in the United States that have been tested and most individuals claiming American Indian heritage type that were tested revealed minor Native American admixture on an IndoEuropean background. However, unlike the case for Hispanics, some of the individuals claiming American Indian heritage were typed as of minor East Asian admixture instead of Native American admixture. Because the founders for Native Americans migrated from East Asia, possibly in different waves at different times in history, the genetic distance between Native Americans and East Asians is lower than between Native Americans and sub-Saharan Africans or IndoEuropeans (Cavalli-Sforza and Cavalli-Sforza, supra, 1995). Among pre-colonial North Americans, proportional affiliation to East Asian or Native Americans would be expected to be different for individuals whose ancestors were part of the first waves than from those whose ancestors were part of later waves.

[0248] The parental samples for Native Americans used in the present study were derived from southern Mexico, and the AIM allele frequencies established for Native Americans obtained from this sample might be expected to be more representative of the ancestors from earlier waves of migration across the Strait. Native Americans from Latin America and South America would likely be more closely affiliated with early wave ancestors than, e.g., those from North America such as the Aleut Indians (and others), which would probably be more closely affiliated with ancestors from later waves across the Strait. The East Asian affiliation for those individuals claiming affiliation with American Indians may be a byproduct of the choice of Southern Mexico Nativos as the parental source, and the use of only a 4 group anthropological scheme, but nonetheless, the answer is not a "wrong" answer in a

120

scientific sense. Rather, it reports affiliations with respect to a coalescent time scale defined by the source of the parental samples and the anthropologically meaningful way in which the world was divided for this study.

[0249] A different test, with more markers to resolve affiliation within the North American groups from each other and from East Asians, would operate on a different coalescent time scale and likely classify these individuals as of minor "American Indian" or "North American Native American" admixture. Nonetheless, the fact that the exemplified BGA test revealed that a significant number of individuals of "Native American" ancestry show more affiliation with East Asians than Native Americans, as defined by the metrics built into the test, may be yet another example that social or human history based notions of population affiliation are semantic, subjective and not always accurate in a biologically meaningful way. Even though Aleuts may appear to resemble East Asians as much as, or even more than, most Native Americans in terms of physical features, and even though they are indigenous to a geographical locale as proximal to East Asia as to temperate North America, they are considered by most to be North American Indians and by extension, Native Americans because their home lies east of the Bering Strait. Similar examples have been observed for certain other population groups, as discussed below, illustrating the disconnect between the measurement of population affiliation using genetic markers and that from geographical and social borders man has devised to ascribe racial identity.

[0250] With deference to the qualifications of the BGA test, it is interesting to compile the results in order to extract some meaningful anthropological and/or sociological knowledge. Using the BGA test, 11 of 201 African Americans tested showed major IndoEuropean and minor African BGA, and Puerto Ricans of majority African descent almost always describe themselves as Hispanic, again pointing towards the deficiency with current notions of race as a dichotomous entity based on man-made constructs. As was observed in the present study, Risch et al. (supra, 2002) and Rosenberg et al. (supra, 2002) have shown that, when tested against methods of reporting BGA that rely on genome markers, majority population affiliation is quite accurately reported on questionnaires. The present testing of over 2,000 individuals indicates that the majority BGA affiliation can be accurately predicted from the

self-reported race, that discordance between the two is not a very significant event, and that determination of majority ancestry affiliation is not the main problem with current self-reporting methods. However, perhaps the most surprising result was the extent of admixture for each population tested. When individuals claimed admixture, it was almost always confirmed with the BGA test; each case of expected sub-Saharan African and East Asian admixture was confirmed with the BGA test, and every Hispanic of Mexican descent registered with either major or minor Native American admixture, as expected.

Slightly more than two-thirds of all "Caucasians" tested exhibited minor East [0251] Asian or Native American admixture, and virtually none of these individuals reported any significant pedigree admixture on his or her questionnaire. Some of this admixture appears to be a function of ethnicity. Not only did individuals of relatively homogeneous self-reported Northern and Eastern European heritage more commonly show East Asian BGA, but Rosenberg et al. (supra, 2002) showed there to be significant structure within the "European-American" or "European" population that support this observation; specifically they showed that Russians commonly exhibit minor East Asian heritage. There are multiple times in history where such East-Asian/IndoEuropean admixture may have taken root, including, for example, the Mongolian invasions of Europe, and the Caucasoid expansion to Scandinavia, whose Lapp inhabitants were derived from Northern Asians, exhibit Mongolian features, and share a common history and culture with East Asians (Cavalli-Sforza and Cavalli-Sforza, supra, 1995). Some of the Native American admixture observed is concordant with history; that Filipinos exhibit extensive Native American admixture, for example, is not entirely surprising in view of the Spanish having conquered most of Latin America and exported Native American slaves to these islands, which were a Spanish territory until recently. The extent of Native American admixture observed for some Filipinos was quite high, perhaps reflecting that Native American admixture is relatively common in this part of the world, and that the pedigree for many Filipinos is dominated by large numbers of individuals of relatively low Native American admixture, rather than recent admixture with individuals of highly polarized BGA proportions. The Native American admixture commonly observed in "Caucasians" likely came from a blending of European and Native American peoples in North America.

122

[0252] In most cases of systematic admixture, such as between Scandinavian/Russian IndoEuropeans and East Asians, sub-Saharan African/IndoEuropean in the U.S., Native American/East Asian mixture in the Philippines or IndoEuropean/Native American in the U.S., the geographical proximity and/or historical mixture of the relevant groups is well established by history. For instance, an African/East Asian mixture was rarely observed in these studies, and history knows few examples of times when these two populations lived in close proximity to or mixed with one another. The type of admixture observed was also interesting when compared to self-held notions of "race". For example, African Americans were more admixed with IndoEuropean Ancestry than were Caucasians with African ancestry, highlighting a difference in how Caucasians and Africans view their heritage, and invoking recollections of the "one drop rule". Given the extent of admixture observed, it is likely that hidden or cryptic BGA structure arising from the process of admixture (a process that is not completely documented and quantifiable from our anthropological literature, as it is based on human constructions) is of potential concern for creating gross (or finer) structure differences between groups of study samples. Such a difference in structure would be expected to reduce the efficacy and power of large population based study designs.

[0253] Given that the majority ancestry was not inconsistent with self-held notions of affiliation for over 2,200 blind test subjects of polarized (low admixture) and self-reported race, the question arises as to how to assure the accuracy of minor admixture proportions such as these, and how such accuracy can be measured given that there is no ultimate arbiter of BGA in existence (except, possibly, genealogical information, see below). Several experiments were performed that address this question and, when considered together, indicate that the minor proportions are accurately determined. First, the minor admixture percentages transmit along family pedigrees in a manner consistent with the genetic law of independent assortment. By definition, a large, unbiased error would make it such that minor proportions were incoherent in the context of family pedigrees, i.e., the results would not be possible given the law of independent assortment, assuming the proportions of the parents to be correct.

123

[0254] Second, the minor admixture proportions are consistent, on average, with self-held notions of admixture. If there was a large, systematic and unbiased error in the estimation of BGA affiliation, this error would impact the integrity of the minor proportion percentages more than that of the major proportion percentages, since most individuals are of relatively polarized BGA affiliation, but the correlation of minor admixture proportions/presence with self-held notions of admixture would probably be very weak, if not imperceptible altogether. This was not observed to be the case. For example, if the error rate was as high as 20%, as many individuals reporting minority Hispanic ancestry would show minor sub-Saharan African and East Asian ancestry as Native American; the present results clearly demonstrate this is not the case.

[0255] Third, of about 2,200 samples blindly tested from North America, an individual with majority East Asian with substantial (>10%) sub-Saharan African admixture, or vice versa, has never been observed. This non-observation is relevant because such individuals are exceedingly rare in North America, from which the validation sample was derived. If there was a large, unbiased and systematic error rate, East Asian/African mixes would be observed as frequently as European-Asian mixes, which were frequently observed.

[0256] Fourth, a highly significant correlation was observed between minority IndoEuropean admixture and skin melanin content in African Americans (see Example 1). If there was a large, unbiased error rate, such a correlation would not likely be obtained.

[0257] Fifth, when the true allele frequencies for AIMs relevant for the resolution of affiliation between two groups is adjusted, such that the parental  $\delta$  value was decreased by 20% for each relevant AIM for a given pair of groups, the overall power of the BGA test was degraded with respect to resolving affiliation proportions between these two groups, but essentially the same results were produced, both in terms of major affiliation and, more importantly, of minor admixture estimations. Parental sampling bias can cause inaccuracies in parental allele frequency estimation and  $\delta$  values in this manner, though certainly less than 20% given that the parental sample for the present studies was comprised of about 100 individuals. Further, the error in allele frequency estimation would have to be obtained

in the same direction for most of the AIMs relevant for resolving affiliation between a pair of groups for such an error to exist. Nonetheless, this result indicates that, even if such an error existed, the performance of the BGA test would be relatively unaffected. In other words, this experiment demonstrated that the BGA test is relatively robust in the face of parental sampling bias and allele frequency estimation.

[0258] Sixth, the distortion of confidence contours along only the axis corresponding to that for which genotyping failures were relevant shows a healthy disconnect among the interdigitating components (subsets of AIMs) of the test. In other words, if samples are fit to several templates for the determination of proportional fit, the elements of these templates should be independent for the test to be valid, and this was the result obtained.

It is difficult to imagine how significant systematic and unbiased error could exist [0259] in light of the above six observations. While each observation, on its own, may not prove the point of accuracy, taken together the results provide ample evidence that there is little or no systematic and unbiased error in the disclosed BGA test. It nevertheless could be argued that the error in the test is not random, but biased in a linear manner. However, the fifth observation (above) argues against this possibility. For example, the finding of minor East Asian admixture on an IndoEuropean background was more frequent than had been expected, and such a result could occur if there was the "right" amount of allele frequency estimation error in the "right" number of markers in the "right" direction (an unlikely, but not impossible, situation). However, because the AIM markers used for these studies are not mutually exclusive to particular group pairs, such an error would manifest itself in the resolution of affiliation for many pairs, not just one, and the first four observations (above) indicate that this is not the case. Also, such an error would seem to require parental sampling from a highly inbred group. While great care was taken to avoid such sampling, every element of population structure was not controlled for because there exists no test to do so a priori. As such, a simulation was performed to estimate the contribution of linear error to the test results (fifth observation, above). The fact that admixture results were relatively impervious to substantial (20%) reductions in δ value demonstrates that the quantity and quality of the AIMs used in the present studies were of adequate power such that reasonable

125

levels of sampling error that may have been expected did not have a significant detrimental impact on the quality of the results. In terms of marker quality, this result is plausible because, given the selection process for AIMs, some of the best markers in the genome were used to determine BGA affiliation. In terms of marker quantity, these results are consistent with the observations that the results from the 71 marker test, as disclosed herein, were very similar to that produced from an earlier 30 marker test (Shriver et al., *supra*, 2003). Thus, reducing the number of markers, while retaining the average marker quality, did not impair the test. Furthermore, in another study, reducing the quality of the makers, but not the quantity, produced the same results. Overall, these observations indicate that the BGA proportions produced by the test are accurate with respect to the confidence intervals presented, and that the test performs in a robust manner.

The present results demonstrate that BGA admixture is more common than [0260] previously believed. If true, then it should be asked whether finer levels of BGA admixture are linked to human biology, e.g., drug responsiveness or disease predisposition. Such "cryptic" structure can only be determined using a molecular test because, unlike crude population structure. a questionnaire cannot be used. Finer levels of population structure beyond the more crude continental levels measured using questionnaires are recognized. For example, significant anecdotal evidence suggests that redheads require a 20% larger dose of many common anesthetics, and exhibit a tendency towards hypertension and bleeding while under anesthesia (Cohen, The Scientist 16:10, 2002). These complex physiological responses would be difficult to explain based on melanocortin-1 (MC1R) variants previously linked to some of the variation in red hair color (Robbins et al., Cell 72:827-834, 1993; Smith et al., J. Invest. Dermatol. 111:119-122, 1998; Flanagan et al., Hum. Molec. Genet. 9:2531-2537, 2000). There are likely specific gene variants responsible for these clinical phenotypes and, if these variants correlate with elements of population substructure or microstructure, as they appear to do, any study attempting to identify linkages or LD between markers and the relevant phenotypically active loci will be challenged from the outset at the study design step. The exemplified BGA test can be extended using AIMs in addition to those disclosed herein to quantify the elements of population structure relevant for this particular problem because a precision and objectivity greater than that provided by self-reporting of socio-cultural race

126

will be needed to identify the elements of structure that can interfere with the design of genetics experiments.

## **EXAMPLE 3**

# APPLICATION OF THE BGA TEST TO GENEALOGY

[0261] This Example demonstrates that BGA admixture estimates can be integrated with genealogical information obtained using traditional genealogical research methods.

[0262] Genealogists collect data that largely is relevant in a geopolitical context (e.g., data relating to which countries a person's ancestors are from, what their religions were, and their last names) rather than in an anthropological context (e.g., what type of population admixture characterizes the person's family tree). There are two main sources for obtaining minority admixture in one's results: 1) recent exogamous admixture events; and 2) ancient affiliations with ethnic groups that are characterized by systematic admixture.

[0263] The results of an exogamous event is determined in a recent genealogical time (e.g., the last 250 years). For example, as shown in Figure 11, a Chinese great grandparent in an otherwise homogeneous IndoEuropean family tree would produce a grandchild of IndoEuropean/East Asian admixture. The individuals that are 100% East Asian (Chinese) are shown with shading (Figure 11), and the admixture results for the male (square) at the bottom of the pedigree (short arrow) are of interest. A person with a single 100% East Asian great grandparent and seven 100% IndoEuropean great grandparents would be expected to have 12.5% East Asian admixture. By the law of genetic assortment, the expected level is actually a range around 12.5%, with values several percent above and below possible. The grandparent indicated by the long arrow is about a 50%/50% East Asian/IndoEuropean mix and her daughter, the subject's mother, is expected to be a 25%/75% East Asian/IndoEuropean mix (Figure 11).

[0264] Ancient affiliations (i.e., considered with respect to an anthropological time frame) have been preserved in modern times by endogamous, relatively geographically isolated, close knit community structure (i.e., ethnicity). For example, modern day demography is shaped by not only the migrations our ancestors made to establish new populations, but also

by admixture between these populations throughout the world. The map below shows these migrations as measured from Y chromosome sequences, which occurred over many tens of thousands of years.

[0265] Admixture between the groups, after each had developed as distinct groups, has occurred many times throughout ancient and even more recent history, and was represented by arrows on the map representing migration patterns. For example, there was extensive East Asian admixture in Russians and Eastern Europeans (Rosenberg et al., supra, 2003), and the extent to which the Mongolian and Hun invasions may have contributed to this admixture over a long time period remain a mystery. There was an even more pronounced East Asian admixture apparent for Native Americans (Rosenberg et al., supra, 2003); arrows were not included on the map for this admixture because far too many would have been required and most of the admixture events are not know. Nonetheless, a person with a fair number of Native Americans or Russians in their family tree could very well exhibit as much East Asian admixture as an individual with a 100% Chinese grandmother and three other 100% IndoEuropean grandparents.

[0266] Although not shown, a time scale was constructed showing the time the most significant migrations occurred, and was correlated to a very, very large family tree. The tree is for a single individual, who resides at the bottom apex of a triangle graph; it is large because it goes back 60,000 years when there are tens of thousands of ancestors for this person. The time scale for the migrations applies to the large family tree as well. The tree was the same as that shown in the pedigree map (Figure 11), only much larger and without the lines connecting the ancestors (spots represented each ancestor, but there were so many that it was not practical to show all of the lines connecting the spots. A pool of spots represented "Russian", which for purposes of this example were assumed to be an ethnicity that arose about 18,000 years ago. Additional spots represented East Asian, and was based on the assumption that the average Russian harbors 10% East Asian admixture. A third set of spots represented the precursors to Russians; these precursors are unknown but, for purposes of this example, were assumed to be Eastern Europeans.

In this example, most of this person's Russian ethnicity came from the left side of [0267] the family tree, which can be assumed to be that part of the tree representative of the subject's father's side of the family. If, as this example indicated, the average Russian harbors 10% East Asian admixture, and half of the person's family tree is predominantly Russian, the person would be expected to harbor 5% East Asian admixture. East Asian admixture is significant for this person even though neither the person's grandmother, grandfather, or any other relative within the past 18,000 years was homogeneous East Asian. The way to visualize this on the family tree is to count all of the "East Asian" spots and divide them by the total number of spots in the tree, to arrive at about 5%. Thus, relatively homogeneous East Asians represented about 5% of the total number of ancestors for this person. Of course, the family tree for some people involves numerous groups that are characterized by small degrees of this type of admixture. Family trees like that exemplified are polarized with certain ethnicities, and it is uncommon to see a tree with an equal distribution of each of the four BGA groups (sub-Saharan African, Native American, IndoEuropean and East Asian) because, until recently, and even now to a certain extent, people have tended to have children with others like themselves. As such, most family trees are not a "mish-mash" of random affiliations, but are highly polarized as exemplified.

[0268] The Pennsylvania Dutch provide another example of admixture due to ancient affiliations, wherein, in this community, it appears that there existed significant East Asian content prior to 1700 in German antecedents. These antecedents established communities that populated the valleys in the upper Rhine River Basin, then, later, more further inland. Since these communities remained relatively isolated, the level of East Asian admixture has remained around the 20% level. Dilution of this level would require external admixture with other IndoEuropean ethnicities such as French or Sardinian, for which East Asian admixture is not detectible. In this case, the German antecedents were of substantial average East Asian admixture, perhaps due to sampling from within a more heterogeneous German population.

[0269] Most genealogists are interested in the type of admixture found in source arrived at according to the methods of exogamous admixture, which gives information about geopolitical affiliation of recent ancestors, rather than anthropological information on distant

ones. This is because there exists little paper data for more distant ancestors compared to recent ones; the further back in time a person goes, the larger number of ancestors exist, making research difficult if not impossible; and the contribution of distant ancestors towards a modern persons genetic constitution is less on average per relative than that of more recent ancestors. As such, genealogists tend to seek information such as that which might be produced due to recent admixture. For example, if a person is trying to prove or disprove a rumor or legend of American Indian ancestry, a 10% Native American admixture result would be very useful if it could be assured that the mechanism for this admixture was due to recent exogamous admixture events; and not to ancient affiliations with ethnic groups that are characterized by systematic admixture. While the BGA test does not allow a distinction between exogamous v. ancient admixture, the results of the test provide an important piece of the genealogical puzzle.

[0270] For some genealogists, depending on the family tree, evidence may strongly suggest that the mechanism of admixture is from recent events. As such, for a person whose family has paper evidence of an American Indian great grandparent, a 10% Native American admixture result according to the BGA test can indicates the event likely arose due to recent admixture. In comparison, for a person of confirmed and homogeneous European ancestry, a 10% Native American admixture indicates the event likely arose due to an ancient admixture.

[0271] Negative results carry different meaning than positive ones for genealogists. For example, if there is circumstantial, but low quality, data suggesting a pure blood African great-grandfather, and the BGA test reveals 100% IndoEuropean, then the rumor would be discounted (taking into account the genetic law of independent assortment, which would make such a result possible, but unlikely if the data was in fact correct). However, if a person's family is suspected to have had a Chinese great grandfather, one cannot prove it from a 20% East Asian admixture result, since it is not possible to distinguish exogamous admixture from ancient admixture.

[0272] It is important for a genealogist to bring other knowledge to bear in order to reconstruct the most likely source for an admixture result. In fact, the BGA admixture data serves as an independent clue for one attempting to reconstruct a family history, and when it

is used with genealogical knowledge, the two combine to form evidence that is more powerful than either on their own. As such, the BGA test provides an ancillary tool that can help fashion a system tailored for the genealogy community by providing BGA admixture results in a manner that places equal emphasis on the anthropological sources, which transmitted ancient or very old (relative to a genealogical time frame, which encompasses the last 250-300 years) admixture to us in modern times, and exogamous admixture due to events in the family tree in the last 200 years.

[0273] A database of several thousand BGA profiles is built from people of various locations throughout the world such that one can query the database with a profile, plus or minus a pre-selected error range. A list of places for which this type of profile has been commonly found can be provided, or, for example, a map of the world that is color-coded can be provided, wherein the colors indicate the likely regional affiliations corresponding to the admixture profile/range. In other words, given a BGA admixture profile, a map can be provided showing the places from which the person's recent ancestors could have been derived. A person with 10% East Asian and 90% Indo European would show high probability of ancestral derivation from China (exogamous admixture) or Russia (more ancient admixture coupled with ethnic homogeneity over the family tree).

[0274] Similarly, the genealogist can provide a map, similarly color-coded, that is derived from paper research, which is based on geopolitical rather than anthropological information. The two maps can then be overlaid, and Bayesian statistical calculations made that combine the information from the BGA test with that of the paper genealogy to provide a most likely estimate of recent family history.

[0275] By way of example, a person with 90% IndoEuropean and 10% East Asian BGA and a paper genealogy of Romanian/British/Spanish ancestors would first query the database with his or her BGA result, and be provided a map where the sources would be shown as possibly from East Asia (due to recent admixture), Russia and Northern/Eastern Europe (both due to a large number of more distant ancestors from relatively isolated and admixed groups). The color-coding would give the probability of derivation from the regions based on the frequency with which the compatible BGA groups are found in each region, and it may be

quite complex depending on the mixture type and the character of our database, which is a function of world-wide sampling. Second, the person would provide (or be provided with) a separate map based on the probable Romanian/British/ Spanish heritage documented from genealogical research, using map drawing tools we could provide. From this map, it would be apparent that the likelihood of recent, homogeneous East Asian ancestors is not high. Third, a program would determine that the most likely origin of the 10% East Asian admixture is from the Romanian ancestors (not the British or Spanish, and not due to a Chinese grandparent, for example).

[0276] This type of presentation allow a person to learn the most likely source of an unexpected admixture result, using prior knowledge obtained through other means such as genealogical research. This is valuable to a genealogist seeking to explain the derivation of a genetic constitution. Without this ancillary tool, a person with 90% IndoEuropean and 10% East Asian admixture would have no ready means to determine whether the test suggested a recent Chinese or Japanese grandparent/great grand parent or a certain ethnic affiliation for which minor East Asian admixture is commonly found.

#### EXAMPLE 4

# ASSOCIATION OF IRIS PIGMENTATION AND BIOGEOGRAPHICAL ANCESTRY

[0277] This Example demonstrates that cryptic population structure as determined using AIMs allows an inference as to a complex genetic trait such as iris color.

[0278] In order to determine whether and how common polymorphisms are associated with natural distributions of iris colors, 851 individuals of mainly European descent were surveyed at 335 SNP loci in 13 pigmentation genes and 419 other SNPs distributed throughout the genome and known or though to be informative for certain elements of population structure. Numerous SNPs, haplotypes and diplotypes (diploid pairs of haplotypes) were identified within the OCA2, MYO5A. TYRP1, AIM, DCT and TYR genes and the CYP1A2-15q22-ter, CYP1B1-2p21, CYP2C8-10q23, CYP2C9-10q24 and MAOA-Xp11.4 regions as significantly associated with iris colors. Half of the associated SNPs were located on chromosome 15, which corresponds with results others have previously obtained

from linkage analysis. Five additional genes (ASIP, MC1R, POMC, and SILV) and one additional region (GSTT2-22q11.23) with haplotype and/or diplotypes were identified, but not individual SNP alleles associated with iris colors (see, also, Intl. Publ. No. WO 02/097047). For most of the genes, multilocus gene-wise genotype sequences were more strongly associated with iris colors than haplotypes or SNP alleles. Diplotypes for these genes explain 15% of iris color variation. These results provide a comprehensive candidate gene study for variable iris pigmentation, and constitute a classification model useful for the inference of iris color from DNA. The results further demonstrate that cryptic population structure can serve as a leverage tool for complex trait gene mapping if genomes are screened with the appropriate AIMs.

Iris pigmentation is a complex genetic trait that has long interested geneticists, anthropologists, and public at large, but is not yet completely understood. Eumelanin (brown pigment) is a light-absorbing polymer synthesized in specialized melanocyte lysosomes called melanosomes. Within the melanosomes, the tyrosinase (TYR) gene product catalyzes the rate-limiting hydroxylation of tyrosine to 3,4-dihydroxyphenylanine, or DOPA, and the resulting product is oxidized to DOPA quinone to form the precursor for eumelanin synthesis. Though TYR is centrally important for this process, pigmentation in animals is not simply a Mendelian function of TYR or any other single protein product or gene sequence. In fact, study of the transmission genetics for pigmentation traits in man and various model systems suggests that variable pigmentation is a function of multiple, heritable factors whose interactions appear to be quite complex (see, e.g., Akey et al., supra, 2002; Box et al., J. Invest. Dermatol. 116, 224-229, 2001). For example, unlike human hair color (Sturm et al., Gene 277:49-62, 2001), there appears to be only a minor dominance component for mammalian iris color determination (Brauer and Chopra, Anthropol Anz. 36:109-120, 1978), and there exist minimal correlation between skin, hair and iris color within or between individuals of a given population. In contrast, between-population comparisons show good concordance; populations with darker average iris color also tend to exhibit darker average skin tones and hair colors. These observations suggest that the genetic determinants for pigmentation in the various tissues are distinct, and that these determinants have been subject

to a common set of systematic and evolutionary forces that have shaped their distribution in the world populations.

[0280] At the cellular level, variable iris color in healthy humans is the result of the differential deposition of melanin pigment granules within a fixed number of stromal melanocytes in the iris. The density of granules appears to reach genetically determined levels by early childhood and usually remains constant throughout later life, though a small minority of individuals exhibit changes in color during later stages of life. Pedigree studies suggested iris color variation is a function of two loci; a single locus responsible for depigmentation of the iris, not affecting skin or hair, and another pleiotropic gene for reduction of pigment in all tissues (Brues, Amer. J. Phys. Anthropol. 43:387-91, 1975).

[0281] Most of what is now known about pigmentation has been derived from molecular genetics studies of rare pigmentation defects in man and model systems such as mouse and Drosophila. For example, dissection of the oculocutaneous albinism (OCA) trait in humans has shown that many pigmentation defects are due to lesions in the TYR gene, resulting in their designation as tyrosinase (TYR) negative OCAs (see, e.g., Oetting and King, Hum. Mutat. 13: 99-115, 1999; see, also, Albinism database, on world wide web ("www"), at URL "cbc.umn.edu/tad/"). TYR catalyzes the rate-limiting step of melanin biosynthesis and the degree to which human irides are pigmented correlates well with the amplitude of TYR message levels. Nonetheless, the complexity of OCA phenotypes has illustrated that TYR is not the only gene involved in iris pigmentation. Though most TYR- negative OCA patients are completely de-pigmented, dark-iris albino mice (C44H), and their human type IB oculocutaneous counterparts exhibit a lack of pigment in all tissues except for the iris (Schmidt and Beermann, Proc. Natl. Acad. Sci. USA 24;91:4756-4560, 1994). Study of a number of other TYR-positive OCA phenotypes showed that, in addition to TYR, the oculocutaneous 2 (OCA2; Durham-Pierre et al., Nature Genet. 7:176-179, 1994; Durham-Pierre et al., Hum. Mutat. 7:370-373, 1996; Gardner et al., Hum. Mutat. 7:370-373, 1992), tyrosinase like protein (TYRP1; Boissy et al., Amer. J. Hum. Genet. 58:1145-1156, 1996), melanocortin receptor (MC1R; Robbins et al., supra, 1993; Smith et al., supra, 1998; Flanagan et al., supra, 2000) and adaptin 3B (AP3B; Ooi et al., EMBO J. 16:4508-4518,

134

1997) loci, as well as other genes (reviewed by Sturm et al., *supra*, 2001) are necessary for normal human iris pigmentation. Each of these genes is part of the main (*TYR*) human pigmentation pathway.

[0282] In Drosophila, iris pigmentation defects have been ascribed to mutations in over 85 loci contributing to a variety of cellular processes in melanocytes (Ooi et al., *supra*, 1997), but mouse studies have suggested that about 14 genes preferentially affect pigmentation in vertebrates (reviewed in Strum et al., *supra*, 2001), and that disparate regions of the *TYR* and other *OCA* genes are functionally distinct for determining the pigmentation in different tissues. Human pigmentation genes break out into several biochemical pathways, including those for tyrosinase enzyme complex formation on the inner surface of the melanosome, hormonal and environmental regulation, melanoblast migration and differentiation, the intracellular routing of new proteins into the melanosome and the proper transportation of the melanosomes from the body of the cell into the dendritic arms towards the keratinocytes. Nonetheless, the study of human OCA mutants suggests that the number of phenotypically active pigmentation loci is manageably small for genetic analysis.

[0283] Though research on pigment mutants has made clear that a small subset of genes is largely responsible for catastrophic pigmentation defects in mice and man, it remains unclear whether or how common SNPs in these genes contribute towards (or are linked to) natural variation in human iris color. A brown-iris locus was localized to an interval containing the OCA2 and MYO5A genes (Eiberg and Mohr, Eur. J. Hum. Genet. 4:237-241, 1996), and specific polymorphisms in the MC1R gene are associated with red hair and blue iris color in relatively isolated populations (see, e.g., Robbins et al., supra, 1993; Flanagan et al., supra, 2000; Valverde et al., Nature Genet. 11:328-330, 1995; Schioth et al., Biochem. Biophys. Res. Comm. 260:488-491, 1999). An ASIP polymorphism was reported to be associated with both brown iris and hair color (Kanetsky et al., Amer. J. Hum. Genet. 70:770-775, 2002). However, the penetrance of each of these alleles appears to be low and, in general, appears to explain but a very small amount of the overall variation in iris colors within the human population (Spritz et al., Nature Genet. 11:225-226, 1995). However single gene studies have not provided a sound basis for understanding the complex genetics of human iris color.

[0284] Because most human traits have complex genetic origins, wherein the whole often times is greater than the sum of the parts, innovative genomics-based study designs and analytical methods for screening genetic data *in silico* are needed that are respectful of genetic complexity - for example, the multi-factorial and/or phase known components of dominance and epistatic genetic variance. The first step however is to define the complement of loci that on a sequence level explain variance in trait value, and of these, those that do so in a marginal, or penetrant sense will be the easiest to find. It is towards this goal that the present study was performed.

[0285] A non-systematic, hypothesis-driven genome screening approach was applied to identify various SNPs, haplotypes and diplotypes marginally (i.e., independently) associated with iris color variation. As disclosed in this Example, a surprisingly large number of polymorphisms in a large number of genes were associated with iris colors, indicating that the genetics of iris color pigmentation are quite complex. The sequences that were identified provide the basis of a classifier model for the inference of iris colors from DNA, and the nature of some of these as markers of BioGeographical Ancestry has implications for the design of other complex trait gene mapping studies.

#### **METHODS**

#### Specimen Collection

[0286] Specimens for re-sequencing were obtained from the Coriell Institute in Camden, New Jersey. Specimens for genotyping were of self-reported European descent, of different age, sex, hair, iris and skin shades and they were collected using informed consent guidelines under IRB guidance. Donors checked a box for blue, green, hazel, brown, black or unknown/not clear iris colors, and each had the opportunity to identify whether iris color had changed over the course of their lives or whether the color of each iris was different. Individuals for whom iris color was ambiguous or had changed over the course of life were eliminated from the analysis.

[0287] For 103 of the subjects, iris colors were reported using a number from 1-11 as well, where 1 is the darkest brown/black and 11 is the lightest blue identified using a color placard. For these subjects, digital photographs of the right iris were obtained, where subjects peered

into a box at one end, at the camera at the other to standardize lighting conditions and distance, and from which a judge assigned the sample to a color group. Comparing the two, 86 of the classifications matched. Of the 17 that did not, 6 were brown/hazel, 7 were green/hazel and 4 were blue/green discrepancies though none were gross discrepancies such as brown/green, brown/blue or hazel/blue. Though such an error is tolerable for identifying sequences marginally associated with iris colors, confidence can be increased for use of the sequences described herein for iris color classification by obtaining digitally quantified iris colors.

#### **SNP Discovery**

[0288] Candidate SNPs were obtained from the NCBI:dbSNP database, which generally provided more candidate SNPs that were possible to genotype. Human pigmentation and xenobiotic metabolism genes were examined, selected based on their gene identities not their chromosomal position. For some genes, the number of SNPs in the database was low and/or some of the SNPs were strongly associated with iris colors, warranting a deeper investigation. For these genes, re-sequencing was performed; of the genes disclosed herein, 113 SNPs were discovered in the CYP1A2 (7 gene regions, 5 amplicons, 10 SNPs found), CYP2C8 (9 gene regions, 8 amplicons, 15 SNPs found), CYP2C9 (9 gene regions, 8 amplicons, 24 SNPs found), OCA2 (16 gene regions, 15 amplicons, 40 SNPs found), TYR (5 gene regions, 5 amplicons, 10 SNPs found) and TYRP1 (7 gene regions, 6 amplicons, 14 SNPs found; see Tables 9 and 10; see, also, Intl. Publ. No. WO 02/097047).

TABLE 9

Candidate Genes Tested for Sequence Associations with Human Iris Pigmentation

Gene	Name	Homology/Model Phenotype
AP3B1	adaptor-related protein	mouse "pearl"
	complex 3, beta 1 subunit	human HPS2
ASIP	agouti signaling protein	mouse "agouti"
DCT	dopachrome tautomerase	TYR-related protein 2
		mouse "slaty"
MC1R	melanocortin 1 receptor	mouse "extension" (e)
OCA2	oculocutaneous albinism II	mouse pink-eyed dilution (p)

137

SILV	silver homologue	mouse "silver" (si)
TYR	tyrosinase	mouse "albino" (c),
<b>ም</b> የሙከ1		Himalayan
TYRP1	tyrosinase related protein 1	mouse "brown" (b)
MYO5A	myosin VA	mouse "dilute" (d)
POMC	proopiomelanocortin	mouse Pomc1
AIM	membrane associated	mouse "underwhite" (uw)
(MATP or AIM-1)	transporter protein	
AP3D1	adaptor-related protein	mouse "mocha" (mh)
	complex 3, delta 1 subunit	
RAB	RAB27A oncogene	mouse "ashen" (ash)

[0289] Resequencing for these genes was performed by amplifying the proximal promoter (avg. 700 bp upstream of transcription start site), each exon (avg. 1400 bp), the 5' and 3' ends of each intron (including the intron-exon junctions, average size about 100 bp) and 3' UTR (avg. 700 bp) sequences from a multiethnic panel of 672 individuals (450 individuals from the Coriell Institutes DNA Polymorphism Discovery Resource, 96 additional European Americans, 96 African Americans and 10 Pacific Islander, 10 Japanese and 10 Chinese; this 672 represented a separate set of samples than that used for the association study described herein). PCR amplification was accomplished using pfu TURBO polymerase according to the manufacture's guidelines (Stratagene). A program was used to design re-sequencing primers in a manner respectful of homologous sequences in the genome, to insure that pseudogenes were not co-amplified or that sequences from within repeats were amplified. BLAST searches confirmed the specificity of all primers used. Amplification products were subcloned into the pTOPO® sequencing vector (Invitrogen) and 96 insert positive colonies were grown for plasmid DNA isolation (the use of 670 individuals for amplification step reduced the likelihood of an individual contributing more than once to this subset of 96 selected).

[0290] Sequencing was perform with an ABI3700 sequencer using PE Applied Biosystems BDT chemistry; sequences were deposited into a commercial relational database system (iFINCH, Geospiza; Seattle WA). PHRED qualified sequences were imported into the CLUSTAL X alignment program and the output of this was used with a second program to identify quality-validated discrepancies between sequences. Those sequences for which at

least two instances of PHRED score 24 or greater variants were identified were selected, and each of these SNPs discovered through re-sequencing was used for genotyping.

### Genotyping:

[0291] For most of the SNPs, a first round of PCR was performed on the samples using the high-fidelity DNA polymerase *pfu* TURBO polymerase and the appropriate resequencing primers. Representatives of the resulting PCR products were checked on an agarose gel, and first round PCR product was diluted,, then used as template for a second round of PCR. The two rounds were necessary because many of the genes queried were members of gene families; the SNPs resided in regions of sequence homology and the genotyping platform required short (approximately 100bp) amplicons. For the remaining samples, only a single round of PCR was performed. Genotyping was performed for individual DNA specimens using a single base primer extension protocol and an SNPstreamTM 25K/Ultra High Throughput (UHT) instrument (Orchid Biosystems; Princeton NJ). Genotypes were subject to several quality controls; two scientists independently pass/fail inspected the calls, requiring an overall UHT signal intensity greater than 1,000 for >95% of genotypes and clear signal differential between the averages for each genotype classes (i.e. clear genotype clustering in 2-D space using the UHT analysis software).

#### **Statistical Methods**

[0292] To test the departures from independence in allelic state within and between loci, the MLD exact test was used (Zaykin et al., supra, 1995). Haplotypes were inferred using the haplotype reconstruction method (Stephens et al., supra, 2001). To determine the extent to which extant iris color variation could be explained by various models,  $R^2$  values were calculated for SNPs, Haplotypes, and Multilocus Genotypes data by first assigning the phenotypic value for blue eye color as 1, green eye color as 2, hazel eye color as 3, and brown eye color as 4. BGA admixture proportions were determined as described (Hanis et al., supra, 1986; Shriver et al., supra, 2003) within the context of a software program developed for this purpose. For  $R^2$  computation, the following function was used:  $Adj-R^2 = 1-\{(n/(n-p))\}(1-R^2)$ , where n is the model degrees of freedom and n-p is the error degrees of freedom. To correct for multiple tests, the empirical Bayes adjustments for

multiple results method was used (Steenland et al., Cancer Epidemiol. 9:895-903, 2000, which is incorporated herein by reference).

#### RESULTS

[0293] To identify SNP loci associated with variable human pigmentation, 754 SNPs were genotyped, including 335 SNPs within pigmentation genes (*AP3B1*, *ASIP*, *DCT*, *MC1R*, *OCA2*, *SILV*, *TYR*, *TYRP1*, *MYO5A*, *POMC*, *AIM*, *AP3D1* and *RAB*, see Table 9), and 419 other SNPs distributed throughout the genome. Alleles for these latter SNPs were informative for certain elements of population structure; 71 were selected from a screen of the human genome based on their exceptionally high δ values (i.e., exceptional AIMs) for IndoEuropean, sub-Saharan African, Native American and East Asian BGA (see Example 2; SEQ ID NOS:1 to 71; see, also, Shriver et al., *supra*, 2003), and the rest were found in or around xenobiotic genes, which tend to exhibit dramatic sequence variation as a function of BGA. Genotypes for these 754 candidate SNPs were scored for 851 European derived individuals of self-reported iris colors (292 blue, 100 green, 186 hazel and 273 brown).

[0294] Before screening these genotypes for association with iris colors, the 71 non-xenobiotic metabolism AIMs were used to determine BGA admixture proportions for each sample, and were tested for correlation between BGA admixture and iris colors. This test showed that each of the 851 Caucasian samples was of majority IndoEuropean BGA, and, though 58% of the samples were of significant (>4%) non-IndoEuropean BGA admixture, there was no correlation between low levels (less than 33%) of East Asian, sub-Saharan African or Native American admixture and iris colors, and no correlation between higher levels (greater than 33% but lower than 50%) of Native American admixture and iris colors; there was, however, a weak association between higher levels of East Asian and sub-Saharan African admixture and darker iris colors.

[0295] It was unclear from the outset whether better success would be realized by considering iris color in terms of 4 colors (blue, green, hazel and brown) or groups of colors. One method of grouping colors is light = blue + green and dark = hazel + brown, and this grouping seems to more clearly distinguish individuals with respect to the detectible level of eumelanin (brown pigment). Given that iris color data was self-reported, partitioning the

sample into brown and not-brown, or blue and not blue could provide greater power to detect significant associations, particularly for alleles associated with only one color. To take advantage of each of these 4 methods, all were considered when screening SNPs for associations; the δ value, chi square and exact test p-values were calculated for a) all 4 colors, b) shades, using light (blue and green) vs. dark (hazel and brown), c) blue vs. brown, and d) brown vs. not-brown (blue, green and hazel) groupings. Significance levels were fixed at 5%, and the alleles of 20 SNPs were associated with specific iris colors, 20 with iris color shades, 19 with blue/brown color comparisons and 18 using the brown/not brown comparison. The overlap among these SNP sets was high but not perfect; a SNP with a significant p-value for association using at least one of the four criteria is indicated as "marginally" associated.

[0296] When multiple simultaneous hypotheses are tested at set p-values there is the possibility of enhanced type I error. As such, a correction procedure was used to compensate for this risk (Steenland et al., supra, 2002); most of the associations were significant after this correction. Most of the marginally associated SNPs were within the pigmentation genes - OCA2 (11 SNPs at the level of colors), TYRP1 (3 SNPs at the level of colors), MYO5A (2 SNPs at the level of colors), AIM (3 SNPs at the level of colors) and DCT (2 SNPs at the level of colors) - though some associations were found within non-pigmentation genes such as CYP2C8 at 10q23, CYP2C9 at 10q24, CYP1B1 at 2p21 and MAOA at Xp11.3 - also referred to as marginally associated SNPs. No significant SNP associations were found within the pigmentation genes SILV, MC1R, ASIP, POMC, RAB or TYR, though TYR had one SNP with a p=0.06. The most strongly associated of the marginally associated SNPs were from the OCA2, TYRP1 and AIM genes, in order of the strength of association.

[0297] Since most of the SNPs identified from this approach localized to discrete genes or chromosomal regions, all of the SNPs from each locus were grouped and inferred haplotypes were tested for association with iris colors using contingency analysis. This higher-order analysis was not confined to those genes with marginal SNP associations, but grouped SNPs for all of the genes tested. For each gene, haplotypes were inferred and contingency analyses was used to determine which haplotypes were statistically associated with iris colors. From

141

the chi-square and adjusted residuals, 43 haplotypes for 16 different loci were either positively (agonist) or negatively (antagonist) associated with iris colors (Table 10). The strongest associations were observed for genes with SNPs that were marginally associated; most of these genes had haplotypes and diplotypes (sometimes referred to as multilocus genewise genotypes or diploid pairs of haplotypes) positively (agonist) or negatively (antagonist) associated with at least one iris color (Table 10). A few of the genes/regions not harboring a marginally associated SNP had haplotypes and diplotypes either positively and/or negatively associated with iris colors (ASIP gene - 1 haplotype, MCIR gene-2 haplotypes, Table 10). In other words, their SNPs were only associated with iris colors within the context of gene haplotypes or diplotypes. For some, associations with iris colors were only found within the context of diplotypes, but not at the level of the SNPs or haplotype (i.e. SILV and GSTT2-22q11.23).

Table 10

The common haplotypes and diplotypes for the 16 iris color genes

Gene au Sequenc			Agonist ²	A2	A 4		
-	<i>-</i>	T 1 160 1			Antagonist ²	•	2
ID. AIM		Haplotypes-16 Genes ¹	Chi P	Color	Chi P	Color	Count ³
	1	CAC	0.0010	Blue	0.004	Brown	(1641)
	2	TGT	0.048	Brown	0.003	Blue	(33)
	3	TAC					(23)
ASIP				•			
•	1	ATA		***			(979)
	2	ATG					(508)
	3	GCG					(196)
	4	ACA	0.017	Hazel			(13)
DCT							` ,
	1	CTGACA		~~*	*en	-	(625)
	2	CTCACA	0.014	Hazel	0.028	Blue	(242)
			0.016	Blue	0.021	Brown	` ,
	3	TCGACA	0.048	Green	0.003	Hazel	(281)
	4	CTCGTA					(320)
	5	CTGACG					(179)
MC1R							• • • •
-	1	TCC	0.016	Green			(152)

142

Gome	<b>t</b>				•	
Gene		A2				
Sequer ID.	Haplotypes-16 Genes	Agonist ² Chi P		Antagonist ²		. 7
ID.	2 CCC	cm P	Color	Chi P	Color	Count ³
	3 CCT			0.026	Green	(1294)
	4 CTC					(143)
MYO5						(113)
WITOS	1 CGATCGGCC	C 0.000	<b>C</b>			/max
•	2 CGATCAGCC		Green			(51)
	4 GTGCTGATC			0.000	D1	(858)
	5 CGATCAACC		Blue	0.008	Blue	(163)
	6 GTACTGATC		Dine			(40)
	8 CGACTGGTT			***		(117)
	10 CGACCAGCC		D			(165)
	13 CTACTGGTT		Brown	******	·	(40)
	14 CGATTAGCC			0.007	 To1	(71)
	16 CGGCCAATC			0.027	Blue	(40)
OCA2	10 COOCCAATC			***		(19)
OCAZ	GGGGACGGC	A 0.002	Dlan	0.01	D	
	1 AAG	N 0.002	Blue	0.01	Brown	(44)
	GAGGCCGGC	A 0.018	Hazel	0.031	Dlue	(44)
	2 AGA	. 0.016	Hazei	0.031	Blue	(42)
	GAGGCCAGC	A 0.022	Brown	0.026	Blue	(43)
	3 AGA	0.042	Green	0.020	Diue	(21)
	GAGGCCGGC		Brown	0.014	Blue	(21)
	4 AAA	- 0.02,	210 1/11	0.011	Blue	(89)
	GAGGCCAGC	A				(0)
	7 AAA					(13)
	TGGGACGCTA	<i>\</i>		***		()
	15 A A G					(17)
	GAGGCCAGC	G 0.019	Brown			
	19 A G A					(23)
	GAGGCCGGC	3 · 0.036	Green	0.012	Blue	, ,
	22 A A A					(15)
	TAAGCCAGC	}				
	25 A A A					(13)
	GGAAATAGCA	A <0.001	Blue	< 0.001	Brown	
	37 A A A					(508)
	GGAAATAGC	G 0.025	Blue	0.011	Brown	
	38 AAA					(200)
	GGAAATAGCA	4	~~~	0.039	Brown	
	39 AGA	•				(71)
	GGAAATAGCA	A	***			4
	41 . GAA					(19)

143

Gene	and						•
Seque			Agonist ²	Agonist ²	Antagonist ²	Antagonist ²	
ID.		Haplotypes-16 Genes ¹	Chi P	Color	Chi P	Color	Count ³
		GGAAATAGCG	0.029	Blue	0.021	Brown	Count
	42				7.021	DIOWII	(174)
		TGAAATAGCG	< 0.001	Brown	0.007	Blue	(174)
	45					2100	(65)
		TGAAATAGCG	0.003	Brown	0.006	Hazel	(00)
	47						(22)
•	4.5	TGAAATAGCA	0.001	Brown	0.036	Blue	( "/
	48				0.015	Green	(35)
		TGAAATAGCA					• •
DOM C	57	AGA	•				(30)
POMC		<b>7</b> 0			•		
	1	T			,		(1187)
	2/2	C					(515)
OIL V	2/2	C/C	0.043	Blue	***		
SILV	1	Tr C			•		
	1	TC					(913)
	2	CC	tereting		***		(296)
	3 1/1	TT					(490)
TYR	1/1	TC/TC			0.035	Blue	
IIK	1				0.04	_	
	1	GCG			0.01	Green	(184)
	2	AAG GCA					(257)
	4	AAA		ter #*			(467)
	5	ACA			******		(231)
	6	ACG	The state of the				(254)
	7	GAG					(189)
	8	GAA			0.022	D1	(83)
TYRP	U	UAA			0.023	Blue	(37)
* * * * * * * * * * * * * * * * * * * *	1	TTTTCG	0.007	Blue	< 0.001	Decree	(0.00)
	2	CTTTTT	0.007	Diue	<0.001	Brown	(968)
	3	CTGACG	0.006	Brown			(86)
	4	CCGACG	0.046	Brown	0.029	Blue	(454)
	5	TTGTCG		DIOWII	0.023	Diue	(98)
	7	CTTACG		•••			(23)
CYP1A		,					(24)
15q22-			•				
•	1	G	0.015	Brown	0.023	Hazel	(769)
	2	C	0.023	Hazel	0.015	Brown	(933)
CYP1E	31-						(200)

144 . .

Gene					2		
Seque	ence	<b></b> 1	Agonist ²	Agonist ²		Antagonist ²	_
ID.		Haplotypes-16 Genes ¹	Chi P	Color	Chi P	Color	Count ³
2p21	_				•		
	1	CC					(957)
	2	TT	0.027	Hazel	0.025	Blue	(403)
	3	CT			****		(337)
CYP2	2C8-					*	• •
10q23	3						
	1	CAA	0.007	Brown			(539)
	2	TAG		~~~			(201)
	3	TGA			0.039	Brown	(513)
	4	TAA			,		(439)
CYP2	2C9-	•					
10q24	1	•					
	1	T Č	0.023	Brown	0.039	Green	(1325)
	2	Ċ	0.039	Green	0.023	Brown	(377)
GSTT	[2-						` ,
22q11	1.23		,				
	1	A G					(821)
•	2	G A			the part part		(778)
	3	AA				***	(99)
•		AG/AG	0.040	Green			(184)
		AG/GA	0.013	Hazel		· ·	(401)
· MAO	Α-						` '
Xp11.	.4-11						
_	1	CG			0.027	Blue	(1133)
	2	T A	0.006	Blue	0.026	Hazel	(480)
	3	CA					(87)
							• •

¹ Sequences of the highest order of complexity within a locus found to be associated with iris colors. All of the major sequences (count  $\geq$  13) for each locus with at least one significantly associated sequence are shown. If no haplotypes or diplotypes for a locus were found to be associated, only the SNP alleles are shown. If no haplotypes were found to be associated for a locus but diplotypes were found to be associated, both the haplotypes and diplotypes are shown.

² Agonist color refers to the color with which the sequence is positively associated. Antagonist color refers to the color with which the sequence is negatively associated. Chi-square P-value is shown.

Number of times the haplotype was observed in our sample of 851.

[0298] At the level of the haplotype, each gene or region had unique numbers and types of associations. For example, OCA2, AIM, DCT and TYRP1 harbored haplotypes both positively associated with blue irides and negatively associated with brown irides (OCA2 haplotypes 1, 37, 38, 42, AIM haplotype 1, DCT haplotype 2, and TYRP1 haplotype 1 Table 10). Others genes such as AIM, OCA2 and TYRP1 harbored haplotypes that were positively associated with brown but negatively associated with blue color (AIM haplotype 2, OCA2 haplotypes 2, 4, 45, 47, TYRP haplotype 4, Table 10), while others such as MYO5A, OCA2, TYRP1 and CYP2C8-10q23 harbored haplotypes that were positively associated with one color but not negatively associated with any other color (MYO5A haplotype 5, haplotype 10, OCA2 haplotype 19, TYRP1 haplotype 3 and CYP2C8-10q23 haplotype 1, Table 10). The MC1R gene harbored haplotypes only associated with green color in our sample and the POMC gene harbored a single SNP with genotypes weakly associated with iris colors (no significant haplotypes or diplotypes were found).

[0299] Overall, the diversity of haplotypes associated with brown irides was similar to that of haplotypes associated with blue irides. Most of the haplotypes were even more dramatically associated with iris colors in a multi-racial sample, because many of the SNPs comprising them are good AIMs, and variants associated with darker iris colors were enriched in those ancestral groups of the world that are of darker average iris color. Most of the SNPs within a gene or region were in LD with others in that gene or region (D'<0.1); only 32 SNP pairs, in the MC1R (1 pair), OCA2 (27 pairs), TYR (2 pairs) and TYRP1 (2 pairs) genes were found to be in LD.

[0300] These analyses resulted in the identification of 61 SNPs in 16 genes/ chromosomal regions associated with iris colors on one level or another, whether the SNP is marginally associated or associated within the context of the haplotype and/or diplotype. The minor allele frequency for most of these SNPs was relatively high (avg. f minor allele = 0.22) and most of them were in Hardy Weinberg Equilibrium (those for which HWE p>0.05, 28/34, Table 10). Nine were not and of these, 2 were of relatively low frequency and the evidence for disequilibrium was marginal (p value close to 0.05). Lack of HWE is usually an indication of a poorly designed genotyping assay, and none of the remaining 7 SNPs

exhibited genotyping patterns that we have previously associated with such problems (such as an absence of one genotype class, or a preponderance of heterozygotes). Indeed one of those for which the evidence of lack of HWE was the strongest was validated as a legitimate SNP through direct DNA sequencing. The chromosomal distribution of the SNPs that were significantly associated in a marginal sense was independent of the distribution of SNPs actually surveyed, indicating that the associations were not merely a function of SNP sampling..

[0301] Chromosome 15q harbored the majority (18/34) of the SNPs that were marginally associated with iris colors, and 14 of these chromosome 15 SNPs were found in two different genes OCA2 and MYO5A. Chromosome 5p had 3 SNPs marginally associated, all in the AIM gene and chromosome 9p had5 SNPs associated, all in the TYRP1 gene. Multiple SNPs were identified on chromosome 10q; the CYP2C8-10p23.33 region had 2 SNPs, and the neighboring region CYP2C9-10p24 also had one. All 3 markers were in tight LD with one another (p<0.001 for each possible pair). Multiple SNPs were also identified on chromosome 2; the POMC SNP located at 2p23 was marginally associated, and SNPs from the CYP1B1-2p21 region were associated within the context of a 2-SNP haplotype (Table 10), and these SNPs were also in LD (p<0.01). Finally, in addition to the OCA2 (15q11.2-q12) and MYO5A (15q21) sequences, a single SNP (15q22-ter) was also implicated on chromosome 15q, but SNPs between each of these three loci were not in LD. SNPs for the MC1R (16q24), SILV (12q13), TYR (11q), MAOA-Xp11.4-11.3 and GSTT2-22q11.23 regions were also associated at the level of the haplotype (Tables 10 and 11), though these were the only regions of these chromosomes for which associations were found.

[0302] The p-values that were obtained indicated that diplotypes explained more iris color variation than haplotypes or individual SNPs. To test this, a corrected ANOVA analysis was performed for the data on each of these three levels. All 61 SNPs were considered, as were their haplotypes (Table 10) and diplotypes (not shown). Diplotypes explained 15% of the variation, whereas haplotypes explained 13% and SNPs explained 11% (Table 4) after correcting for the number of variables. The most strongly associated 68 genotypes of the

147

543 genotypes observed for the 16 genes/regions, based on chi-square adjusted residuals, explained 13% of the variation (row 4, Table 11).

Table 11 ANOVA- SNP and Haplotype data

				No.			
Row	Source	Model DF	Error DF	Variables	F Value	R2 Value	Adj.R ²
1	SNP data	62	788	62	2.63	0.17	0.11
2 -	Haplotype data	212	638	216	1.58	0.34	0.13
	Multilocus Gene-wise	;					0.15
3	Genotype data	543	307	572	1.27	0.69	,
	Multilocus Gene-wise	,					0.13
4	Genotype data	68	782	68	2.82	0.2	

From a screen of 754 SNP loci, 61 were identified that were statistically associated [0303] with variable iris pigmentation at one level of intragenic complexity or another. The remaining SNPs had δ values and chi-square p-values that were not significant on any level of intragenic complexity. Diplotypes for these 61 alleles explained most of the iris color variance in the sample; the lowest amount was explained at the level of the SNP, suggesting an element of intragenic complexity to iris color determination (i.e., dominance).

[0304] Only about half of the 61 SNPs identified were associated with iris colors independently; the others were associated only in the context of haplotypes or diplotypes. Even at this level of complexity, the sequences from no single gene could be used to make reliable iris color inferences, indicating an element of intergenic complexity (i.e., epistasis) for iris color determination as well. Aside from the fact that many of the identified SNPs were significant after imposing the correction protocol for multiple testing, five lines of evidence indicated that the identified SNPs are not spuriously associated. First, for all of the genes identified as marginally associated SNPs, multiple such SNPs were identified; i.e., the distribution of SNPs among the various genes tested was not random. Second, some of the non-pigment gene SNPs are located near pigment genes, e.g., CYP2C8 (10q24.1) and CYP2C9 (10q24), which are located proximal to two pigment genes not tested directly -HPSI (10q23.1-4) and HPS6 (10q24.34) and the chromosome 2p SNP at the CYP1B1 locus

148

(CYP1B1-2p21) located proximal to POMC at 2p23 (and in LD with the POMC SNP). Third, though a roughly equal number of pigmentation and non-pigmentation gene SNPs were tested, of the 34 marginally associated SNPs, 28 of them (82%) were in pigmentation genes. Thus, the distribution of SNPs among the various gene types also was not random. Fourth, the associations were generally stronger for the SNPs in the context of within-gene haplotypes, a result that would not necessarily obtain for SNPs spuriously associated (i.e., the result suggests the gene sequences themselves are associated, not merely a single polymorphism within each gene). Fifth, when applied to a sample including individuals of multiple ancestries, the linear and non-linear variables from these and the other genes combined performed even better than when applied just to individuals of majority European ancestry. Since most individuals of non-European, or minority European descent exhibit low variability in iris colors (on average of darker shade than individuals of European descent) this improvement may not seem surprising. However, this result would not have necessarily been obtained were the SNPs not truly associated with iris colors.

[0305] Though corrections for multiple testing left most of the SNP-level associations intact, a number of the associations did not pass the multiple-testing examination, but are presented in order to avoid possible type II error; the sequences may be weakly associated with iris colors and possibly relevant within a multiple-gene model for classification (i.e., epistasis). For these, it would seem more prudent to eliminate false-positives downstream of SNP identification, such as from tests of higher order association, using various other criteria such as those described above, or possibly using the utility of the SNP for the generalization of a complex classification model.

[0306] Mutations in the pigmentation genes are the primary cause of oculocutaneous albinism, so it was natural to expect that common variations in their sequence explain some of the variance in natural iris colors and, in fact, this result was observed. However, a number of the associations were for SNPs located in other types of genes (CYP2C8 at 10q23, CYP2C9 at 10q24, CYP1B1 at 2p21 and MAOA at Xp11.3). The inclusion of non-pigmentation genes in this study was intentional; the screen was not restricted to pigmentation gene SNPs, but included two types of AIMs – those that were selected from the

149

genome based on  $\delta$  values between sub-Saharan, IndoEuropean, Native American and East Asian population allele frequencies, and those selected based on their location within xenobiotic metabolism genes. The latter are included based, in part, on previous evidence indicating that xenobiotic metabolism genes harbor an unusual concentration of AIMs, and that some of these AIMs are relevant for the measurement of "cryptic" population structure, presumably because xenobiotic metabolism gene products are responsible for detoxification of floral alkaloids and tannins present in indigenous diets, and selection and genetic drift have shaped the geographical distribution of their sequences. Such cryptic structure may be correlated with iris colors to a degree that enables accurate classification, even though they may not be helpful for elucidating a biological mechanism.

It was hypothesized that a) some of these SNPs would be indicators of not merely crude or continental population structure, but sub-structure and perhaps even microstructure, b) iris colors were correlated with these elements of structure within the Caucasian group. and c) these markers can serve as proxies for phenotypically active loci for the purpose of classification or trait value inference. The commonly held notion is that genetic screening is only properly conducted towards identifying phenotypically active loci through linkage disequilibrium. However, when classification is the goal, rather than the identification of phenotypically active loci, population structure can be helpful if the trait value correlates with the structure and if markers for the structure can be identified. For example, an iris color classification tool can be useful to a forensic scientist for the objective and science-based construction of a partial physical profile from crime scene DNA. Currently, forensics investigators construct physical profiles using surprisingly unscientific means; only in rare cases are eye-witness accounts available, and often times human accounts are subjective and unreliable in certain circumstances. For a forensics application, an investigator is less interested in the biological mechanism of the phenotype than in an ability to make an accurate inference of trait value. Of course, identifying markers in LD with phenotypically active loci (or the phenotypically active loci themselves) provide for more accurate classification, as well as for a better understanding of biological mechanism, but the hunt for these elusive loci in heterogeneous populations is impractical because LD only extends for a few Kb and expensive genome-wide scans are required.

150

[0308] That a number of the SNPs identified herein as associated with iris colors were located in xenobiotic metabolism genes suggests the identified markers are associated with iris colors through correlation with cryptic population structure. In other words, the non-pigmentation gene markers are probably correlated with, but not necessarily in LD with, phenotypically active loci for iris color. Through such a correlation, both markers and active loci are enriched for in certain branches of the IndoEuropean pedigree, even though they may not be in LD with one another. These results based on such correlations are meaningful in a classification context only with respect to the sample used. For example, AIMs selected based on their inter-continental δ values were not associated with iris colors in individuals of mainly European descent, but were strongly associated with iris colors in a more international sample because the AIMs, are specifically relevant for the element of structure correlated with iris colors in this sample. In contrast, these same AIMs were not associated with iris colors within the sample of majority European-derived individuals examined because there is little variation in crude structure within this sample. Instead, within any (majority) Caucasians or European American sample, there will exist sub-structure or micro-structure (cryptic structure) due to variation in ethnic or other sub-population level affiliations, and only those SNPs specifically relevant for measuring this cryptic structure would be needed if the structure correlates with the trait. It bears noting that no systematic structure in the mainly European-derived study sample unrelated to the phenotype measured was identified, indicating that the use of qualified AIMs is imperative for reproducing the present results with another trait.

[0309] Other interpretations of the present results are possible, for example, that the associations could have been observed through LD with as of yet to be defined pigmentation genes. Indeed, CYP2C8 and CYP2C9 are located on chromosome 10 near the HPS1 and HPS2 pigmentation genes (not tested directly), CYP1A2 is located at 15q22-ter, on the same arm as OCA2 and MYO5A, CYP1B1 is located at 2p21, in the vicinity of the POMC gene at 2p23 and MAOA is located on the same arm of the X chromosome (Xp11.4-11.3) as the OA1 pigmentation gene (not tested directly). The distances between these loci associated with iris colors and "neighboring" pigmentation genes is far greater than the average extent of LD in

151

the genome, and even if these associations are through LD, it would seem that, again, population structure would need to be invoked as an explanation.

[0310] LD is known to extend for megabases in recently admixed populations, such that as few as a couple thousand AIMs can be used to gain full genome coverage in these populations, and it is of some interest that two-thirds of the European American samples used in this study were of significant (4%) BGA admixture. Though European Americans are not recognized as a traditionally defined admixed group (like Hispanics or African Americans), the BGA admixture observed may be linked to finer, cryptic levels of population structure. While the relevance of the present results to LD and/or population structure is not clear, if the results are due to LD rather than correlation, they would suggest that just as AIMS can be used to leverage population admixture for trait mapping in recently and extensively admixed populations, they also can be used to leverage cryptic population structure in a similar manner. Thus, regardless of whether the results are due to correlation or LD, the large number of non-candidate gene associations identified indicates that the measurement of population structure has broader implications for the cost-effective development of pharmacogenomics and complex disease gene classifiers.

[0311] Linkage studies have implicated certain pigmentation genes as specifically relevant for pigmentation phenotypes, and most of the pigmentation gene SNPs identified herein clustered to certain genes such as OCA2, MYO5A, TYRP1 and AIM. Further, certain aspects of the present support the previous literature. Most of the SNPs identified were on chromosome 15, which linkage analyses identified as the primary chromosome for the determination of "brownness" (Eiberg and Mohr, Eur. J. Hum. Genet. 4:237-41, 1996); it was suggested that the candidate gene within the interval containing this locus (BEY2) was most likely the OCA2 gene, though the MYO5A gene is also present within this interval and, as disclosed herein, was associated with iris colors. OCA2 associations were by far the most significant of any gene or region tested, while MYO5A SNPs were only weakly associated (but haplotypes and diplotypes more strongly). MYO5A alleles were not in LD with those of OCA2, suggesting these results were independently obtained and that results described by

152

Eiberg and Mohr may have been a reflection of the activity of two separate genes (Eiberg and Mohr, *supra*, 1996).

[0312] Two OCA2 coding changes were reported to be associated with darker iris colors (Rebbeck et al., Cancer Epidemiol. Biomarkers Prev. 11(8):782-784, 2003). In addition, the "red hair/blue iris" SNP alleles previously (Valverde et al., supra, 1995; Koppula et al., supra, 1997) were identified, confirming that these sequences are associated with iris pigmentation, though the previously described associations were with blue irides and at the level of the SNP, whereas, in the present study, the associations were with green irides and only apparent at the level of the haplotypes and diplotypes. Associations were also identified in the ASIP gene (see Kanetsky et al., supra, 2002), though, in the present study, this gene association was not at the level of the SNP; one of the ASIP SNPs identified herein (marker 861) is the 8818 G-A SNP transversion described as associated with brown iris colors (Kanetsky et al., supra, 2002) though, in the present study, the association was with hazel color at the level of the haplotype.

[0313] The associations between TYR haplotypes and iris colors was relatively weak, which is not inconsistent with results obtained by others in the field of oculocutaneous albinism who have failed to find strong associations in smaller samples. Though the present results independently verified findings for OCA2, ASIP and MC1R, they also show that several other pigmentation genes harbor alleles associated with the natural distribution of iris colors (TYRP1, AIM, MYO5A and DCT). As such, the present results indicate that most of the previous studies associating pigmentation gene alleles with iris colors, taken independently, represent mere strokes of a larger, more complex portrait.

[0314] Interestingly, most of the SNPs identified herein are non-coding, either silent polymorphisms or residing in the gene proximal promoter, intron or 3 'UTR. This result, while not altogether unusual, can indicate that the SNPs are in LD with other phenotypically active loci, or it may be a reflection that variability in message transcription and/or turnover may explain part of the variability observed in human iris colors. Though a large number of SNPs was screened, some of the genes harbor a large number of candidate SNPs and not all were tested. For example, the OCA2 has about 200 known candidate SNPs in NCBI dbSNP.

As such, the OCA2 gene may yet have more information variable human iris pigmentation, such information being accessible using methods as disclosed herein.

#### **EXAMPLE 5**

### USE OF AIMS TO PREDICT DRUG RESPONSIVENESS

[0315] This Example demonstrates that AIMs can be used to develop chemopredictive and diagnostic tests because many drug response traits, as well as many human genetic traits, correlate with elements of population structure.

[0316] The distribution of SNPs available for genotyping by chromosomal arm is shown in Figure 12. At each of the approximately 400 SNPs examined, Caucasian individuals taking LipitorTM (180), for whom response was known in terms of cholesterol (TC), low density lipoprotein (LDL), liver transaminase ASTSGOT and ALTGPT measurements, were genotyped. 150 ZocorTM patients of known response in terms of TC and LDL change, and 1,000 individuals of known hair and eye color, also were genotyped. Those SNPs with delta values of significance ( $\delta$ >0.20) among the various trait classes were selected. For example, in about 70% of patients, LipitorTM caused a decrease in LDL, whereas in 30% of patients it had no effect. For any given SNP, the delta value ( $\delta$ ) is the difference in minor allele frequency among those individuals for whom LDL decreased by at least 20% versus those for whom LDL did not so decrease. The δ value was measured for each SNP in Figure 12 for each test (ZocorTM: LDL, TC, ASTSGOT, ALTGPT response; LipitorTM: LDL, TC response). For eye color, the ôvalue was measured in terms of dark (hazel or brown) versus light (blue or green) eyes. For hair color, the δ value was measured in terms of black or brown color versus blonde.

[0317] For LipitorTM response, the number of SNPs of significant (δ>0.20) value for each of the 4 endpoint measures is shown in Figure 13. Those SNPs with delta values of significance for ZocorTM response using LDL and TC endpoint measures then were selected (Figure 14). Next, those SNPs with delta values of significance for Iris color were selected (Figure 15); and, likewise, for hair color (Figure 16). The distribution of SNPs with good δ values was similar for each trait in Graphs A-E, but with certain elements of specificity.

154

The specificity can be appreciated by focusing on chromosome arm 6p, which has many important SNPs for TC (Total Cholesterol) response to LipitorTM (Figure 13), but none for ZocorTM response(Figure 14). Chromosome 2 harbors SNPs with good δ values for eye color (Figure 16), but not hair color (Figure 15). Chromosome 15 harbors many markers predictive of LipitorTM responsiveness, but not ZocorTM response. This specificity is likely a function of linkage disequilibrium with other loci deterministic for each of these traits, but which has nothing to do with the expression of the other traits; this type of finding is the goal of traditional measures of gene mapping. Alternatively it can be due to correlation with certain elements of population structure

[0318] The similarity is apparent in that chromosomes 10 and 22 have a relatively high number of SNPs with good  $\delta$  values for each of the four mechanistically unrelated traits, as does chromosome 1. Overall, the distribution of important SNPs for one trait is not dissimilar to that for another. It is the similarity between the profiles that is of interest here, and that illustrates the value of the present methods.

[0319] The elements that the four graphs share in common correlate with the number of SNPs genotyped (Figure 12; Figures 13 to 16 roughly resemble Figure 12). At first pass, this result appears to indicate that the :importantness" or significance for these SNP alleles are spurious, and merely a function of the number of SNPs genotyped for each chromosomal arm (i.e., the more SNPs one genotypes from a chromosome, the more SNPs of good delta value one will find on that chromosome). However, the SNPs in Figure 12 are not just any type of SNP; most of the SNPs available in the Figure 12 are xenobiotic metabolism and pigmentation gene SNPs, and most all are good AIMs.

[0320] For iris color, it was shown that most of the SNP associations remain significant (chi p<0.05 after correction for multiple testing, thus indicating that the SNP associations are not spurious. That the distribution of SNPs associated for each of the 4 traits resemble one another to a large extent, and that this distribution is similar to the distribution of available SNPs, most of which are good AIMs, indicate that most of the SNPs measured in these experiments are reporters of population structure, and that similar elements of population

155

structure are correlated with response to each of the two drugs (however measured) as well as to hair and iris color pigmentation.

[0321] It is notable that these four traits appear to be mechanistically unrelated (at least to current knowledge) and it is not intuitively obvious as to how hair or eye color can relate to the response to two randomly selected drugs. However, the similarity in profiles for important SNPs for each of the traits suggests that each can be predicted, to a significant extent, with a knowledge of the sequence for a common set of chromosomal markers. Because these markers are known to be excellent indicators of BGA, the results indicate that each of the four unrelated traits can be predicted to some extent by measuring BGA rather than by measuring the particular SNPs measured in Figures 12 to 16.

[0322] Simply measuring BGA as disclosed herein does not impart as much predictive power for each of the four traits as measuring the specific AIMs in the plots above. However, using the markers in Figure 15 enabled a good classification accuracy for iris color. These results indicate that different AIMs in the plots above are informative for different elements of population structure or substructure, and while most of the SNPs in Figure 12 are good indicators of crude population structure in terms of continental BGA, it has not yet been determined to what extent each is also informative for other finer levels of structure, such as between Scandinavian and Mediterranean ancestry, or even within ethnic groups. Such "cryptic" structure could not previously be defined in a reliable, confidence qualified manner because, prior to the present disclosure, it was not possible to define the subtle elements of structure recognized in large populations of individuals of common race in biogeographically meaningful ways). For example, red haired individuals of majority IndoEuropean descent are known to require 20% more anesthesia than other IndoEuropean (or other) patients (Cohen, supra, 2002). These red haired individuals also exhibit a tendency towards hypertension and bleeding under the influence of certain common anesthetics, thus presenting a serious clinical problem of unknown etiology.

[0323] Red haired individuals are common in the United Kingdom (Ireland and Britain), which also contains individuals sharing more ancestors with one another than with individuals of other regions of Europe. As such, they can be considered as a branch of

indeterminate structure off the family tree of humanity. Only one gene is linked to red hair color (MCIR), and it is difficult to imagine this gene is of such pleiotropic effect that its sequence contributes to variability in such diverse and complex physiological responses as those to numerous anesthetics. Further, not every person with red hair harbors the known red hair MC1R variants, yet most exhibit the aberrant anesthesia response. Rather, it is more likely that red hair color correlates with an element of population structure that also correlates with anesthesia response; i.e., the gene(s) for red hair color and aberrant anesthesia response are unique to or enriched for in particular branches of a human pedigree, within which these two traits are more common. Thus, the genes for red hair color and aberrant anesthesia response are distributed as a function of population structure, and, similarly, so have many other traits, as disclosed herein.

[0324] The AIMs and methods disclosed herein are suitable for the measurement of various levels of structure, including crude continental structure, as well as structure relevant for ethnicity and even cryptic structure (e.g., almost 30 markers have been identified that are capable of resolving Pacific Islanders from other East Asians; i.e., a finer level of structure than continental structure). The informativeness of the AIMs arises over the course of evolutionary human development through founder effects, migration, bottlenecks, genetic drift and/or selection, but these forces need not be focused on hair color or anesthesia response for there to have arisen AIMs correlated with these two types of traits. Much like this case, where there must be AIMs that are informative for disparate phenotypes in North Western Continental Europeans, there also must be AIMs that are informative for disparate phenotypes in IndoEuropeans in general. The results demonstrating the similarity in AIMs with good delta values for LipitorTM response, ZocorTM response, hair color and eye color (Figures 12 to 16) are indicative of a level of population structure that is informative for each of these phenotypes.

[0325] It should be noted that the magnitude of association does not match between the traits. For example, the strongest AIM for iris color is not the strongest AIM for LipitorTM response, etc. Also, the direction of the association is not necessarily the same among the traits; thus, agonist associations (associated positively) for blue iris color can be either agonist

or antagonist associations (associated negatively) for a particular LipitorTM response outcome.

[0326] Randomly selected AIMs that distinguish specifically between Africans and East Asians can, but need not, harbor information about a particular set of traits because they can, but need not, be markers of the particular element of human population structure correlated with the trait (i.e., the branch of the human family tree within which these traits are more common). Similarly, AIMs that distinguish between Indo Europeans and Africans would not necessarily carry with them the information necessary to help with the prediction of anesthesia response or red hair. Only those AIMs with alleles that distinguish between individuals of the particular branches of humanity within which the traits values are concentrated or underrepresented carry the information necessary to predict trait value, in this example, anesthesia and red hair, or, as disclosed, response to LipitorTM, ZocorTM, hair color and eye color. Certain SNPs are good AIMs for certain gross elements of population structure (Europeans versus sub-Saharan African) or sub-substructure (Northern Europeans versus Mediterranean IndoEuropeans) or micro-structure (Scottish vs. Irish vs. British; or red haired Northern Europeans versus Northern Europeans of other hair colors; or Northern Europeans that respond to a drug versus other Northern Europeans that do not respond to the drug). Some AIMs are informative for some levels of population structure but not others, and most of the SNPs in the human genome do not carry information of any level of population structure at all (i.e. they are not AIMs).

[0327] The primary element of the disclosed methods is that most human traits can be predicted through a detailed measurement of AIMs associated with population structure at various levels, provided the trait correlates with that element of structure. A secondary element is that classifiers, or collections of SNP markers, and methods for predicting trait value from DNA can be constructed for most human traits through such an appreciation of population structure. As disclosed herein, such applications can be accomplished through correlation, not just through extended LD found in certain admixed groups such as Hispanics or African Americans, but for any sample of subjects, regardless of race or ethnic background, provided that the AIMs used are appropriate for the element of population

structure with which the trait is correlated. These results can be attained because the methods of the invention provide a means to mine the genome for good AIMs, qualify their value as AIMs and accurately measure population structure against the backdrop of human phenotypes.

[0328] The trend observed in the studies represented by Figures 12 to 16 has been observed for many other traits, where the SNP associations are of such "penetrance" that they withstand corrections for multiple tests (Steenland et al., supra, 2000). As such, by measuring AIMs across the human genome, elements of population structure, sub-structure, or microstructure relevant for predicting or inferring the value of virtually any given human trait can be learned, even if the markers are not in linkage disequilibrium with the phenotypically active loci for the trait. This correlation applies to any human trait, simple or complex, of clinical, recreational, forensics or other value, or not, because every human trait is correlated to greater or lesser degree with certain elements of population structure.

[0329] It is not possible to know a priori whether a trait segregates with population structure versus substructure versus sub-substructure versus microstructure unless one measures AIMs across the genome in individuals and identifies which (if any) correlate with trait value. Virtually any trait correlates with at least one element of structure, some with gross structure (as in the case of human skin pigmentation), some with substructure (as in the case of iris, hair or skin pigmentation between South Asian IndoEuropeans (Indians) and Northern European IndoEuropeans (e.g., Irish), and some with microstructure (as in the case of red hair or anesthesia response among Continental Europeans). It is not important to know with which level of population structure a trait correlates in order to measure and find AIMs for the inference of that trait, it is only necessary to measure and test a plurality of general AIMs for statistical association with that trait. As such, methods are provided for linking human gene sequences with traits such that they can be predicted or inferred. Such a method is valuable, for example, in the clinical and forensic fields because it is not important that biologically or mechanistically relevant sequences be measured when predicting a common trait (response to a drug or predisposition to develop disease), as it is to make an accurate inference of that common trait (drug response or disease disposition).

159

#### **EXAMPLE 6**

### ALGORITHM FOR MAXIMUM LIKELIHOOD ESTIMATE

[0330] An software program was written based on the algorithm of Hanis et al. (*supra*, 1986) for using multilocus AIM genotypes to determine the Maximum Likelihood Estimate of individual BGA admixture. A flow chart illustrating an algorithm useful for determining proportional ancestry is provided in Table 12. An example as to how the algorithm operates in shown in Table 13, and the results of an ancestry proportion calculation is shown in Table 14.

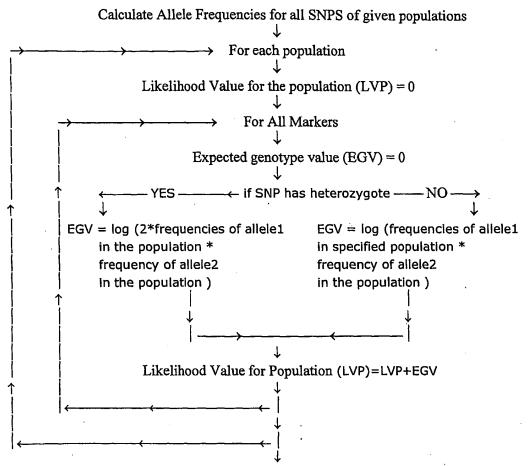
[0331] The  $\delta$  value is an expression of the ancestry informativeness of the marker (Dean et al., 1994). For a biallelic marker, the frequency differential ( $\delta$ ) is equal to  $p_x$  -  $p_y$  which is equal to  $q_y$  -  $q_x$ , where  $p_x$  and  $p_y$  are the frequencies of one allele in populations X and Y and  $q_x$  and  $q_y$  are the frequencies of the other. To test the departures from independence in allelic state within and between loci, we have used the MLD exact test (Zaykin et al., *supra*, 1995). The collection of 71 AIMs used in Example 2 was selected to maximize the cumulative  $\delta$  value within, and minimize differences in the cumulative  $\delta$  value between each of the six possible pairs of the four dimensional (sub-Saharan African, Native American, IndoEuropean and East Asian) problem.

[0332] The algorithm inverts the population specific allele frequencies to obtain a likelihood estimate of proportional affiliation corresponding to a multilocus genotype using three groups at a time; three groups were used mainly for computational convenience, and also because a 4-dimensional admixture is likely to be relatively rare. For example, the likelihood of 100% IndoEuropean, 0% Native American, 0% East Asian is calculated, then the likelihood of 99% IndoEuropean, 1% Native American, 0% East Asian is calculated next, and so on until all possible IndoEuropean, Native American and East Asian proportions are considered, and then the process is repeated for all possible IndoEuropean, Native American and African proportions, and all possible Native American, African and East Asian proportions. The likelihood of maximum value is selected as the Maximum Likelihood Estimate (MLE). When plotting a single MLE on a triangle plot, the space within which the

likelihood is within 2-fold, 5-fold and 10-fold of the MLE also are delimited (see Figure 3); these intervals generally are not plotted when multiple MLEs are shown in a single triangle plot.

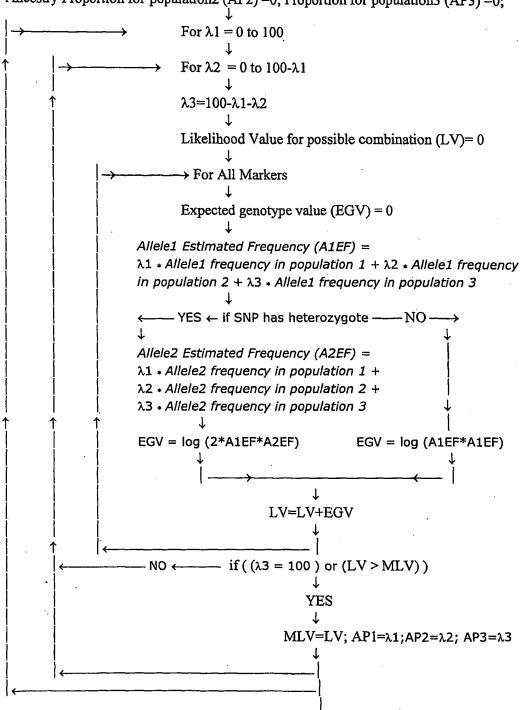
TABLE 12

Algorithm for Ancestry Calculation - Flow Chart



Select 3 populations, which have Maximum Likelihood Values

Maximum Likelihood Value (MLV)=0; Ancestry Proportion for population1 (AP1) =0; Ancestry Proportion for population2 (AP2) =0; Proportion for population3 (AP3) =0;



 $\lambda 1$ ,  $\lambda 2$ ,  $\lambda 3$  are corresponding Ancestry proportions for Blind Sample.

162

TABLE 13

Example of Proportional Ancestry Determination Using Algorithm

SAMPLEID	SNP1	SNP 2
ANC30000	GT	π

- I. Pick best three population for blind sample
- II. Get proportions which has maximum likelihood value

## 1. Pick up best three populations:

## Algorithm:

Pick best three likelihood values out of those 4 population values.

For those selected three population estimate proportions.

#### 1. Starts with

$$\lambda_{1=0}$$
,  $\lambda_{2=0}$  and  $\lambda_{3=1}$   $0+0+1=1$ 

## 2. Compute Likelihood value:

### Estimate Expected genotype for SNP1

SNP1 has heterogeneous genotype; Alleles are G and T.

## **Estimated Allele Frequency from SAMPLE:**

Allele1 Estimated Frequency (A1EF)=  $\lambda_1.p(G,1)+\lambda_2.p(G,2)+\lambda_3.p(G,3)$ 

Where p(G,1) - G Allele frequency in population 1

p(G,2) - G Allele frequency in population 2

p(G,3) - G Allele frequency in population 3

The mixing proportions  $\lambda_1$ ,  $\lambda_2$  and  $\lambda_3$  are treated as unknown parameters.

Allele2 Estimated Frequency (A2EF)= $\lambda_1$ . p(T,1)+ $\lambda_2$ . p(T,2) + $\lambda_3$ . p(T,3)

Where p(T,1) - T Allele frequency in population 1

p(T,2) - T Allele frequency in population 2

p(T,3) - T Allele frequency in population 3

The likelihood of the parameters is obtained by multiplying the probabilities for each observed genotype in the new observation under the assumption of Hardy-Weinberg Law.

Since SNP1 has heterogeneous genotype

Expected genotype for SNP1 = log(2*A1EF*A2EF);

#### Estimate Expected genotype for SNP2

#### **Estimated Aliele Frequency from SAMPLE:**

Allele1 Expected Frequency (A1EF)= $\lambda_1$ . p(T,1)+  $\lambda_2$ . p(T,2) +  $\lambda_3$ . p(T,3)

Where p(T,1) - T Allele frequency in population 1

 $p(T_12) - T$  Allele frequency in population 2

p(T,3) - T Allele frequency in population 3

The mixing proportions  $\lambda_1$ ,  $\lambda_2$  and  $\lambda_3$  are treated as unknown parameters.

Since SNP2 has Homogenous genotypes

Expected genotype for SNP2 = log(A1EF*A1EF);

164

### LIKELIHOOD VALUE

Compute LIKELIHOOD VALUE by adding all Expected genotypes of all SNPS

LIKELIHOOD = Expected genotype for SNP1 + Expected genotypes for SNP2;

Compute LIKELIHOOD value by using different unknown parameters. (Repeat step 2)

Get Maximum Likelihood value and corresponding unknown parameters.
 Those unknown parameters are nothing but proportions

TABLE 14
Ancestry Proportion Calculation:

African fi	requenc	ies			
SNP1	G	0.8	T	0.2	
SNP2	T	0.7	A	0.3	
SNP3	С	1.0			
Europear	n freque	ncies			
SNP1	G	0.9	T	0.1	
SNP2	Т	0.7	A	0.3	
SNP3	С	0.8	T	0.2	
Native A	merican	(NA) fi	reque	ncies	
SNP1	G	0.6	T	0.4	
SNP2	Т	0.5	A	0.5	
SNP3	С	0.7	T	0.3	
Middle East (ME) frequencies					
SNP1	G	0.7	T	0.3	
SNP2	T	0.9	A	0.1	
SNP3	C	0.9	T	0.1	

**Ancestry Frequency Table** 

165

#### **EXAMPLE**

SAMPLEID	SNP1	SNP2	SNP3
ANC30000	GT	TT	СТ

- III. Pick best three population for blind sample
- IV. Get proportions which has maximum likelihood value
  - a. Pick three best populations

## African(Assume blind sample 100% African):

SNP1 Alleles: G, T

"G" allele frequency in African P(G) = 0.8 "T" allele frequency in African P(T) = 0.2

Expected genotype value for SNP1 = log (2 * P(G) * P(T))= log (2 * 0.8 * 0.2)

= -0.4948

SNP2 Alleles: T, T

"T" allele frequency in African P(T) = 0.7

Expected genotype value for SNP2 = log(P(T) * P(T))

 $= \log (0.7 * 0.7)$ = -0.3098

SNP3 Alleles: C, T

"C" allele frequency in African P(C) = 0.9999 "T" allele frequency in African P(T) = 0.0001

Expected genotype value for SNP3 =  $\log (2 * P(C) * P(T))$ 

 $= \log(2*0.9999*0.0001)$ = -3.6990

Likelihood for African = -0.4948 - 0.3098 - 3.6990 - 4.5036

166

# European(Assume blind sample 100% European):

SNP1 Alleles: G, T

"G" allele frequency in European P(G) = 0.9"T" allele frequency in European P(T) = 0.1

Expected genotype value for SNP1 = log (2 * P(G) * P(T))= log (2 * 0.9 * 0.1)= -0.7447

SNP2 Alleles: T, T

"T" allele frequency in European P(T) = 0.7

Expected genotype value for SNP2 = log(P(T) * P(T))= log(0.7 * 0.7)

= -0.3098

SNP3 Alleles: C, T

"C" allele frequency in European P(C) = 0.8 "T" allele frequency in European P(T) = 0.2

Expected genotype value for SNP3 = log (2 * P(C) * P(T))= log (2 * 0.8 * 0.2)= -0.4948

2. Likelihood for European = -0.7447 - 0.3098 - 0.4948

= -1.5493

## Native American(Assume blind sample 100% NA):

SNP1 Alleles: G, T

"G" allele frequency in NA P(G) = 0.6 "T" allele frequency in NA P(T) = 0.4

Expected genotype value for SNP1 = log (2 * P(G) * P(T))= log (2 * 0.6 * 0.4)

- 0.3187

SNP2 Alleles: T, T

WO 2004/016768

167

"T" allele frequency in NA P(T) = 0.5Expected genotype value for SNP2 = log(P(T) * P(T)) $\log (0.5 * 0.5)$ - 0.6020

SNP3 Alleles: C, T

"C" allele frequency in NA P(C) = 0.7"T" allele frequency in NA P(T) = 0.3

Expected genotype value for SNP3  $\log (2 * P(C) * P(T))$ log(2*0.7*0.3)- 0.3767

Likelihood for Native American = -0.3187 - 0.6020 -

0.3767

= -1.2974

## Middle East(Assume blind sample 100% ME):

SNP1 Alleles: G, T

"G" allele frequency in ME P(G) = 0.7"T" allele frequency in ME P(T) = 0.3

Expected genotype value for SNP1 =  $\log (2 * P(G) * P(T))$  $\log (2 * 0.7 * 0.3)$ 

- 0.3767

SNP2 Alleles: T, T

"T" allele frequency in ME P(T) = 0.9

Expected genotype value for SNP2 log(P(T) * P(T))

log(0.9*0.9)- 0.0915

SNP3 Alleles: C, T

"C" allele frequency in ME P(C) = 0.9"T" allele frequency in ME P(T) = 0.1

Expected genotype value for SNP3 log (2 * P(C) * P(T))

log (2*0.9 * 0.1)

- 0.7447

168

- 5. Likelihood value for European = -1.5493
- 6. Likelihood value for Native American = -1.2974
- 7. Likelihood value for Middle East = -1.2129

In this case we drop African and consider other three for proportions.

## Maximum likelihood value

Now starts giving some values to unknown parameters

```
I = European J= Native American K = Middle East
Always
              I + j + k = 1
I = 0; j = 0; k=1
{
       SNP1 Alleles: G, T.
       "G" allele frequency in European
                                           P(G,1) = 0.9
       "G" allele frequency in NA
                                           P(G,2) = 0.6
       "G" allele frequency in ME
                                           P(G,3) = 0.7
       Allele1 Estimated Frequency(A1EF) = I * P(G,1) + J*P(G,2) + K* P(G,3)
                                           = 0^{\circ} 0.9 + 0 = 0.6 + 1 = 0.7
                                           = 0.7
       "T" allele frequency in European
                                           P(T,1) = 0.1
       "T" allele frequency in NA
                                           P(T,2) = 0.4
       "T" allele frequency in ME
                                           P(T,3) = 0.3
       Allele 2 Estimated Frequency(A1EF) = I * P(T,1) + J*P(T,2) + K* P(T,3)
                                           = 0 * 0.1 + 0*0.4 + 1*0.3
```

= 0.3

169

Expected genotype value for SNP1 = log (2*A1EF*A2EF) = log (2*0.7*0.3); = - 0.3767

# SNP2 Alleles: T, T.

"T" allele frequency in European P(T,1) = 0.7 "T" allele frequency in NA P(T,2) = 0.5 "T" allele frequency in ME P(T,3) = 0.9

Allele 1 Estimated Frequency(A1EF) = I * P(T,1) + J*P(T,2) + K* P(T,3)= 0 * 0.7 + 0*0.5 + 1*0.9= 0.9

Expected genotype value for SNP2 (EGV2) = log(A1EF*A2EF) = log (0.9*0.9); = - 0.0915

### SNP3 Alleles: C, T.

"C" allele frequency in European P(C, 1) = 0.8 "C" allele frequency in NA P(C, 2) = 0.7 "C" allele frequency in ME P(C, 3) = 0.9

Allele 1 Estimated Frequency(A1EF) = I * P(C,1) + J*P(C,2) + K* P(C,3)= 0 * 0.8 + 0*0.7 + 1*0.9= 0.9

"T" allele frequency in European P(T,1) = 0.2 "T" allele frequency in NA P(T,2) = 0.3 "T" allele frequency in ME P(T,3) = 0.1

Allele 2 Estimated Frequency(A1EF) = I * P(T,1) + J*P(T,2) + K* P(T,3)= 0 * 0.2 + 0*0.3 + 1*0.1= 0.1

Expected genotype value for SNP3(EGV3) = log (2*A1EF*A2EF)= log (2*0.9*0.1); = -0.7447

i. Likelihood value for unknown parameters

= EGV1+EGV2+EGV3 = -0.3767 - 0.0915 - 0.7447 }

170

= -1.2129

for European = 0; NA=0; MiddleEast =1; likelihood value is -1.2129

Repeat above loop for all possible combinations

0.0,0.0,1.0 -1.2129 1.0, 0.0, 0.0, 0.0, 1.0, 0.0, 0.1, 0.0 0.9, 0.1, 0.1,0.8, 0.1,0.2, 0.7, 0.1,0.3,0.6,

etc

0.1,0.4, 0.5

Get **Maximum likelihood** value and corresponding proportions are ancestry proportions.

[0333] Although the invention has been described with reference to the above example, it will be understood that modifications and variations are encompassed within the spirit and scope of the invention. Accordingly, the invention is limited only by the following claims.

#### What is claimed is:

- 1. A method of inferring, with a predetermined level of confidence, a trait of an individual, comprising:
  - a) contacting a sample comprising nucleic acid molecules of a test individual with hybridizing oligonucleotides,

wherein the hybridizing nucleotides can detect nucleotide occurrences of single nucleotide polymorphisms (SNPs) of a panel of at least about ten ancestry informative markers (AIMs) indicative of a population structure correlated with the trait, and

wherein said contacting is performed under conditions suitable for detecting the nucleotide occurrences of the AIMs of the test individual by the hybridizing oligonucleotides; and

b) identifying, with a predetermined level of confidence, a population structure that correlates with the nucleotide occurrences of the AIMs in the test individual, wherein the population structure correlates with a trait,

thereby inferring, with a predetermined level of confidence, the trait of the individual.

- 2. The method of claim 1, wherein the panel comprises at least about twenty AIMs.
- 3. The method of claim 1, wherein the trait comprises biogeographical ancestry (BGA).
- 4. The method of claim 3, wherein the panel comprises AIMs set forth as SEQ ID NOS:1 to 331.
- 5. The method of claim 3, wherein the panel comprises AIMs set forth as SEQ ID NOS:1 to 71.

WO 2004/016768

PCT/US2003/026229

172

- 6. The method of claim 3, wherein the panel comprises AIMs set forth as: SEQ ID NOS:7, 21, 23, 27, 45, 54, 59, 63, and 72 to 152; SEQ ID NOS:3, 8, 9, 11, 12, 33, 40, 59, 63, and 153 to 239; SEQ ID NOS:1, 8, 11, 21, 24, 40, 172, and 240 to 331; or combinations thereof.
- 7. The method of claim 1, wherein at least one AIM of the panel is not linked to a gene linked to the trait.
- 8. The method of claim 3, wherein the BGA comprises a proportion of a sub-Saharan African, Native American, IndoEuropean, or East Asian ancestral group, or a combination of said ancestral groups.
- 9. The method of claim 8, wherein the BGA comprises a proportion of at least three ancestral groups.
- 10. The method of claim 3, wherein the BGA comprises proportions of at least sub-Saharan African and IndoEuropean ancestral groups; Native American and IndoEuropean ancestral groups; East Asian and Native American ancestral groups; or IndoEuropean and East Asian ancestral groups.
- 11. The method of claim 3, wherein the BGA comprises proportions of at least Native American, East Asian, and IndoEuropean ancestral groups; or sub-Saharan African, Native American, and IndoEuropean ancestral groups.
- 12. The method of claim 1, wherein the trait comprises responsiveness of the individual to a drug.
  - 13. The method of claim 12, wherein the drug is a cancer chemotherapeutic agent.

173

- 14. The method of claim 12, wherein the drug is a statin.
- 15. The method of claim 1, wherein the trait comprises susceptibility to a disease.
- 16. The method of claim 15, wherein the disease has an ethnic predisposition.
- 17. The method of claim 16, wherein the disease is a cancer, diabetes, or hypertension.
  - 18. The method of claim 17, wherein the cancer is prostate cancer.
  - 19. The method of claim 15, wherein the disease is a neurological disorder.
  - 20. The method of claim 19, wherein the schizophrenia or Parkinson's disease.
  - 21. The method of claim 15, wherein the disease is alcoholism.
  - 22. The method of claim 1, wherein the trait comprises a pigmentation trait.
- 23. The method of claim 22, wherein the pigmentation trait comprises eye color, skin color, hair color, or a combination thereof.
- 24. The method of claim 1, further comprising identifying, with a predetermined level of confidence, a sub-population structure of the population structure that correlates with the nucleotide occurrences of the AIMs in the test individual, wherein the sub-population structure correlates with a trait.

- 25. The method of claim 1, wherein the hybridizing oligonucleotides comprise oligonucleotide primers, said method further comprising contacting the sample with a polymerase, under condition suitable for generation of a primer extension product, wherein determining the nucleotide occurrence of a SNP comprises detecting the presence of the primer extension product.
- 26. The method of claim 1, wherein the hybridizing oligonucleotides comprise oligonucleotide primers, said method further comprising contacting the sample with a polymerase, under condition suitable for generation of a primer extension product, wherein determining the nucleotide occurrence of a SNP comprises determining the nucleotide sequence of the primer extension product at a position corresponding to the position of the SNP.
- 27. The method of claim 1, wherein the hybridizing oligonucleotides comprise amplification primer pairs, said method further comprising contacting the sample with a polymerase, under condition suitable for generation of an amplification product, wherein determining the nucleotide occurrence of a SNP comprises detecting the presence of the amplification product.
- 28. The method of claim 1, wherein the hybridizing oligonucleotides comprise amplification primer pairs, said method further comprising contacting the sample with a polymerase, under condition suitable for generation of an amplification product, wherein determining the nucleotide occurrence of a SNP comprises determining the nucleotide sequence of the amplification product at a position corresponding to the position of the SNP.
  - 29. The method of claim 1, which is performed in a high throughput format.
  - 30. The method of claim 1, which is performed in a multiplex format.

- 31. A method of estimating, with a predetermined level of confidence, proportional ancestry of at least two ancestral groups of an individual, comprising:
  - a) contacting a sample comprising nucleic acid molecules of a test individual with hybridizing oligonucleotides,

wherein the hybridizing oligonucleotides can detect nucleotide occurrences of single nucleotide polymorphisms (SNPs) of a panel of at least about ten ancestry informative markers (AIMs) indicative of biogeographical ancestry (BGA) for each ancestral group examined, and

wherein said contacting is under conditions suitable for detecting the nucleotide occurrences of the AIMs of the test individual by the hybridizing oligonucleotides; and

b) identifying, with a predetermined level of confidence, a population structure that correlates with the nucleotide occurrences of the AIMs of each of the ancestral groups examined, wherein the population structure is indicative of proportional ancestry,

thereby estimating, with a predetermined level of confidence, proportional ancestry of the individual.

- 32. The method of claim 31, wherein the proportional ancestry comprises a proportion of a sub-Saharan African ancestral group, a Native American ancestral group, an IndoEuropean ancestral group, an East Asian ancestral group, or a combination thereof.
- 33. The method of claim 31, wherein the proportional ancestry comprises proportions of sub-Saharan African and IndoEuropean ancestral groups; Native American and IndoEuropean ancestral groups; East Asian and Native American ancestral groups; or IndoEuropean and East Asian ancestral groups.

- 34. The method of claim 31, wherein the proportional ancestry comprises proportions of Native American, East Asian, and IndoEuropean ancestral groups; sub-Saharan African, Native American, and IndoEuropean ancestral groups; or sub-Saharan African, Native American, and East Asian ancestral groups.
- 35. The method of claim 31, wherein the panel for at least one of the ancestral groups comprises AIMs as set forth in SEQ ID NOS:1 to 331.
- 36. The method of claim 31, wherein the panel for at least one of the ancestral groups comprises AIMs set forth as SEQ ID NOS:1 to 71.
- 37. The method of claim 31, wherein the panel for at least one of the ancestral groups comprises AIMs set forth as:

SEQ ID NOS:7, 21, 23, 27, 45, 54, 59, 63, and 72 to 152; SEQ ID NOS:3, 8, 9, 11, 12, 33, 40, 59, 63, and 153 to 239; SEQ ID NOS:1, 8, 11, 21, 24, 40, 172, and 240 to 331; or combinations thereof.

- 38. The method of claim 31, wherein at least one AIM of the panel is not linked to a gene linked to the trait.
- 39. The method of claim 31, wherein the proportional ancestry comprises proportions of three ancestral groups, and wherein identifying a population structure that correlates with the nucleotide occurrences of the AIMs of the test individual comprises:

performing a likelihood determination for affiliation with each of a sub-Saharan African ancestral group, a Native American ancestral group, an IndoEuropean ancestral group, and an East Asian ancestral group;

thereafter selecting three ancestral groups having a greatest likelihood value;

determining a likelihood of all possible proportional affiliations among the three ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with the nucleotide occurrences of the AIMs of the test individual is identified; and

identifying a single proportional combination of maximum likelihood, thereby estimating the proportional ancestry of the individual.

40. The method of claim 31, wherein the proportional ancestry comprises proportions of three ancestral groups, and wherein identifying a population structure that correlates with the nucleotide occurrences of the AIMs comprises:

performing six two-way comparisons comprising likelihood determinations for affiliation between each group with each other group;

thereafter selecting three ancestral groups having a greatest likelihood value;

determining a likelihood of all possible proportional affiliations among the three ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with the nucleotide occurrences of the AIMs of the test individual is identified; and

identifying a single proportional combination of maximum likelihood, thereby estimating the proportional ancestry of the individual.

41. The method of claim 31, wherein the proportional ancestry comprises proportions of three ancestral groups, and

wherein identifying a population structure that correlates with the nucleotide occurrences of the AIMs of the test individual comprises performing three three-way comparisons among the groups,

determining a likelihood of all possible proportional affiliations among the three ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with the nucleotide occurrences of the AIMs of the test individual is identified; and

identifying a single proportional combination of maximum likelihood, thereby estimating the proportional ancestry of the individual.

42. The method of claim 31, wherein the proportional ancestry comprises proportions of four ancestral groups, and

wherein identifying a population structure that correlates with the nucleotide occurrences of the AIMs of the test individual comprises performing six two-way comparisons, three three-way comparisons, or one four-way comparison among the groups;

determining a likelihood of all possible proportional affiliations among the four ancestral groups having the greatest likelihood value, whereby a population structure or proportional affiliation that correlates with the nucleotide occurrences of the AIMs of the test individual is identified; and

identifying a single proportional combination of maximum likelihood, thereby estimating the proportional ancestry of the individual.

- 43. The method of claim 40, further comprising generating a graphical representation of the comparison of the three ancestral groups, said graphical representation comprising a triangle with each an ancestral group independently represented by a vertex of the triangle, wherein the maximum likelihood value of proportional affiliation for an individual comprises a point within the triangle.
- 44. The method of claim 43, wherein the graphical representation further comprises a confidence contour indicating a level of confidence associated with estimating the proportional ancestry.
- 45. The method of claim 31, further comprising identifying, with a predetermined level of confidence, a sub-population structure of the population structure that correlates with the nucleotide occurrences of the AIMs in the test individual, wherein the sub-population structure correlates with ethnicity of the test individual.

46. The method of claim 45, wherein identifying the subpopulation structure comprises

identifying chromosomes of the test individual that contain the AIMs indicative of an ancestral group of the proportional ancestry of the test individual,

contacting a sample comprising nucleic acid molecules of the test individual with second hybridizing oligonucleotides;

wherein the second hybridizing oligonucleotides can detect nucleotide occurrences of SNPs of a second panel of AIMs, and

wherein the AIMs of the second panel are present on the chromosomes of the test individual that contain the AIMs indicative of the ancestral group of the test individual; and

identifying a sub-population structure that correlates with the nucleotide occurrences of the AIMs of the second panel, wherein the sub-population is indicative of ethnicity of the ancestral group of the test individual.

- 47. The method of claim 45, wherein the ancestral group is IndoEuropean, and wherein the ethnicity comprises Northern European or Mediterranean.
- 48. The method of claim 31, further comprising generating an ancestral map of the world, wherein locations of populations having a proportional ancestry corresponding to the proportional ancestry of the test individual are indicated on the ancestral map.
  - 49. The method of claim 48, further comprising
  - a) overlaying the ancestral map with a genealogical map, wherein the genealogical map indicates locations of populations having geopolitical relevance with respect to the test individual, and
  - b) statistically combining the information of the ancestral map and genealogical map to obtain a most likely estimate of family history of the test individual.

- 50. The method of claim 31, wherein identifying a population structure that correlates with the nucleotide occurrences of the AIMs comprises comparing the nucleotide occurrences of the AIMs of the test individual with known proportional ancestries corresponding to nucleotide occurrences of AIMs indicative of BGA.
- 51. The method of claim 50, wherein the known proportional ancestries corresponding to nucleotide occurrences of AIMs indicative of BGA are contained in a database.
  - 52. The method of claim 51, wherein the comparing is performed using a computer.
- 53. The method of claim 50, wherein each of the known proportional ancestries corresponding to nucleotide occurrences of AIMs indicative of BGA further comprises of photograph of a person from whom the known proportional ancestry was determined.
  - 54. The method of claim 53, wherein the photograph comprises a digital photograph.
- 55. The method of claim 54, wherein digital information comprising the digital photograph is contained in a database.
- 56. The method of claim 55, wherein the digital information in the database is associated with a known proportional ancestry corresponding to nucleotide occurrences of AIMs indicative of BGA of the person in the photograph.
- 57. The method of claim 51, further comprising identifying a photograph of a person having a proportional ancestry corresponding to the proportional ancestry of the test individual.

- 58. The method of claim 57, wherein identifying the photograph comprises scanning a database comprising a plurality of files, each file comprising digital information corresponding to a digital photograph of a person having a known proportional ancestry, and identifying at least one photograph of a person having nucleotide occurrences of AIMs indicative of BGA that correspond to the nucleotide occurrences of AIMs indicative of BGA of the test individual.
- 59. An article of manufacture, comprising at least one photograph of a person having a known proportional ancestry corresponding to a population structure comprising nucleotide occurrences of AIMs indicative of biogeographical ancestry (BGA).
  - 60. The article of claim 59, which is contained in a file.
- 61. A plurality of files comprising the article of manufacture of claim 59, wherein files of the plurality comprise at least one photograph of a person having a known proportional ancestry corresponding to a population structure comprising nucleotide occurrences of AIMs indicative of BGA.
- 62. The file of claim 60, which comprises a plurality of photographs, wherein photographs of the plurality comprise a photograph of a person having a known proportional ancestry corresponding to a population structure comprising nucleotide occurrences of AIMs indicative of BGA.
- 63. The file of claim 62, wherein photographs of the plurality comprise photographs of different persons having the same known proportional ancestries.
- 64. The file of claim 62, wherein photographs of the plurality comprise photographs of different persons having different known proportional ancestries.

182

- 65. The article of manufacture of claim 59, wherein the at least one photograph comprises a digital photograph.
- 66. The article of manufacture of claim 65, wherein the digital photograph comprises digital information.
- 67. The article of manufacture of claim 66, wherein the digital information is contained in a database.
- 68. The article of manufacture of claim 65, comprising a plurality of digital photographs.
- 69. A plurality of the articles of manufactures of claim 65, comprising at least two digital photographs.
- 70. The plurality of claim 69, wherein the digital photographs comprise digital information.
- 71. The plurality of claim 70, wherein the digital information is contained in a database.
- 72. A kit, comprising a plurality of hybridizing oligonucleotides, which comprise at least fifteen contiguous nucleotides of at least five polynucleotides as set forth in SEQ ID NOS:1 to 331, or polynucleotides complementary thereto.
- 73. The kit of claim 72, wherein the hybridizing oligonucleotides comprise at least fifteen contiguous nucleotides of at least five polynucleotides as set forth in SEQ ID NOS:1 to 71, or polynucleotides complementary thereto.

- 74. The kit of claim 72, wherein hybridizing oligonucleotides of the plurality comprise at least one nucleotide corresponding to a position of the polymorphism of the polynucleotide, or polynucleotide complementary thereto.
- 75. The kit of claim 74, wherein hybridizing oligonucleotides of the plurality comprise nucleotide position 50 of a polynucleotide as set forth in any of SEQ ID NOS:1 to 34, 36 to 49, 52 to 55, or 57 to 98, 100 to 105, 107 to 162, 164 to 331, or a polynucleotide complementary thereto.
- 76. The kit of claim 72, wherein hybridizing oligonucleotides of the plurality comprise at least one probe, at least one primer, or a combination thereof.
  - 77. The kit of claim 76, comprising at least one amplification primer.
- 78. The kit of claim 76, comprising at least one amplification primer pair, which comprises a forward primer and a reverse primer.
- 79. The kit of claim 78, further comprising reagents for performing an amplification reaction using the at least one amplification primer pair.
- 80. The kit of claim 72, further comprising at least one ancestry informative marker (AIM), wherein the AIM corresponds to a hybridizing oligonucleotide of the plurality.
- 81. The kit of claim 72, further comprising a detectable label, which can be bound to or incorporated into at least one hybridizing oligonucleotide of the plurality.
- 82. The kit of claim 72, wherein hybridizing oligonucleotides of the plurality are detectably labeled.

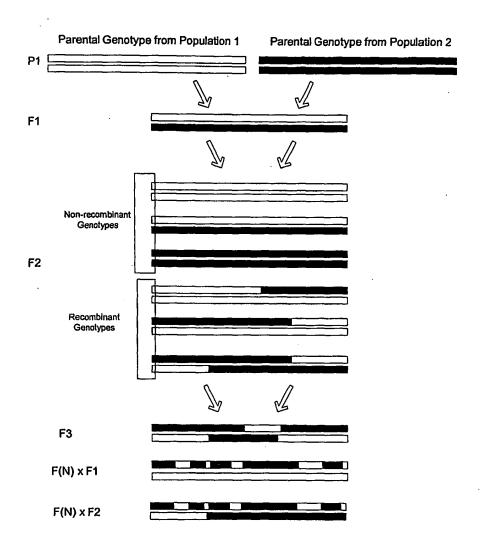


FIGURE 1

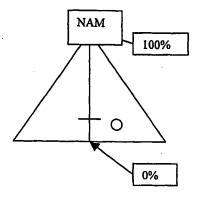


FIGURE 2A

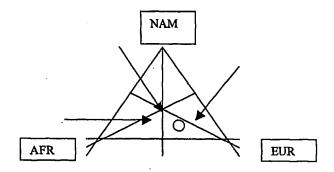


FIGURE 2B

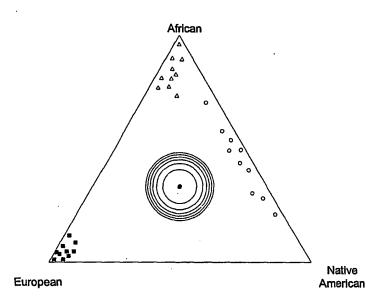


FIGURE 3

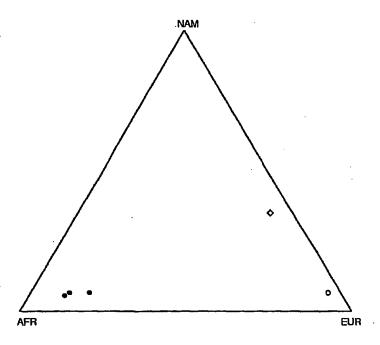


FIGURE 4

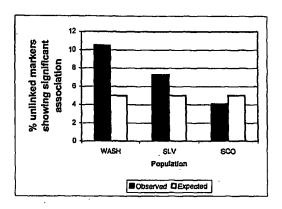


FIGURE 5A

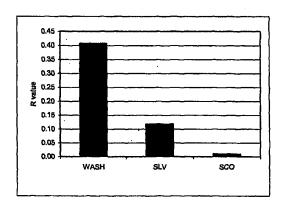


FIGURE 5B

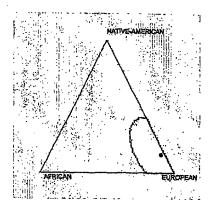


FIGURE 6A

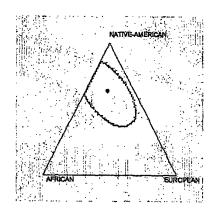
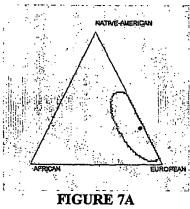
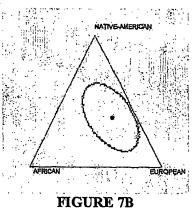


FIGURE 6B





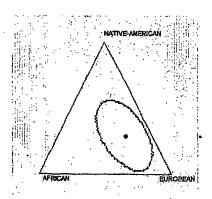


FIGURE 7C

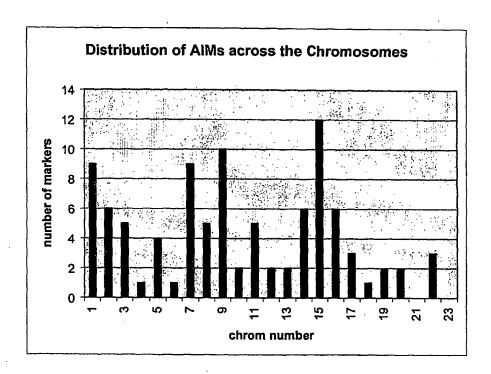


FIGURE 8

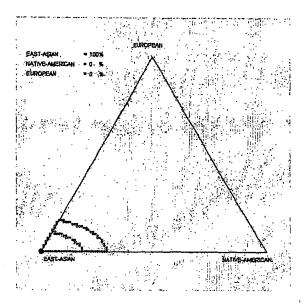


FIGURE 9A

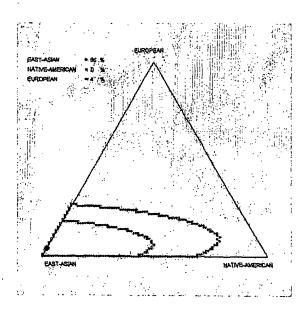
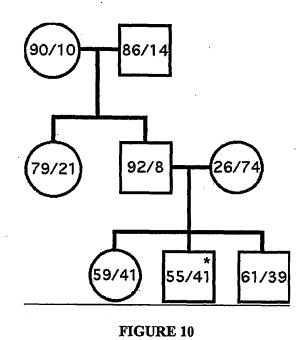


FIGURE 9B



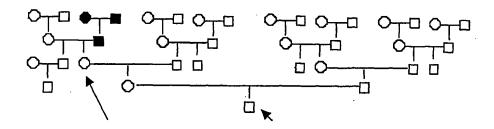


FIGURE 11

10/11

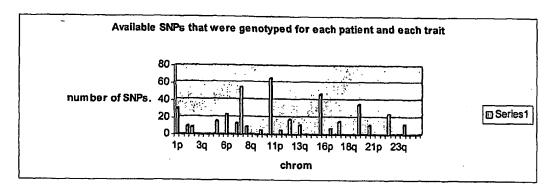


FIGURE 12

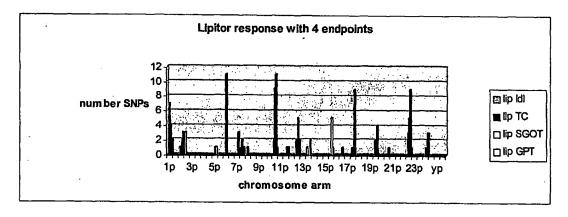


FIGURE 13

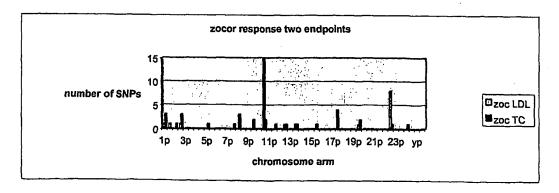


FIGURE 14

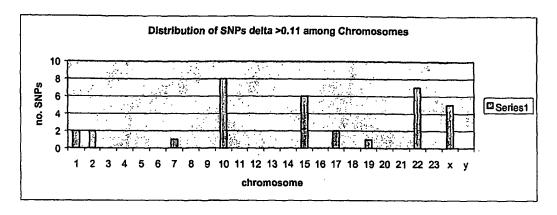


FIGURE 15

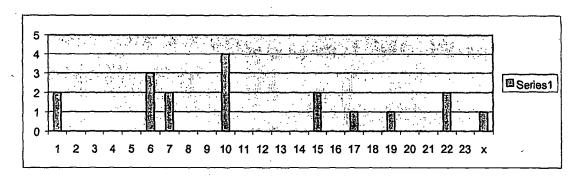


FIGURE 16

## SEQUENCE LISTING

<pre>&lt;110&gt; DNA Print Genomics, Inc. FRUDAKIS, Tony N. SHRIVER, Mark D.</pre>	
<120> COMPOSITIONS AND METHODS FOR INFERRING ANCESTRY	
<130> DNA1170-2WO	
<150> US 60/404,357 <151> 2002-08-19	
<150> US 60/467,613 <151> 2003-05-02	
<160> 363	
<170> PatentIn version 3.1	
<210> 1 <211> 250 <212> DNA <213> Homo sapiens rs236919	
<400> 1 tgagcgtgga ggtggagtgg gaagatgggt caacttgcag cagttctccy gggactg	tat 60
gaagagggca tecegittet cagagtetti gigeaggeea gitaeettie caagetg	
ggggccatcc actgtcttcc catcgtcatc tggtccccag ctgttgagaa ataaata	
agaggggaca gecagageca ggetgegget acceaeceag egecteecag teteage	
ctgccctttg	250
<210> 2 <211> 170 <212> DNA <213> Homo sapiens rs594689	
<400> 2 gccgggcaga gacagcggct agagaccccg aggtggggga cacaaatgar ctccttg	gaac 60
gtcccccaa tttctcctac cctacaaagg gagggaatcc caaaaggacc cttctac	
tgtgactggt tttcatgaag gactatgatt cccatcatcc ctcaggggca	170
<210> 3 <211> 189 <212> DNA <213> Homo sapiens rs725192	
<400> 3 caaatccctg aaagagtaag cattatgcta atgggtctga aaatataaay gaagaa	tcaa 60
aatctattct gctttaaaga aggcaccagt ttaggaggca ggctctaagt acttga	
ataaaaggaa ctcccctttg cctaacaagt cccatatagc agcctccttc atacat	

tttaacgtc	189
<210> 4 <211> 110 <212> DNA <213> Homo sapiens rs1800498	
<400> 4 aggaatgatg cctggatgcc tgggggtgtg aagaaaagag ccttgggtty gactagggaa	60
cetggggeca etectteete tgetaetaaa teaceaagtg atettgttet	110
<210> 5 <211> 170 <212> DNA <213> Homo sapiens rs593226	
<400> 5 cttccttcta tgacccctac ctgttgtggg tctccaggca agcagtgcay aggtagagcc	60
atccttaggt agcctttaga cttaatatta ggtgagctct ccccacagat agcctctcct	120
	170
ttatttgaat ggaattatat tttaagtttg gaaatatttt tcagcttatt	170
<210> 6 <211> 170 <212> DNA <213> Homo sapiens rs1337038	
<400> 6 ctctcaaaca ttagetttta cccttttacc ttcaggttcg taccccaccr tatttgcata	60
attgtattac ttccctaaga accaattttt tgttgttgtt gtttgctagt acttttgtgc	120
cacctgacaa acttacaagc tatttttaaa tgcttgcttt taaatagagg	170
<210> 7 <211> 126	
<212> DNA	
<213> Homo sapiens rs729531	
<400> 7 ccagtaggtg tcagctacgc acagcettte tetttetaaa teeatatttr taagaaaacg	60
geettteate cataatttte cetagttaaa gggateaatt tettetgate tgaageaege	120
aaaaca	126
<210> 8 <211> 170 <212> DNA <213> Homo sapiens rsllll108	
<400> 8 gaaacgccat ggctgttctc ataccaagtg ctatttaata agacagatcr ctcaaactgg	60

PCT/US2003/026229

gatcccaaga gggttcaatt aagagcataa tgtaattcca ggctaccagt gctgccctca	120
gcccaagtat ctcatacctg ctctatgaag tctgtcctga tgtccccaga	170
<210> 9 <211> 170 <212> DNA <213> Homo sapiens rs1007407	
<400> 9	
tggcctgtga agtccagaag ctctttccgc gctcggctct ttgagcctcr gtttccccat	60
ctgtcaggcg tgtgtatgtg ggagggggt acggagtaga tttacctaga tcactcgggg	120
ttgtcccttc cagttttgag ggtgatgtcg gggtgtctgt gaggaggagg	170
<210> 10 <211> 170 <212> DNA <213> Homo sapiens rs741272	
<400> 10	
gtcgctgtca gttgaaatag attacaagag ggatctcctg gcagaatttr tttagtttag	60
ctcaaaggag aatgttatag tcaagggtac atttggttgc aaggacaaga gagtcactcg	120
agttggctta agtaaaaggg tacttgtccc aaaaacataa cagccagtcc	170
<210> 11 <211> 170 <212> DNA <213> Homo sapiens rs1076808	
<400> 11 aatototttt coagaagtot aagtgaaaat otoactttgt ototgtactr gttaggoaag	60
gctaagttgc tatgacaaat agacctcaaa ttctgctgta acagctcaga ctataacact	120
gcattttgct gtcatgtaat atcagtggtg ggcctttctg gtcatttctc	170
<210> 12 <211> 170 <212> DNA <213> Homo sapiens rs716873	
<400> 12	
etecatgetg catgtaggtt tataaaccaa taccetgete agacaccaer ttettetaac	60
accetaatga tggggtcaca tttctctctt taggtactac aatctggttt tgaagttgcc	120
caaagaaact tgaatggett cetettteat teecacaagg etetgaaaac	170
<210> 13	

<212> DNA

WO 2004/016768 PCT/US2003/026229 4/68

<213> Hom	o sapiens	rs730570				
<400> 13 gacactcccc	tactccaggo	ctgcagcact	cacctgcatc	tcacactgcr	atgcaaaatt	6
tgtgtgatta	atgggttcaa	atgtgcccc	ctcccctgga	·ctttacaggg	tgctggagct	120
gctggcttgc	tgttcccagt	gccaagtccc	atatgggtgc	tcagccatat		170
<210> 14 <211> 311 <212> DNA <213> Hom		886993 (OC	A2E10_102)			
<400> 14 ttaatatggt	gtcctgctaa	aagattatcc	ttgtcttctt	cttttccccr	tagatgatct	60
tagtagccat	attttcagaa	acgggatttt	tcgattattg	tgctgtaaag	gtaggtatga	120
tgttgcattt	aataattcta	tcctgattaa	tttatatatg	tatttttctg	acattatata	180
tttaggaaac	aaacatttaa	aacaaacatt	gaaaaattcc	atccttcttt	taaaggttgc	240
ttetgeaġgg	cagggtatac	ttgctatgtt	aagttgtatg	gctctgagca	gcactttcag	300
ctgctcagta	a					311
<210> 15 <211> 242 <212> DNA <213> Home	o sapiens	886895 (OC)	A2E11_263)		·	
<400> 15 cctcttcacg	cctgtgacca	taaggtacgc	aaagcacctc	tgccgtgggr	gttgcggcca	60
ggttctggca	ggcaggggct	ctgcctgcac	tgcctggctc	caggttccat	tctcaggtgc	120
atgaaaaggt	gggggcrgtt	gagcccacag	ctcactgcat	tccagtccag	ctcgtgtctg	180
ctttgtgtga	ctgcagtaca	tgctacaagc	agtggggcct	cagaagctgg	tggcagaaat	240
gc						242
<210> 16 <211> 110 <212> DNA <213> Homo	o sapiens :	rs1426208				
<400> 16	accettetet	tactttaaaa	acaccataga	tataataa	tantangaa	60
	tagttctgca				Lyaccaaccc	60 110
<210> 17 <211> 95 <212> DNA		·				

<213> Homo sapiens rs2862

<400> 17	
totgagtgca agataaaaag gaaatagcaa ttoaaggcac aaagctaagy acatgcaaca	60
atatagatga tttgggggtg ggacagtaca gaatt	95
<210> 18	
<211> 110 <212> DNA	
<212> DNA <213> Homo sapiens rs972801	
1415 Homo Saptems 18972801	
<400> 18	
ttcttctgaa tctagatgca gcatttagca actctatact attcaccaay attatcttgc	60
tttatgtttt caacctataa aattcaactc agtttctgaa aaaggtgtag	. 110
2	110
<210> 19	
<211> 110	
<212> DNA	
<213> Homo sapiens rs251741	
<400> 19	
gaatctgctg agactgacgc tggcgtccca accagacact tggaggcatr gggcatgccc	60
	80
ttcaagtcta gaaccctgcc aggtacctat ccctcctgcc caggccaaag	110
<210> 20	
<211> 73	
<212> DNA	
<213> Homo sapiens rs1650999	
<400> 20	
catatggtgc aataaaattg tggtacagca aagacaaaga gaaaaaccay agaaagaacc	60
gcagtaaaaa cat	77
JJ	73
-210. 23	
<210> 21 <211> 110	
<212> DNA	
<213> Homo sapiens rs173537	
<400> 21	
ccctcccctc ttgggtcttg tagtcggctg tgggtggcaa gtacagcagy gatggcagac	60
aggtgtgatg agttcaagga cagggaaata acatccacag ccccatccca	110
<210> 22	
<211> 64	
<212> DNA	
<213> Homo sapiens rs67302	
<400> 22	
tgtcacctcg gatcacccgg gaacgtagac tccaaactcc caatggctty agcagaaatt	60
gage	. 64

<210>	23				
<211>	110				
<212>	DNA				
<213>	Homo sapiens rs2124	98	•		
<400>	23 .				
gggtgaa	itgg atggatgagt gctgg	gagge tgtati	gtaa ataacaaa	ar toggatetga	60
			-	020 0	
aggaagg	ıtat gggactetga agtet	gcatt ccttt	acct tttgctat	at	, 110
				- <del></del>	,
<210>	24				
<211>	110				
<212>	DNA				
<213>	Homo sapiens rs1010	022	•		•
10107	nome sapiens raidio	923			
<400>	24				
		<b></b>			
gacaaac	tga attaaacgtg gtatt	ctart tggatt	cctt gattttta	ay gtaagcatga	, 60
		<b>.</b>			
accacac	taa ctcttccttg ctato	rgaaa tactgo	aatt cttgttct	ag	110
010	0.5	•			
<210>	25 .				•
<211>	110				
	DNA				
<213>	Homo sapiens rs4306	67			
	•				
<400>	25		•		
gcgctga	tgt ggttgtggag gtcac	tgtgg agacca	caga ggggtagt	tr tcatgtccag	60
	3			• •	
gtcccaa	ctg ttaagcatga aggga	agtat agtttt	aaag gcttttgt	qt	110
			<del>"</del>		
<210>	26				
<211>	110				
<212>	DNA				
<213>	Homo sapiens rs2363	36	•		
	•				
<400>	26				
	ccc tgtgggcccc cagaa	ataac totcac	agge atgaetge	er gaggagatag	60
J . J . J .	· · · · · · · · · · · · · · · · · · ·		agge acgaecgg	cr ggggagacgg	00
acaaaaa	tcc tgccattatg aagtg	dotto caadao	teae caeaette	30	110
34333	ere egocarcacy aageg	ggeec caagag	cagageeg	ay	110
<210>	27				
	110				
	DNA	•			
		61			
~~137	Homo sapiens rs7830	0-2			
<400>	27 .	•			
-					
cactyct	ctg tcttggagag ggttc	aytge tgggtg	yyga gacatctg	ar aacagattat	60
+0000++	224 ta242222			·	
Lycaact	aag tgagaaaagc tgaaa	yıtaa attgoo	taga gtgtgcat	tc	110
-010:			•		•
	28				
	110				
<212>	DNA			-	
-917-	TT				

28					
gget tttgeteece	tacccacctg	actctactga	ggctcacctr	ggcgccccct	60
atcc ccagaaccgg	gccattgcca	cacccaatca	gggtgtctgg		110
	•		•		
29					
110					•
DNA			•		
Homo sapiens	rs434504				
etet etggetgeat	agaggtttta	atgaaacaag	gggataaccr	teteecteet	60
ota ttancactac	cactegettt	tacaaaaaaa	aasaataa		110
cty ttyatattyt	cagegggeee	cgcaggcaga	gcaccctgec		110
		•			
30	•	•	,		•
110					
DNA .					-
Homo sapiens	rs2814778 (I	FY-null)			
30					
agac agaagggctg	ggacggctgt	cagcgcctgt	gcttccaagr	taagagccaa	60
atga gggccatcag	gttctgggca	gggacagggg	gcatcaggaa		110
21					
		-			•
	r=1325609			•	
nomo sapiens	181323003				
31					
agag tgctgagaga	cgagggatga	tggctccact	gttgtgctgs	ccacattgta	60
gagt ccaggctcct	gctcttatca	gaacagcaat	ggtagctgct		110
		•			
	•				
	~-6003				
HOMO Saprens	186003				
32	•	•			
	gttattgtat	aaaattcaag	agaacatgcr	ttatggttgc	60
	3		<b></b>	0000330030	
ggt acaaaaccac	tggagggaag	gatgaagaag	tggttcaatg		110
	-0 000 0		30		
33					
110	•				
DNA					
DNA Homo sapiens	rs2065160				
Homo sapiens	rs2065160				
Homo sapiens					ـ ـ ـ
Homo sapiens		ctcgatgagt	aaatatgggy	caaatgaagt	60
	29 110 DNA Homo sapiens 29 etct ctggctgcat cctg ttgacactgc 30 110 DNA Homo sapiens 30 agac agaagggctg atga gggccatcag 31 110 DNA Homo sapiens 31 agag tgctgagaga gagt ccaggctcct 32 110 DNA Homo sapiens 32 atct ctgatgtaaa gggt acaaaaccac	atcc ccagaaccgg gccattgcca  29 110 DNA Homo sapiens rs434504  29 ctct ctggctgcat agaggtttta  acctg ttgacactgc cagtgggttt  30 110 DNA Homo sapiens rs2814778 ()  30 agac agaagggctg ggacggctgt  atga gggccatcag gttctgggca  31 110 DNA Homo sapiens rs1325609  31 agag tgctgagaga cgagggatga  gagt ccaggctcct gctcttatca  32 110 DNA Homo sapiens rs6003  32 atct ctgatgtaaa gttattgtat agggt acaaaaccac tggagggaag	acc ccagaaccgg gccattgcca cacccaatca  29 110 DNA Homo sapiens rs434504  29 20 210 Ctct ctggctgcat agaggtttta atgaaacaag 2110 DNA Homo sapiens rs2814778 (FY-null)  30 30 31 30 31 30 31 30 Agac agaagggctg ggacggctgt cagcgcctgt 32 33 34 35 31 31 31 31 31 31 31 31 31 32 32 33 31 32 32 32 34 32 34 35 34 35 35 36 37 38 38 38 38 38 38 38 38 38 38 38 38 38	acc tttgctccc tacccactg actctactga ggctcacctr acc ccagaaccgg gccattgcca cacccaatca gggtgtctgg  29 110 DNA Homo sapiens rs434504  29 21ctc ctggctgcat agaggtttta atgaaacaag gggataaccr actg ttgacactgc cagtgggttt tgcaggcaga gcaccctgcc  30 110 DNA Homo sapiens rs2814778 (FY-null)  30 agac agaagggctg ggacggctgt cagcgcctgt gcttccaagr atga gggccatcag gttctgggca gggacagggg gcatcaggaa  31 110 DNA Homo sapiens rs1325609  31 agag tgctgagaga cgagggatga tggctccact gttgtgctgs gagt ccaggctct gctcttatca gaacagcaat ggtagctgct  32 110 DNA Homo sapiens rs6003  32 atct ctgatgtaaa gttattgtat aaaattcaag agaacatgcr aggt acaaaaccac tggagggaag gatgaagaag tggttcaatg	acc tttgctccc tacccactg actctactga ggctcactr ggcgcccct acc ccagaaccgg gccattgcca cacccaatca gggtgtctgg  29 110 DNA Homo sapiens rs434504  29 20 210 2110 2110 2110 2110 2110 211

<400> 39

			•
<210>	34		
<211>	110		
<212>	DNA		
<213>	Homo sapiens rs1406869		
<400>	34		
	cca tattgaaaag agccatccat gctcat	age atggetetev	cctccctcct 60
	J J J S S S S S S S S S S S S S S S S S		
tatgtgt	iggt aatcatgtga gaaacagtca gctgag	cttg tgacatgaaa	110
<210>	35		
<211>	116	•	
<212>	DNA		
<213>	Homo sapiens rs1004571		
400	25		
<400>	35		ht
getgee	gga cccctactga gagaatagag accttg	get teatetette	ttgccrgaga 60
gctctg	ecta tggcetegea gacageaate agteaa	get cagecaceca	cacttc 116
<210>	36		
<211><212>	110 DNA		
<213>	Homo sapiens rs172982	•	
<400>	36		
agattg	caag caaggcattc tcgagtttca attctg	gaag agaaagcacr	ttggagcatt 60
taataa	room poetrametan paganopen tembet		127
cggcag	gacg aactagetee agggeaaaga taetgt	gec ggcactcact	110
<210>	37		•
<211>	110		
<212> <213>	DNA		
<b>\213</b> >	Homo sapiens rs138335		
<400>	37		•
gatgttt	aat tcaacatgca gttcagaaaa atgaca	gatt tgtcttgtas	aaaaagacct 60
aacacaa	aget aageetttaa gaaaaccaac etcaac	igca tgaggcactg	110
*	•		•
<210>	38 .		•
<211>	110		
<212>			
<213>	Homo sapiens rs3287		
<400>	38		
	atcc atcctccaac acacacacaa ataagc	agct aatggcaatr	ctagtggtct 60
tcccaat	tca caagacctgt gcttcaaatt gttttc	etga taatgtggag	110
<210>	39		
<211>	110		
<212>	DNA		•
<213>	Homo sapiens rs959929		

gtagtgtagt ctttattctt ggtgaaagga aagaaatgtg caaatgctts tttgcctgtt	60
gcacagtttc aaggatgaat ttatccacac atccactttt gaaggtgcct	110
<210> 40 <211> 110 <212> DNA <213> Homo sapiens rs716373	
<400> 40 ccaggtccct gtggctggac ggggcacagc ttgggagcag ggcttcatgy tacctttgct	60
ctgtgcagca aagccctgca tggtcactgg tgtctggctc tctcctcctc	110
<210> 41 <211> 110 <212> DNA <213> Homo sapiens rs729253	
<400> 41	<b></b>
ttgcctatcc tagtaccacc ctcactgctt ctggtgctca cccatttctr gaacaaagct ctgcattttt tatatcaaat gtgcagacgt gtgccacaag cacacctcct	110
organization curational gracing gracial and calcacticit	110
<210> 42 <211> 110 <212> DNA <213> Homo sapiens rs1108718	
<400> 42	<b></b>
actagetece agaaagtagt ttateaegge teaacaaaag ateaataaer ttgattgaat tecatteata etagaaaaca tatetattta actgtaceet cagaaaaatt	60 110
<210> 43 <211> 110 <212> DNA <213> Homo sapiens rs1399272	
<400> 43 aactttaaac ctattgteta ccacatccaa ccaccacatt gctttggctr cagagatcag	60
tttaaagagc agatgattaa aagaattaac cacactttag aggtgaagaa	110
<210> 44 <211> 110 <212> DNA <213> Homo sapiens rs361065	,
<400> 44 cagtggtgac ttgttagtta tgtgactaca aacaaggcac agaaaactgr ttgagcttca	60
ggtttttcat atccttgaga attatagcat ctgtatgtca gggttattaa	110

<211> 110 <212> DNA <213> Homo sapiens rs361055	
<400> 45 aatagaaaaa ccaccagaat tagctacacg gaatacacgg atcactgccr tetcetcaga	60
gggatcaatg cgtttaagaa aaacacttgg ctcagcccaa ggagaaccag	110
<210> 46 <211> 110 <212> DNA <213> Homo sapiens rs733563	
<400> 46 ccagcgtaac caccttctcc tgactttagc cctgacatcc ctgctttctr gtaatcccca	60
gctccaggca agcccgaagg ccagcaccct acactcaccc ttcctgggtg	110
<210> 47 <211> 785 <212> DNA <213> Homo sapiens 712047 (HMGCRE12E14-3_786)	
<400> 47 acccaaaagg tatcactaat tgtccttaaa actcttctca ttgccttacy tatgatgtat	60
ttttaaactg gcaaatatat aaatgccaac ttacacctat tgctctgcag cctctattgg	120
tgctggccac aagacaacct tctgttgttg ccattggaac ctgaaattct ttttcatcta	180
agcaaagggg teetgeeact ecaacaggga tgggeatata tscaataaca tteteacaac	240
aagctcccat cacctaaaag gtaaagtcag gcaccaaatg aaaatctata tagtaaatgc	300
acaaaatttt atctcagctt gtcagtataa ctatcttcaa acttaatcct ttagtatgta	360
ttctttttaa acaaaatgtt taattcacct ttaaaaagtg tttaaaatac taaaatcttt	420
gagtaatgac tataaaagca ggaaatatat ttttaatttc tatgacctac atcataagca	480
gaataaaaaa ttaggataaa afgatttaaa aggaaaatgt tttataaaac tatgtacata	540
tatgaaacta atcaagaaaa aagattaaaa acatacaaag caaatctctc ttaatcgaga	600
aaataacatm ccaaggagta attataatcc ctgtaaggta ggtactggag agaagaaggt	660
tetgaaaget tettggaaag taaetgtegg egaatagata eaceaegete atgagtttee	720
atcagagttt ccaacttgta ggctgggata tgcttagcat tgactaactg gatgatctca	780
gcatc	785

<210> 48 <211> 110

<212> DNA

<213> Homo sapiens rs960709

400 40	
<pre>&lt;400&gt; 48 ggccctaagc cagagctgga gctcaggccc cc</pre>	cagagttcc gtccagggcr ctccaggact 60
cttcttaaca aagctataga aattccgagc to	gaaaagacc cataaagact 110
<210> 49 <211> 110 <212> DNA <213> Homo sapiens rs3317	
<400> 49	
atttacatct atttatttcc attcagtgta to	cacatette aggataggtr ataacagtgt 60
gaagggtgtg ctcattttct tcagctgtga gt	tagaggagt cttcccgaga 110
<210> 50 <211> 104 <212> DNA <213> Homo sapiens rs3340	
<400> 50 gaaatcgaat acgtccattt ctttgtaaaa ta	aacaataac gttraaggca aaagcaagat 60
tetgtaaacc aacattggaa aaggggacac ag	gggaggggc agag 104
<210> 51 <211> 98 <212> DNA <213> Homo sapiens rs933199	
<400> 51	
agggaaaggg aagtaggcat gtaccatttt aa	•
gaagaaagac tttctgcttt tctctgacta ti	tctgtat 98
<210> 52 <211> 110 <212> DNA <213> Homo sapiens rs1476597	
<400> 52 ctgtccctat cttcaccagc ctgccctcaa a	tcattggtt caaatcgcts tcgagaagtt 60
tgggagetee cagggegggg getttetaac a	getetgeet agetegggee 110
<210> 53 <211> 110 <212> DNA <213> Homo sapiens rs1155513	
<pre>&lt;400&gt; 53 ggaaaagagt gtggaagtta actaaatctg c</pre>	agctaatac cacagaaccr tggagatgat 60
ctgaatgaag aaacttagaa aatgtggtgg g	tcaqaacaa qaqtttactc 110

<210> 54	
<211> 110	
<212> DNA	
·	
<213> Homo sapiens rs984654	
<400> 54	
aaaaaatcct gataagcact gttaggcata ttaactttaa tgattacaay ttttaggaca	60
$\cdot$ '	
ctctgtggcc tagacttaga aacacaacta atgtccagaa aaagattcct	110
•	
<210> 55	
<211> 110	
<212> DNA	
<213> Homo sapiens rs2161	
-	•
<400> 55	
tecectecet eggeacecae teceaggeea agagegteet atgaateatr catttgttee	60
tgttattgct gttcacagag tggcaactct tgcaaaggga ggggtacaaa	110
egergese geography eggenness serveses geography	
<210> 56	
<211> 51	
<213> Homo sapiens CYP3A4E10-5_292	
400 56	
<400> 56	51
ttttacccaa taaggtgagt ggatgrtaca tggagaagga gggaggaggt g	21
<210> 57	
<211> 110	
<212> DNA	
<213> Homo sapiens rs869337	
<400> 57	
atccctgcaa cttggcacgt cttgctgtaa atggaagcag tggcatgtcy gtgtcaggca	60
tgaaagccgt tcataaaacc accttgctga gcacttcctg tctgtcaggc	110
<210> 58	
<211> 110	
<212> DNA	
<213> Homo sapiens rs285	
<400> 58	
ctgcaagggt tttgcttaat tctcaattca atgtctcttc atcttttagy agctgtgggg	60
segundada occasoran continuous augustos activities and a designation activities activities and a designation activities ac	
	110
ttttgttgtt gttcttctgt ttttgcttag tatctgacta ctttttaatt	110
	110
ttttgttgtt gttcttctgt ttttgcttag tatctgacta ctttttaatt	110
ttttgttgtt gttcttctgt ttttgcttag tatctgacta ctttttaatt	110
ttttgttgtt gttcttctgt ttttgcttag tatctgacta ctttttaatt <210> 59 <211> 110	110
ttttgttgtt gttcttctgt ttttgcttag tatctgacta ctttttaatt  <210> 59 <211> 110 <212> DNA	110
ttttgttgtt gttcttctgt ttttgcttag tatctgacta ctttttaatt <210> 59 <211> 110	110
<pre>ttttgttgtt gttcttctgt ttttgcttag tatctgacta ctttttaatt &lt;210&gt; 59 &lt;211&gt; 110 &lt;212&gt; DNA &lt;213&gt; Homo sapiens rs716840</pre>	110
ttttgttgtt gttcttctgt ttttgcttag tatctgacta ctttttaatt  <210> 59 <211> 110 <212> DNA	110

agtcagg	gega tigeticati eteagetitg	tggagagtga	tatacacaga		. 110
<210><211><211><212>	60 79 DNA				
<213>	Homo sapiens rs3176921(CF	RH2942)	•		
<400> ggagtag	60 gete ttgtcateat etaaaaact	tgaactgcat	tttgagagay	ttattggcct	. 60
tgcttct	gca ggctcataa				79
.010-	<b>C1</b>				
<210> <211> <212>	61 110 DNA				
<213>	Homo sapiens rs1467044				
<400> agaatco	61 cccc ttgccattcc catccctgta	tggtccttcc	tctagggctr	tttgggaata	60
gtactta	atgg tetaactggt tetgeagetg	cccttcgtgt	ggttataaat		110
<210> <211>	62 110				
<212>	DNA				
<213>	Homo sapiens rs998599				
<400>	62 etgt gttttgtegt gtatgtagga	tttacccaqt	agcaaaactr	taaattcagg	60
	aago taaaacttoo taaagoaata				110
Jeres		oo o o o o o o o o o o o o o o o o o o			
<210>	63				
<211><212>	110 DNA				
<213>	Homo sapiens rs1041321				
<400>	63			1.	
	agtc tgagcatctg aaggaggatg	•		caggicigia	60
aatgtc	gtgt gtatgaatga atccacacca	gtcccttccc	atagacaaca		110
<210>	64				
<211><212>	110 DNA				
<213>	_				
<400>	64				<b>.</b> .
	tcac teteetteee agateataga			cacgactetg	60
ctacaga	acct gcctgtgatt tcaggctaac	cgctgagggt	cacagagata		110
<210>	65				
<211>	110				

<212> DNA

<213>	Homo sapiens	rs913258				
<400>	65					
atcagag	gga gagaataaat	gaccctgata	ctcttactta	ggaaggcats	ctccccctc	60
catacac	tcc ctcgagaato	atggctttaa	agacacagag	atgagaaaga		110
<210>	66				•	
<211><212>	DNA				•	
	Homo sapiens	rs2695				
<400>	66					
agcatac	tga agactattco	c taaatggcta	tttgtgttgg	gtggtcaagr	ctattcagaa	60
aatctca	gag gaggacaaat	ga .				82
						•
<210>	67 ⁻					
<211> <212>	110 DNA					
	DNA Homo sapiens	rs675837				
<400>	67 :tat ttatgtcta:	a ctcctcagca	aaggaaggcc	ttacttaacr	tgataaacat	60
						110
tcacatt	tta ttttagtta:	a geatttettt	aaatgtaaga	aattggaggc		110
-010-	68					
<210> <211>	68 110					
	DNA	•				
<213>	Homo sapiens	rs662117				
<400>	68				•	•
ctaaagg	ggga cactactgt	g tccaggagca	tctcctctct	cccaacacay	acacacacaa	60
caaaata	aga tcaaacctg	c tgatttaata	tcaattttc	aacaaaaatt		110
						•
<210>	69					
<211>	110					
<212> <213>	DNA Homo sapiens	rs590086				
	69 :ttt atgagtcct	c teceseattt	ttaaattaaa	taccaaacar	tattassaa	60
cacgee	cee acgageeee	c cggcgcccc	ccgocccggo	caocgaager	cocogaaago	•
agagatt	ttt gaaagcccc	a ttgatttttg	tgtcatccat	ctcagatcct		110
<210>	70		•			
<211> <212>		•				
	Homo sapiens	rs9032				
<400>	70					
	agtg cctcttgta	a gcagccgcct	caacactgaa	cgcagaaaas	aggacaaaac	60
catctg	gtg tcagcctgt	c agcatatcac	tatataaatt	aaaagtcaaa		110

•	
<210> 71	
<211> 110	
<212> DNA	
<213> Homo sapiens rs1076160	
<400> 71	
gccagacett agtggggaaa gcgtttcaga cccagacagt atgtctcccr catggcatag	60
agaatggett gatgaettat tteetaaaag aeggagegge agteeeacta	110
<210> 72	
<211> 100	
<212> DNA	
<213> Homo sapiens 32934	
<400> 72	
gcgcacgcac cagagccagt ggcaaagcaa cttaagctgc aggctcctcy tgcccacccc	60
	-
cageegegae geeeetgggg geeettagea gaetgeeaag	100
<210> 73	
<211> 100	
<212> DNA	
<213> Homo sapiens 235936	
<400> 73	
atagtattte ttttetteag tagaatggte aggtetetaa gtgetgeagy ggettgaaet	60
cttgatgaca aacatgtagg aaaacttcat gcagccagtt	100
<210> 74	
<211> 100	
<212> DNA	
<213> Homo sapiens 520354	
400 84	
<400> 74	60
cttcaaaggc cagaggcttg gagctggcac aaggagtcct tgtgcagaar agaacagaag	00
charteters transferent contaggers annotative	100
gtggttttgc tgccctacat ccatccccgg aaccttctca	100
<210> 75	
<211> 100	
<211> 100 <212> DNA	
<213> Homo sapiens 539619	
CZISV HOMO BAPICHS SSVOIV	
<400> 75	
tccctataag gtatactgtt taaagactag gtccttggct ttgccttcar gtaacttgaa	60
- cooleaning generalize changing and green and	
gatgtttttg ttgaaatatc ttggagacta aaaaatagtc	100
<210> 76	
<211> 100	
<212> DNA	
<213> Homo sapiens 715029	
Table topon , about	

	76 ccc ctactcagtt	gccttgaata	ggaattatac	tgaccaaaar	ctagathtaa	60
	•					
aatgaga	aac tttaacatac	ccagttagac	ccacctacac		•	100
	•					
<210>	<b>7</b> 7					
	100					
	DNA					
<213>	Homo sapiens	719185				
<400>	77					
tgagaag	aga gagcaactgg	aagggtgagg	ggagatactg	agttttagay	ggttttcgaa	60
aaagcaa	gtc actgcccaac	aagtgatgtt	caqtaqacaa			100
-		3 3 3				
<210>	78					
	100					
	DNA					
		726176		•		
<400>	78					
caaaaag	gca gctaaaaaga	caaataaata	taactggtgg	cacttactty	agtgtctgag	60
ctttaaa	gtc aaatttggct	ctacaaatc	tcatctcaac			100
0000000	goo aaacccggcc	. ccggcaggcc	ccaccegade			100
		•				
	79			•		
	100					
	DNA					
<213>	Homo sapiens	734463				
<400>	79					
gttaaca	tat ctgaaattca	taggtaacac	ctttcccaag	ataatttagy	tctggtatta	60
catttac	aat ottoatooto	attttt	tataatatt			100
gacccag	aat cttcatcctg	geeeeeegg	cctagigitt	•		100
			•			
	80					
<211>	100					
	DNA					
<213>	Homo sapiens	739485	•		•	
<400>	80					
tcaatag	ata ggttgatcac	tctataaaat	agtagtctgt	tggcacttay	gtgcgtaacc	60
		•				
agaggcc	atc atgttctgag	actgcccag	ggctttttgg			100
<210>	81					
<211>	100					
<212>	DNA					
<213>	Homo sapiens	765480				
-400:	01				·	
<400>	tcc tgcagagggg	. aaatoataso	tagasatsas	taataaata	anttontata	-
ugagaag	cycayayyyy	, audicalaac	Loggagtacc	coccaacay	accegatata	60
a+a+aaa	ata attatatat			•		7.00

WO 2004/016768 PCT/US2003/026229 17/68

<210>	82	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 781062	
<400>	82	
ttttat	tagt atagtggatg tetettttgt tteeetttet tgteatttew etttetttee	60
cctgag	gcaa atactattat gaatttagtg tcttcatttt	100
010	0.0	
<210>	83	
<211>	100	
<212>		
<213>	Homo sapiens 878201	
<400>	83	
	·	60
	egtt gtggactttg ggetgggeet tgeaatttet etggeeaagr gatgttggee	80
teetae	ctgc tgaggacatg ccctgaagaa cgtgtgcccg	100
ccccgc	cese esassacaes eccesaasaa esesteces	100
<210>	84	
<211>	100	•
<212>		
<213>		
<400>	84	
	ccat tetgtgeeat eccatgeeae tecetteeet gecaeteeaw getattagaa	60
		•
ttqqaq	gacag agaccccaat tacttttaat cctcagaatt	100
ttggag	gacag agaccccaat tacttttaat cctcagaatt	100
ttggag	gacag agaccccaat tacttttaat cctcagaatt	100
ttggag	gacag agaccccaat tacttttaat cctcagaatt 85	100
		100
<210>	85 100	100
<210> <211>	85 100	100
<210><211><211><212>	85 100 DNA	100
<210><211><211><212>	85 100 DNA	100
<210><211><211><212><213>	85 100 DNA Homo sapiens 886229	100
<210><211><211><212><213>	85 100 DNA Homo sapiens 886229	
<210> <211> <212> <213> <400> ccttac	85 100 DNA Homo sapiens 886229	
<210> <211> <212> <213> <400> ccttac	85 100 DNA Homo sapiens 886229  85 ecctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa	. 60
<210> <211> <212> <213> <400> ccttactagtta	85 100 DNA Homo sapiens 886229  85 ecctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact	. 60
<210> <211> <212> <213> <400> ccttactagtta	85 100 DNA Homo sapiens 886229  85 ecctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact	. 60
<210> <211> <212> <213> <400> ccttactagtta <210> <211>	85 100 DNA Homo sapiens 886229  85 ecctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100	. 60
<210> <211> <212> <213> <400> ccttact tagtta  <210> <211> <212>	85 100 DNA Homo sapiens 886229  85 ecctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA	. 60
<210> <211> <212> <213> <400> ccttactagtta <210> <211>	85 100 DNA Homo sapiens 886229  85 Except gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA	. 60
<210> <211> <212> <213> <400> ccttact tagtta  <210> <211> <212> <213>	85 100 DNA Homo sapiens 886229  85 ecctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA Homo sapiens 898455	. 60
<210> <211> <212> <213> <400> ccttac  tagtta  <210> <211> <212> <213> <400>	85 100 DNA Homo sapiens 886229  85 Exectt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA Homo sapiens 898455 86	60 100
<210> <211> <212> <213> <400> ccttac  tagtta  <210> <211> <212> <213> <400>	85 100 DNA Homo sapiens 886229  85 ecctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA Homo sapiens 898455	. 60
<210> <211> <212> <213> <400> ccttac  tagtta  <210> <211> <212> <213> <400> gtetce	85 100 DNA Homo sapiens 886229  85 ecctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA Homo sapiens 898455  86 etttg aacatctggc ctggactcat ccccatgtct gtccatctgy tgtcatccca	60 100
<210> <211> <212> <213> <400> ccttac  tagtta  <210> <211> <212> <213> <400> gtetce	85 100 DNA Homo sapiens 886229  85 Exectt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA Homo sapiens 898455 86	60 100
<210> <211> <212> <213> <400> ccttac  tagtta  <210> <211> <212> <213> <400> gtetce	85 100 DNA Homo sapiens 886229  85 ecctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA Homo sapiens 898455  86 etttg aacatctggc ctggactcat ccccatgtct gtccatctgy tgtcatccca	60 100
<210> <211> <212> <213> <400> ccttact tagtta  <210> <211> <212> <213> <400> gtctcc ggcctg	85 100 DNA Homo sapiens 886229  85 cctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA Homo sapiens 898455  86 ctttg aacatctggc ctggactcat ccccatgtct gtccatctgy tgtcatccca agget gcccacagcc cgggccgcag tcaccactca	60 100
<210> <211> <212> <213> <400> ccttac tagtta  <210> <211> <212> <213> <400> gtctcc ggcctg	85 100 DNA Homo sapiens 886229  85 cctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA Homo sapiens 898455  86 ctttg aacatetgge etggacteat ecceatgtet gteeatetgy tgteateeca ageet geceacagee egggeegeag teaccactea	60 100
<210> <211> <212> <213> <400> ccttact tagtta  <210> <211> <212> <213> <400> gtctcc ggcctg	85 100 DNA Homo sapiens 886229  85 cctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA Homo sapiens 898455  86 ctttg aacatctggc ctggactcat ccccatgtct gtccatctgy tgtcatccca agget gcccacagcc cgggccgcag tcaccactca	60 100
<210> <211> <212> <213> <400> ccttac tagtta  <210> <211> <212> <213> <400> gtctcc ggcctg  <210> <211>	85 100 DNA Homo sapiens 886229  85 ccctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa aatat tttttaagga tttattatgt gtgaagtact  86 100 DNA Homo sapiens 898455  86 ctttg aacatctggc ctggactcat ccccatgtct gtccatctgy tgtcatccca ggcct gcccacagcc cgggccgcag tcaccactca  87 100 DNA	60 100

20,00	
<400> 87 tgattcacgt gtggcaggga gggccgtcat taatgaaaat gcgagggcgk cgcacgagtg	60
ttgatgactc agcaggcctt tctacttctg tatgagtcag	100
<210> 88 <211> 100	
<212> DNA	
<213> Homo sapiens 994174	
<400> 88	
cttctcatca aataagtttg agtgttaagc atattgtgga atttggagar tggcactgag	60
tactattaga agcagtactg acgcaaatac attgcatact	100
<210> 89	-
<211> 100	
<212> DNA <213> Homo sapiens 1009738	
<213> Homo sapiens 1009738	
<400> 89	
tctctccaca gggccccagc caccgaaggc tcaaaaaaaaa aaaaaaaaaa	60
aaaccctgca tggggagttg tgagaatctg aagcctgata	100
<210> 90	
<211> 100	
<212> DNA <213> Homo sapiens 1023996	
<pre>&lt;400&gt; 90 tagagaagaa tattetaaat ettettgtgt etteteettg ttggtetter ttetaateat</pre>	60
tggaatcatg aagaagttta gtttctcttc tgtgtgactg	100
<210> 91	
<211> 100 <212> DNA	
<213> Homo sapiens 1324758	
<400> 91	
tgaaaaatgc ttcccagaaa tatgctcata tcattcatcc attgacaatm atcatttgtt	60
ggacatcaac tgcttgtact acattagctt atgacatgaa	100
ggacarcaac tgccogtaco acarragere acgacacgaa	
· <210> 92	
<211> 92 <211> 100	
<212> DNA	
<213> Homo sapiens 1361107	
<400> 92	
gtacttattc ccacagecaa etteaaatga ecceeteeet ggeeceedem caagaaaaaa	60
acagggcagc tgaaaagttt gcttaaactt cttaatactt	100

$\cdot$	
<210> 93	
<211> 100	
<212> DNA	
<213> Homo sapiens 1367990	
400 00	
<400> 93	
ctttaagatc catgtaaaaa tattgcaaat gagggatttt agtagtaagr aaaatttgcc	60
·	
aatttttact cgattagttt ggctattttt atatattctt	100
<210> 94	
<211> 100	
<212> DNA	
<213> Homo sapiens 1443995	
<400> 94	
ccatctggcc taccctaggt taacgctagc ttgcattttg ctttgcttty tgattttaat	60
acatatgtca gtattactaa acaatatatt gtttagtttt	100
<210> 95	
<211> 100	
<212> DNA	
<213> Homo sapiens 1553018	
<400> 95	
tcaagegatt cttgtgcctg agcctcttga gtagctggga ttacaggcaw gtgccactgt	60
gcttggctaa ttcatatata tatatatata tatatatata	100
georggeoda recaracara racarara racararara.	
<210> 96	
<211> 100	
<212> DNA	
<213> Homo sapiens 1806913	•
<400> 96	
tttaaaatgt tactttattt tagaataagg tagagttgaa aattgtcagr aatacaaaca	60
occurred processes calendad collaborate and all and and all and and all and a second collaborate	• •
ttatttaagt atttaatggt ttatatatgt gaagtttaa	100
ttatttaact atttaatcct ttatatatgt gaagttttaa	100
<210> 97	
<211> 100	
<212> DNA	
<213> Homo sapiens 1806995	
•	
<400> 97	
ataatactta attttttggc tatgaagagg agaaaaatca tcattttcty atttttctca	60
acadactica acceptage tatgatagg against teaction acceptage	- 00
	100
tcaggggact ctgaagggct tgggaggagt gatctgccca	100
<210> 98	
<211> 100	
<212> DNA	
<213> Homo sapiens 1966625	
-510	
.400. 00	
<400> 98	

atttagtccc agaagggcgt gaggacatca tgtagacaca aaagagcatr caacagcaaa	60
ttattggggg taaatattta atatcttgcg tctggggaac	100
ctattggggg taaatatta atatttoggg tolggggaa	
<210> 99	
<211> 89	
<212> DNA	
<213> Homo sapiens 2001433	
<400> 99	
caccacttta cattetetea acaacagege acaagggtwe cagtttetet geateettga	60
caacacttgg gattttctgt tattttgat	89
<210> 100	
<211> 100	
<212> DNA	
<213> Homo sapiens 2012067	
<400> 100	
ggttgtgggg ttetecaggt getetgtege ceaggetgga gtgeagtggy gegatetegg	60
at an at many such as tracks as a grant to a strack of tracks	100
ctcactgcaa cetecteete eegggtttaa gtgattettg	100
<210> 101	
<211> 100	
<212> DNA	
<213> Homo sapiens 2012661	
<400> 101	
<400> 101 aatggttgca tattattgtc aagtgggtgt taccacatga atatttacak ttgttcagct	60
catctgtgaa tttcttctct tcattcccag gactcaccat	100
<210> 102 <211> 100	
<211> 100 <212> DNA	
<213> Homo sapiens 2035324	
<400> 102	
aactaactta gagaagccac taaaggacaa ttcattctaa ttttcacgtr aatcaaagac	60
that there is a character of a contract of the contract	100
ttatctgact aatgctgaat aagttgggga gagatttcat	200
<210> 103	
<211> 100	
<212> DNA	
<213> Homo sapiens 2048326	
.400- 102	
<400> 103 aaaaagaatg gcatttggca agatgagact ttggttccta actgctctar gacctaggct	60
andanganey yeareregen anarganare regarrees assessar garreanges	
ggcttgaggt tcaaagtgcc ggttactctc catttgactg	100

<211> 100	
<212> DNA	
<213> Homo sapiens 2061254	
<400> 104	
agcagtggtg agaggggaag agatttttct atgccatgtt tcctgacagk agcagaacct	60
ageagegges agaggggas agaeteree argeoarger recegangs, agoaganoor	
atalkangan atganggat tataattita taganahat	100
gtcttcagac ctgaaacact tctcctttta tagtcactat	100
<210> 105	
<211> 100	
<212> DNA	
<213> Homo sapiens 552647	
<400> 105	
tactgcatga aggaccaaac caacaagcca gtcaggacac caaaagcacm caaggaagca	60
aggaacaaga tgctgcagtt cttaaaataa actgtaggtg	100
<210> 106	
<211> 97	
<212> DNA	
<213> Homo sapiens 1022573	
400 106	
<400> 106	
attgacatca acaaaagtta tcacagggtt aaactgacac acgtttmatg atatgcgtag	60
tcattgctaa gtaagtaggc ataagaacaa ggtaatt	97
<210> 107	
<211> 100	
<212> DNA	
<213> Homo sapiens 1076245	
<400> 107 .	
aggtgccccg gcccagagaa cctgccagtg aaattgcgtc ctacccacar cccctcgttc	60
ccatagetat ggeettaett ggggeeetet ttgeetetga	100
Contragation agreement aggreence to green and a	
<210> 108	
<211> 100	
•	•
<212> DNA	
<213> Homo sapiens 1365259	
<400> 108	<b>~</b> 0
tattaaccac ctccaacctt ctgtctagag atcatcctta ttatcactaw actctagttt	60
	100
ggctataaag tatataaaga agtagatgtc aaaatggaca	100
•	
<210> 109	
<211> 100	
<212> DNA	
<213> Homo sapiens 1409133	
<400> 109	
taagacccct gtcatttgtt caacagatgt teceetecaa ggeteaccay aatetgeece	60

aaatccatct ctctggtttt atcttctaaa attccccacc	100
<210> 110	
<211> 100	
<212> DNA	
<213> Homo sapiens 1566088	
<400> 110	60
gttcctcttc aggtcggtca agaccaggtt tggaactcag cttgttcacr tctgtttcca	60
agtcatcatc ctaaaatgga atagtcatag aagagattac	100
<210> 111	
<211> 100 <212> DNA	
<213> Homo sapiens 1934073	
<400> 111	
tttttcccat tctgtgggtc tgttcacttt gttgtttttg tttttttgtk tgtttttgtt	60
tttgttttgc tatgcataaa ctttttagtt ttatgcaatc	100
<210> 112	
<211> 100	
<212> DNA	
<213> Homo sapiens 147316	
<400> 112	
cttttggggc tgaggtaata gtggaatgct cccttacagg aagctacccr ggacttcaca	60
ggccctgtgc cagtgacttc cttgaaacca ggaatattga	100
<210> 113	
<211> 100	
<212> DNA	
<213> Homo sapiens 738987	
<400> 113	
ttgcaaacgt ggagtgaggg aaggggatta aaaagggaga aggtgaatar gtgactttga	60
atccaaccta cagtggttaa ttttcattta actatctggc	100
<210> 114	
<211> 100	
<212> DNA	
<213> Homo sapiens 740910	
<400> 114	
attgttaagt ataacagttt gctaagtaag gtgagtggta taatcatatr tttgtaaaaa	60
gcaaaacaaa ataaagccaa aactcgaacc tatctttgtg	100
<210> 115	
<211. 100 -211. 100	

<211> 100

<212> DNA	
<213> Homo sapiens 764232	
400- 115	
<pre>&lt;400&gt; 115 tgtcaaaggg gtctttattt aagcctcaga acacatctag tagaagacar gtaatatgag</pre>	60
	00
gaaagaggtt gaggtaggac aagactgaaa actttgtaaa	100
<210> 116	
<211> 100 <212> DNA	
<213> Homo sapiens 978009	
<400> 116	
gtccacaggg cccctgccct gcctttctgt ctgtcataga gcaggaacgr ctgaccatcc	60
	1.00
agatgtccct cagcgagaaa ccctgactat acaaatccat	100
<210> 117	
<211> 100	
<212> DNA	
<213> Homo sapiens 1327678	
<400> 117	
atctccatct ataaaatggg aatcacagta gtacccacct catatgaccr ttgtgaggat	60
tagatgacat tttatatgga aaatgtctgc agtaaaccgt	100
<210> 118 <211> 100	
<212> DNA	
<213> Homo sapiens 1954787	
<400> 118	
ttgagaaatt ttgaggaagt acaaccaaaa gcaattggag actggttaty ggaaggtgcg	60
gaattgggtg aaggcacgat gcctgggtag ctggtgctgc	100
24400222-2 4422040240 2001222042 002202020	
<210> 119	
<211> 100	
<212> DNA <213> Homo sapiens 2049252	
22137 Nomo Baptens 2047232	
<400> 119	
acaatctgtc ttgcactaaa actaggcgtt aaagcaccat ctaagaattw tttttttgag	60
ctagaagatg gacccattta agaaaggtca cagttcettc	100
<210> 120	
<211> . 100	
<212> DNA	
<213> Homo sapiens 154497	
400. 120	
<400> 120 agatgagcag ccagttaagg ttaaagaatc agtggatttg gaataaggay ggaggtttgg	60
-ancaraces consecutas contratures adealacera aurentain adadaceras	50

aggattetgg gettgggaaa aataagagea aaggtaetga	100
<210> 121 <211> 100 <212> DNA <213> Homo sapiens 268666	
<pre>&lt;400&gt; 121 ggtcaggggt tgggctgact catccagacc tctaaacatt atctgaggcy acctaggggg</pre>	60
tcagggggag ttaaggcagt ggcctctggc tgcctcggtt	100
<210> 122 <211> 100 <212> DNA <213> Homo sapiens 327219	
<400> 122 cttccttcct tccttccttcct tccttccttc cttccttccy tccctctct	60
tototttott totttottto ttttattttt gttttgggtg	100
<210> 123 <211> 100 <212> DNA <213> Homo sapiens 434363	
<400> 123 agagcagaaa cccaaaagct gttagctgaa aaacaaacaa aaaatacctw tggtgaagcg	60
tgatttttgg gtgaagttaa aggcaggaag ctggtagagt	100
<210> 124 <211> 100 <212> DNA <213> Homo sapiens 653220	
<400> 124 ccaagtactt acaggatttc tccaaatgtg ttgtattgat aataatctgr acaagctaga	60
aaacaacaaa cacatgtaaa agtcatttaa ctactcaccc	100
<210> 125 <211> 100 <212> DNA <213> Homo sapiens 655351	
<400> 125 tttcagaagc taaccacctg acttgagact agaattgcac atgttgcatr ccattcttgc	60
ctttgactct ttcagacatt gtatccaata tcatctatca	100
<210> 126 <211> 100 <212> DNA	

<213> Homo sapiens 699863	
<400> 126 taaggtacta taattgatat acttgtatac agggtcctta taaaatacar ctcataccta	60
aatccagata tttaataatt gggcccatta agtccaatca	100
<210> 127 <211> 100	
<212> DNA <213> Homo sapiens 878213	
<400> 127 gtcatgaaaa ggttaatcta caatgcactt agacagctaa taataagtar aacatttcca	60
aagaaatagg aacctgggat tgtctctttt taataaatta	100
<210> 128	
<211> 100 <212> DNA <213> Homo sapiens 918880	
- <400> 128	<b>50</b>
cccacattag gcataaaaca taaatgtata ccttaataaa tttcggttay gtaaaagagc gcctgggtaa cctcacccag gttgagatgg aaaatgtgcc	60 100
	•
<210> 129 <211> 100 <212> DNA	
<213> Homo sapiens 957452 <400> 129	
tatacaggat ggggattcca gacaatcaga gtatgatgtt ctcttgtcay tgatcacatt	60
cactagtgac aactaattag cacatccgag taatcaaaaa	100
<210> 130 <211> 100	
<212> DNA <213> Homo sapiens 1004660	
<400> 130 gcaccgcctg tetataccag tgctgatecc acctetecc tececettey gecetectge	60
ctttgcaatt ggctgtttcc ctccagcttt gtccatcctc	100
<210> 131	
<212> DNA <213> Homo sapiens 1152202	
<400> 131 gcgggtggag ccctcctcta gctatcagca tgactgctgt gttttctgay tcaccagttc	60
cctgtccaag gccatcccta agctctgttt tcttccagtt	100

<210> <211> <212> <213>	132 100 DNA Homo sapiens 1251587	
<400> atggtad	132 ctat gctgtaataa aaacctgaag tgtgggagaa gtttcagaay caagaactga	60
gaggaag	gctg aaaggacttt gaggagagtg taagcagaat	100
<210><211><212><212><213>		
<400> ttttata	133 acaa attcatcttc ttatagttgg gatgttatta caaaagttts gatatcttaa	60
	ctaa tgttttagag tttcatattt attgacctga	100
<210><211><211><212><213>		
<400>	134 tttc aaaggtgtca tttatgtata catattattt ttatttttaw ttcacataca	60
gatacc	taaa ctattccatt tttaagagta gaaatcatgc	100
<210><211><211><212><213>	135 100 DNA Homo sapiens 1549681	
<400> agggga	135 gtga acagaggatg aaagagagaa gaaagggcag tetcageter etgtgecaga	60
	aget cagegtataa cagggaetee ataaatatgg	100
<210> <211> <212> <213>	100	
<400> tgcagg	136 gcaat ttagtagaca ctgaagcata tttttacctt tccaaattgr aagtggataa	60
caaggt	caaca atageaetgt ecetggagee atectgteet	100
<210><211><212><213>	100	

<400> 137 gatactcatc cctgggtccc acgcccagag agtctgatgt aatcagtgts gggtgtggac	60
agggcattgg gatttttaaa agctccccag gtgattgcag	100
<210> 138	
<211> 100	
<212> DNA	
<213> Homo sapiens 1932378	
<400> 138	
ctatatttta cgtaagttca aggtatgaca atgcatatgt ggcaaagatr tatgtagtct	60
gctttataaa aagtgtctaa ggaaacaaaa ttttctttcc	100
<210> 139	
<211> 100	
<212> DNA	
<213> Homo sapiens 1978355	
<400> 139	
atgtaacaaa tacaattaaa tggagatgca atttgattta ttgaaaatar catgatttta	60
atgeagatta actgtaggea gtacettace agaatataat	100
	,
<210> 140	
<211> 100	•
<212> DNA	
<213> Homo sapiens 2031262	
<400> 140	
attotgatag agaccatatg acctecaagt ggattagett teaatgaatk etacaaacca	60
gaagagtett teececatet teatetteat tetetaaett	100
<210> 141	
<211> 100	
<212> DNA	
<213> Homo sapiens 518116	
<400> 141	
taaggtgaga gaggcagaag gagggtggaa agaatggcgt ttttggtgcr gtgcaaacta	60
atttgcccaa actgaaaaag ctggtgctct caccattatg	100
<210> 142	
<211> 100	
<212> DNA	
<213> Homo sapiens 546445	
<400> 142	
cagtccttcc gccgccgggg ctaacacaat agtgaaagct gcgctcctgr tcgtcggcag	60
gctggcactg ttatacagga aaagggtcgt gtgcgacctt	100

<210>	143	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 715005	
<400>	143	
	aggt ctttaaagca gcacatgatt ttcaagtgag ttattcaaty aatcgtttct	60
gerere	aggs coccaaagea geacabgace cocaagegag coacceaacy aaccageses	•
atataa	ctct gatgcccctt tggactgtct gcttattgaa	100
gcacca	cee gaegeeeee eggaeegeee geeeaeegaa	
212	144	
<210>	144	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 763003	
	<b>.</b>	
<400>	144	
ctgagci	tcag ccagggacca ggaaccaggg accagagtca ggggagccgk ctagaacaga	60
agggca	cgtc ggtgctaggc tgggggaggc gcagagatgg	100
	·	
<210>	145	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 913315	
	•	
<400>	145	
	aagg ggaggccaca ggagttacag aggactctgg aaatacagar agatctgtga	60
tttact	cagt gggtatttga tagactaaga aggcaaattc	100
cocaco		
<210>	146	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 974828	
(213)	nomo saprens 374020	
, <400>	146	
		60
ccugca	gttt tccacatgga attcctcttc ttgcactttg gcctaagcay gcatttttc	30
<b>.</b>		100
taatgg	tgga aaatattaaa cagacttttg ctttagaagg	100
<210>	147	
<211>	100	
<212>		
<213>	Homo sapiens 1109525	
	•	
<400>		
tcagcc	caga gaaagagtca gtcccataat cccagatctc caccacatcy gcacagaggc	60
		•
ctgtta	lggec agggggetge eetgeteece getgeceeca	100
<210>	148	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 1325561	

<pre>&lt;400&gt; 148 attaaaaggg aactttttat ctcatatttg aaacataagt cacaagaatr actaacattt</pre>	60
atgetttaaa ttattaatee tagtttteag aattttatta	100
<210> 149 <211> 100	
<212> DNA	
<213> Homo sapiens 1382794	
<400> 149	
aaaactttta ccaataattt tcatggagga ttttatgaaa taaccaaaak ctattaaaaa	60
tgcttatgct gaaaaatata aaatattcag ttttctatta	100
ogeoverget gaadaaaaa aaababbabag boobbbabaa	
<210> 150	
<211> 100	
<212> DNA	
<213> Homo sapiens 1813081	
<400> 150	
tgcaaaagag aaggccacac cttgctcaga aagacctgat gtgtggggcy agtgcaaggg	60
gtcaaaaact gaggagaggc cagaaggcca aagcagcttg	100
<210> 151	
<211> 100	
<212> DNA	
<213> Homo sapiens 1412302	
<400> 151	
tcacataatg gaagaatggc ccggcctaga ccagaggttc ctaattaggr gtccatgagc	60
cacgttcaat tatatatatt ttttcttctg gtatttttct	100
<210> 152	
<211> 100	
<212> DNA	
<213> Homo sapiens 1536847	
<400> 152	60
cagtttaagg tggaaagggg gtaggaggaa ggaggaagtg tatttctttm gtcacattag	60
ccagattctc taccttgaca ttgtatctcc cagatctatt	100
· <210> 153	
<211> 100	
<212> DNA <213> Homo sapiens 231727	
•	
<400> 153	60
tettteetet tttatgttte ettggetaca tgetgggata ggggeteatr gtaagtttge	60
cagattcaac caaaaaacgc cacaaaaaca agaaaacagg	100

<210> <211> <212> <213>	154 100 DNA Homo sapiens 235936	
<400> atagta	154 httc ttttcttcag tagaatggtc aggtctctaa gtgctgcagy ggcttgaact	60
cttgat	gaca aacatgtagg aaaacttcat gcagccagtt	100
<210><211><211><212><213>	155 100 DNA Homo sapiens 246103	
<400> tttaac	155 cetec etecetgegg gegaatacee agttgteeca gtacattttr ttgatgagte	60
ctttt	tgtt tttgtttttt gagacacagt ctcgctttgt	100
<210><211><211><212><213>	156 100 DNA Homo sapiens 260707	
<400>	156 tgat geggeettae aaactatttg atgteeaaac eccateetty acegggeeta	60
		100
<210> <211> <212> <213>	DNA	
<400> acagtg	157 gccac cttcacgcca gggcctggga ggtctgtatg caggttaccr ggaagaataa	60
gtcagt	tttag tgagcattcg gcccataaaa ccaccgtgca	100
<210><211><212><213>	100	
<400> agtgtg	158 gtcac tcctaactct tcttacctcc tttctttggt ttaaggtctm ttagtgtctg	60
tactgg	ggtag tgttttcaca gctttctcct ctgacatggc	100
<210><211><211><212><213>	100 DNA Homo sapiens 386243	
<400>	±23	

cctcacccac gatggcggcc agtctcccga agcgcgaggc cccgcccagy tctcggcctc	60
tecegeggee ggteeetgea ceaagetege gagetgaetg	100
<210> 160	
<211> 100	
<212> DNA	
<213> Homo sapiens 410852	i
<400> 160	
tgggcctggg gggttcatgt ccaatgttgt ccaatcactg gataattctr acatctaact	60
aaacctcttt tataggaaaa agagatgctt taaaattgtt	100
<210> 161	
<211> 100	
<212> DNA	
<213> Homo sapiens 448083	
<400> 161	
ctgcaggcag actcagatca gcagagaagc atctcgcctc tggctgtgcy gtccaggttg	60
agctgcgtgt ggcagtgagc acagaggaga aatgcaggga	100
<210> 162	
<211> 100	
<212> DNA	•
<213> Homo sapiens 463295	
•	
<400> 162	
tgggaacttt accacttttt attattttgt teettttete tgetggaaay gegtttgetg	60
tcaatattat gccatatatc gctgtgccaa cttaagttgc	100
<210> 163	
<211> 90	
<212> DNA	
<213> Homo sapiens 500243	
<400> 163	
tototatoag tataacaaag gagtoagact tgatggoots taagggtoot tocaagtact	60
	_
ttaacattct agaaatttac cggacagaag	90
<210> 164	
<211> 100	
<212> DNA	
<213> Homo sapiens 715029	
<400> 164	
aagctcccc ctactcagtt gccttgaata ggaattatac tgaccaaaar ctagatttaa	60
aatgagaaac tttaacatac ccagttagac ccacctacac	100

<211>	100	
<212>	DNA	
<213>	Homo sapiens 715491	
<400>		
atgctg	tect agecaatate tgeteaceet etteagecat tecetgtacy geacattege	60
<b>.</b>		100
tecage	catg actgccatcc acggcccact catcacatac	100
	$\cdot$	
<210>	166	
<211>	100	
<212>	•	
<213>	Homo sapiens 718830	
<400>		
acatat	ttga tgacagagac acaatccctg tcctcaagga gctttcatty tagtagagaa	60
		100
gatgaa	aacc agtacagttt ggtaagttag atgatattgg	100
<210>	167	
<211>		
<212>		
	Homo sapiens 719185	
<400>	167	
tgagaa	gaga gagcaactgg aagggtgagg ggagatactg agttttagay ggttttcgaa	60
aaagca	agtc actgcccaac aagtgatgtt cagtagacaa	100
.010.	160	
<210><211>	168	
<212>		
<213>	Homo sapiens 723053	
<400>	168	
aagagg	aagg gggatgtgaa acatatacag tacattactc tctgcgatcy tcctctgtgc	60
tatcaa	caag caagaaaaaa gagatctgga gaaaacagat	100
.010	100	
<210>	169 100	
<211><212>		
<213>		
<400>		
gtcact	gggc agtccatgac tgattgccgt gctagttgtg tgggcactam tccctttaga	60
	•	
tcttta	gagg aggctgagtt cactttagac agagaggttt	100
.010	170	
<210>	170	
<211> <212>		•
<213>		
<400>	170	
	aggca gctaaaaaga caaataaata taactggtgg cacttactty agtgtctgag	60

•	
ctttaaagtc aaatttggct ctggcaggtc tcatctgaac	100
-010. 171	•
<210> 171	
<211> 100	
<212> DNA	
<213> Homo sapiens 726733	
<400> 171	
atgtttttga gtttgttttg ggaaatctac tttgcacatt ttgttaattr tctttgaact	60
caagetttee acaggtteaa etgetattet teecaettag	100
<210> 172	
<211> 100	
<212> DNA	
<213> Homo sapiens 728552	
<400> 172	60
taggaactga aagtacctga gctgtaaaac tagagaaacc tttcatatgk aaagggcaag	•••
ggaaaaagga gttaacatgt ataagcccga ggaagttgaa	100
<210> 173	
<211> 100	
<211> DNA	
<213> Homo sapiens 728612	
(213) HOMO Baptems /20012	
<400> 173	
agagagetgt actgggagee tggaaaagga aggtetaate ggcaatttty etttetetae	60
ccatgtattt ggaaaaata atgaattatc tggttgtggg	100
ccacgcacce ggaaaaaaa aogaabaaba oggaogogogog	
<210> 174	
<211> 100	
<212> DNA	
<213> Homo sapiens 729531	
<400> 174	
ccagtaggtg tcagctacgc acagcettte tetttetaaa tecatatttr taagaaaacg	60
and the state of t	100
gcctttcatc cataattttc cctagttaaa gggatcaatt	100
<210> 175	
<211> 100	
<212> DNA	
<213> Homo sapiens 730939	
<400> 175	
ttatttact tttttgtatt tgaggttttg atcttgctct gttttgtgtr agaaataaat	60
200000000 000000000 0000000000000000000	
tagaaattca gcattatttc cttctcaaat gtttagccat	100
-210- 176	
<210> 176	
<211> 100	

<212> DNA	
<213> Homo sapiens 734356	
<400> 176	
cgtgaagcaa aatctcattg gtaagtctga gttgcccaat gagggtttay aagaatacat	60
tcacgttttc taacttcatt accttttctt acttgaattt 1	100
<u>,</u>	
<210> 177	
<211> 100	
<212> DNA <213> Homo sapiens 734463	
<400> 177 gttaacatat ctgaaattca taggtaacac ctttcccaag ataatttagy tctggtatta	60
goodadaaa oogaaaaaaa ooggaaaaa oogaaaaa aaaaaaaa	
gatttagaat cttcatcctg gtttttttgg tctagtgttt	100
<210> 178	
<211> 100 <212> DNA	
<213> Homo sapiens 739485	
<400> 178	
tcaatagata ggttgatcac tctataaaat agtagtctgt tggcacttay gtgcgtaacc	60
agaggecate atgttetgag actgeceeag ggetttttgg	100
<210> 179 <211> 100	
<212> DNA	
<213> Homo sapiens 742737	
<400> 179	
ctggtaataa cttggttggt ctgaacccat gcacagctca gaagtgtgar tgagttaata	60
cccccaagga aaacttttga ctaattgggg gaagaaagca	100
<210> 180	
<211> 100	
<212> DNA	
<213> Homo sapiens 752331	
<400> 180	۲۵
cttccttaat gatcagaaca gagataaaaa tagtgtttag aatgtcctgr aagcctttgg	60
gaatgttaaa ttagaagaca tctattttga tacagatgtg	100
<210> 181	
<211> 100	
<212> DNA <213> Homo sapiens 756827	
<400> 181 aaacatttee ttetgtteea eccetggtat tettgtatgt gtgttetger taggagggee	60
~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	

tgggagttcc tgttgcttct ctgtgaaggg gggaggactt	100
<210> 182 <211> 100 <212> DNA <213> Homo sapiens 761057	
<400> 182 tcaggttcac atgtgcattt ggttcattat gtgtctttag tgttttaaty tggaacagtt	60
ctcttcccac ctttggatgt aagtggcaca tttgtttccc	100
<210> 183 <211> 100 <212> DNA <213> Homo sapiens 763807	
<400> 183 aagtaggcaa gaccctaaaa agaagccacc tgcctctccc agtccctccr ggaggactcc	60
aagagggcaa atctgcaggc agtgagctca gggccagcgg	100
<210> 184 <211> 100	
<212> DNA	
<213> Homo sapiens 765480	
<400> 184 agagaagtcc tgcagagggg aaatcataac tcggagtacc tcctaaatay acttgctata	60
agagaageee egeagagggg aaaceaeaac eeggageaco oootaaaaa, abbogoonea	
atctcacctc cttctatttt ttattttttt ttaattttca	100
<210> 185	
<211> 100	
<212> DNA	
<213> Homo sapiens 775251	
<400> 185	60
ageteetttt gatatteeet geeceagage teatgaceag aacceagagy tgatttaaaa	60
tattttgaaa aatggaggag gcagactgct cccagcagcc	100
<210> 186 <211> 100	
<211> DNA	
<213> Homo sapiens 784566	
<400> 186	
acacaggtcc atctgtggac attgtatatt taaaaaggcc tgtgggaacr atccatgaca	60
gggccactca tagaagaagc tgcaaatttt tagacggtgg	100
<210> 187	
<211> 100	
<212> DNA	

<213> Homo sapiens 877044	
100	
<400> 187	60
tecetetage tgetggggg eteaettgga tggteetaaa ggeaeceeey ttetteeaat	60
ggtggggtgc cctccagtgt tcacgggatg tgggctcata	100
·	
<210> 188	
<211> 100	
<212> DNA	
<213> Homo sapiens 878201	*
<400> 188	
cttccccgtt gtggactttg ggctgggcct tgcaatttct ctggccaagr gatgttggcc	60
3 3 3 3 3 5 5 6 6 6 6 6 6 6 6 6 6 6 6 6	
teetgeetge tgaggaeatg eeetgaagaa egtgtgeeeg	100
<210> 189	
<211> 100	
<212> DNA	
<213> Homo sapiens 879496	
<400> 189	C 0
gctatggttt tttgtttgtt tgtttagcac cagcaggcct gatagcaatr gcaattttta	60
	100
agaaaagaaa aaattagttg ggagaaaggt agtgtatgag	100
	•
<210> 190	
<211> 100	
<212> DNA	
<213> Homo sapiens 880456	
<400> 190	
tocattocat totgtgccat cocatgocac tocottocot gocactocaw gotattagaa	60
ttggagacag agaccccaat tacttttaat cctcagaatt	100
·	
<210> 191	
<211> 100 <212> DNA	
<213> Homo sapiens 885679	
C2139 Hollo Baptens CCCCC	
<400> 191	
caaggaaggg ctattataat totatttgaa actggctgca tttatcttcw tctctctcag	60
caaggaaggg ccaccaac cocaccaga accagacaca cocaccaca, coocacaca	
the beautiful through the section of	100
aagtcacatt teeettgeee ttggatgagg actggaatge	
	•
<210> 192	
<211> 100	
<212> DNA	
<213> Homo sapiens 886229	
<400> 192	
ccttaccctt gttagtggat agtggggatt taaaatagca gataatattr tatttagtaa	60
tagttaatat titttaagga titattatgi gigaagtaci	100
•	

<210>	193	
<211>	100	•
<212>	DNA.	
<213>	Homo sapiens 902336	
	•	
<400>	193	
aatttq	gaaa ctatagtacg tatcaggtat cgagaaggaa gtgaatataw ctcctataag	60
ctaact	ttct agtcttttt tcttgagcat tttttttca	100
00000		
	·	
<210>	194	
<211>	100	
<212>	DNA	
<213>		
\213 /	Nome Bapterin 303004	
<400>	194	
	atto otoagottta gaagaaaatg titttotaaa agggaacaay actotgaatt	60
LLGagi	actic creagettea gaagaaaatg teettetaaa agggaacaay acceegaate	•
	been between antenetter themselves	100
CCLLL	tttc tctaaaatgg aatcactttt tacaaatctc	100
	·	
<210>	195	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 912142	
<400>	195	
taatga	acag gatgeaeetg tteeatgaae eetetetete tteeetagar ggagaeaeee	60
agatco	rtcat cagtggcagc agacacctcc aagtctagga	100
<210>	196	
<211>	100	
<212>		
<213>	Homo sapiens 912265	
<400>	196	
ggcttt	gggc ataaagcagc cggctggctg ttggatctac agagtcaaay gtgaacaggg	60
aggcct	tacg aagcaagcct tgttttgctg agagctctgt	100
<210>		
<211>		
<212>		
<213>	Homo sapiens 924799	
<400>		
tctagi	tgaaa agactaatta ttttttgtat tatcctgttt ccaattttgy ggatcacttc	60
atact	aaact gaagacaaac teetatataa teettatgte	100
•	$\epsilon_{\rm c}$	
<210>	198	
<211>	100	
<212>		
<213>	Homo sapiens 975423	

<400> 198	
tagacttatc ttccaggctc gtgaagagtg cagtaatgct aatgttgagr gatgttaggc	60
acagatcaca tagcatatto ototoatcat cotgtoagga	100
<210> 199	
<211> 100	
<212> DNA	
<213> Homo sapiens 985627	
400 100	
<400> 199	60
tacattgaag accaagccat teteagtetg etetteaett etaattetam ggttattetg	00
· · · · · · · · · · · · · · · · · · ·	100
attgctcagg acaatgctgt ctgggaagac tgccccacag	100
<210> 200	
<211> 100	
<212> DNA	
<213> Homo sapiens 985716	
·	
<400> 200	
catgaagcag taaatggagc aattaacctg gggcctgaat gggaagaagk ttctggaggc	60
aacagggagt ctggagcaaa atatcaccag atgacaatat	100
anengaarae ebaaraera annoneera nogreeneera	
•	
2210. 201	
<210> 201	
<211> 100	
<212> DNA	
<213> Homo sapiens 994174	
<400> 201	
cttctcatca aataagtttg agtgttaagc atattgtgga atttggagar tggcactgag	60
tactattaga agcagtactg acgcaaatac attgcatact	100
*	
<210> 202	
<211> 100	
<212> DNA	
<213> Homo sapiens 996320	
.	
<400> 202	
tcctagtttt tgactttact gcctatatta cagaggaagt acggtatgtr ttgaagaaat	60
tagaagteea eecatagete aaatatgett eagtateett	100
tagaagttoa coodiagote aaatatgees cagaascoo	
2210- 202	
<210> 203	
<211> 100	
<212> DNA	
<213> Homo sapiens 997929	
<400> 203	
tocatgtgcc caacttggaa ttotottoto tgattttoco ataacttaay tgaagtgaga	60
actttgggga gttcaagcac tgtttcactt atttcctttt	100

<210><211><212><213>	204 100 DNA Homo sapiens 998188	
<400> ggcctc	204 ttat ggctgtattc tattagctct gggtttactg tgatattatr tatatttgtc	60
ttgggc	caaa cagcttgctt gcctcagttg aagtctaaaa	100
<210> <211> <212> <213>		
<400> ggcatg	205 aata tatatcaaag tttcttttt cccaaggata aacaaattgy cccagcacca	60
ttaatt	gatg tactaatgtg ctcaaaacac aaaataaatg	100
<210> <211> <212> <213>		
<400> atgcac	206 aaat gaaaggttee etgtgeteta teeetgeaca acateetter tetgetgtgg	60
gagaaa	tcat cttaaatact taaatgtaga attctgcagc	100
<210><211><212><212><213>	DNA	
<400> agtcta	207	60
tgaaaa	attgt attttattga actgcatatt aatgcattct	100
<210> <211> <212> <213>	DNA	
<400> tgtttt	208 cacta aagttgattg aaattagtta gttgatattc atcctaccak attttgagtt	60
agagaa	aattt agatagcaat atttgcacct aaactatttt	100
<210><211><212><213>	100 DNA	

400	
<400> 209 actacaacag aaacacactg tcacacttgc tagaatataa ctgtacttgr gcttctcctt	60
tcctgtgaag tagtgctggg ctttctagag tttaattctc	100
<210> 210	
<211> 100	•
<212> DNA	
<213> Homo sapiens 1041111	
<400> 210	
ctgctgaaag atgaatcact atgaatttgg caggggaatg gaaagatcay gagcagctca	60
gcaatacagt ggccccatct agagatggtc acccaggact	100
<210> 211	
<211> 100	
<212> DNA	
<213> Homo sapiens 1240709	
<400> 211	
ggacttccag gcaggatgct gaaggataaa tgtggacacg ggtgagggar gcacctgtgg	60
aatctgcagg gaggctggac tccagtcctg atgtcggccc	100
<210> 212	
<211> 100	
<212> DNA	
<213> Homo sapiens 1259291	
<400> 212	60
ggcattatca totoaaaggg caaaagaagg atggotaaac cattggaaay gtggattogc	60
tatttttgct atggtcccat ttagccccta cccatgtctg	100
<210> 213	
<211> 100	
<212> DNA	
<213> Homo sapiens 1324758	
<400> 213 tgaaaaatgc ttcccagaaa tatgctcata tcattcatcc attgacaatm atcatttgtt	60
tgaaaaatgo ttoocagaaa tatgotoata teattoatoo attgacaatii attatotigot	00
ggacatcaac tgcttgtact acattagctt atgacatgaa	100
<210> 214	
<211> 100 <212> DNA	
<213> Homo sapiens 1361107	
<400> 214	
gtacttattc ccacagccaa cttcaaatga cccctccct ggcccccccm caagaaaaaa	60
acagggcagc tgaaaagttt gcttaaactt cttaatactt	100

<210> 215 <211> 100 <212> DNA <213> Homo sapiens 1367990	
<400> 215 ctttaagatc catgtaaaaa tattgcaaat gagggatttt agtagtaagr aaaatttgcc	60
aatttttact cgattagttt ggctattttt atatattctt	100
<210> 216	
<211> 100 <212> DNA <213> Homo sapiens 1395381	
<400> 216 - ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca	60
atgaagaaat taaactggac tgtcccaaag aagatgaaaa	100
<210> 217	
<211> 100 <212> DNA <213> Homo sapiens 1466386	,
<400> 217	60
ctccatgtta ttcctgacaa acagttgtcc catgaacaac atcccaacar atcatacttt	60
tcaagtagca attatcagag ggtgctaata attccaggtt	100
<210> 218 <211> 100	
<212> DNA <213> Homo sapiens 1473138	-
<400> 218 gctcttaagt caagtcactt gtggaacttg ccatataaac aatggtttgy aatgtaatgt	60
gttgaagtat aaaatagagt tgtgttcaaa gtactggggt	100
<210> 219 <211> 100	
<211> 100 <212> DNA <213> Homo sapiens 1541306	
<400> 219 acgggcgagt atccatggtg ctgattcttc cagttgtccc aaaacaaaty ttcaatgaat	6 0
tetgaggeee caaggettee ettgttggee aagteaetga	100
<210> 220 <211> 100	
<212> DNA <213> Homo sapiens 1541515	
<400> 220	,

cagaagttat ttgctcttgt gcataattga aaacaaatgt tctgtgttgk gtacaattat	60
tttgatctta aaggctatgc ttcgtagaaa aagagaaggt	100
<210> 221	
<211> 100	
<212> DNA	
<213> Homo sapiens 1544656	
<400> 221	
cagettgtee cegttaggtg acaataagca gacagaagag ccaagcagcy atggaggaaa	60
atcaggtacc taaatttagc tcaactcctg cctcattcag	100
<210> 222	
<211> 100	
<212> DNA	
<213> Homo sapiens 1618967	
<400> 222	
ttttagette ttttetaete teteaatett atcegggett gtttgggggy gaactettta	60 -
fiftagette tecterate coccarette accoggisted Secusing Secusing	
tettttatag aagataaagt ttggttttat taaatttttt	100
•	
<210> 223	
<211> 100	
<212> DNA	
<213> Homo sapiens 1806913	
<400> 223	60
tttaaaatgt tactttattt tagaataagg tagagttgaa aattgtcagr aatacaaaca	60
ttatttaact atttaatcct ttatatatgt gaagttttaa	100
tracticaact actiaacter tracatacyt gaagetteaa	
<210> 224	
<211> 100	
<212> DNA	
<213> Homo sapiens 1806995	
400 004	
<400> 224 ataatactta attttttggc tatgaagagg agaaaaatca tcattttcty atttttctca	60
alaatacita attititigge tatgaagagg agaadaatta teattitity attitiona	
tcaggggact ctgaagggct tgggaggagt gatctgccca	100
<210> 225	
<211> 100	
<212> DNA	
<213> Homo sapiens 1883252	
<400> 225	
ataaatgtot ggtttaaagg agaaatcaat actgttatto ototocaaar attaagcoca	60
and and a second a	
gtctaggaag agaagattgg agttttagac ttgattagtt	100

	·	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 1939382	
<400>	226	
acatta	ggca agttatatta cttccttgct ccttactatc ttcatctgtm aaatggggaa	60
		100
agtatt	taac atagggccaa ctacaagcaa attggctttt	100
<210>	227	
<211>	100	
<212>		
	Homo sapiens 1957979	
72207	none Baptons and a second	
<400>	227	
	gtgg tatatactgc caaaaatgta tetteetaat tgteagtety aaaaaaggta	60
J	5.55	
gtttcc	aaac tacctctttt ttaaacatct atcagttttt	100
<210>	228	
<211>	•	
<212>		
<213>	Homo sapiens 1964211	
400	000	
<400>	zzo getac tegggagget gaggeaggat aategtttga aeeetggagr tggaggttge	60
LCCLAS	derge cedadagger daggeaggar aaccaesaga accaesagast saasaasa	
anthrac	getga gattgtgeea etgeacteea geetggaeta	100
. agugas	30034 3400343004 0030400000 300033	
	•	
<210>	229	
<211>	100	
<212>	DNA .	
<213>	Homo sapiens 1966625	
<400>	229	60
atttag	gtccc agaagggcgt gaggacatca tgtagacaca aaagagcatr caacagcaaa	00
		100
ttatt	ggggg taaatattta atatcttgcg tctggggaac	
<210>	230	
<211>	•	
<212>	•	
<213>	Homo sapiens 2006120	
•		
<400>	230	
agaac	cagag gctgggaagg acaagtcacc tgcttgtgag tggcaggggm aggattggga	60
		100
gcagt	ggact cggtctcagg actgcggccc tcaggaccaa	100
.210	. 221	
<210> <211>	·	
<211>		•
<213>		
-3-27	• • • • • • • • • • • • • • • • • • •	
<400>	231	
ggttg	rtgggg ttctccaggt gctctgtcgc ccaggctgga gtgcagtggy gcgatctcgg	60

ctcactgcaa cctcctcctc ccgggtttaa gtgattcttg	100
<210> 232 <211> 100 <212> DNA <213> Homo sapiens 2012661	
<400> 232 aatggttgca tattattgtc aagtgggtgt taccacatga atatttacak ttgttcagct	60
catctgtgaa tttcttctct tcattcccag gactcaccat	100
<210> 233 <211> 100 <212> DNA <213> Homo sapiens 2023675	
<400> 233 atgcaaatgt gttcatggtc atgggtggta cactggctag ttttaaatay gagccctcag	60
agctgcagga gaaggccaca tgaatgcctc caccacagac	100
igotgoagga gaaggoodoa ogaaaggoodo odoaangoo	
<210> 234 <211> 100 <212> DNA <213> Homo sapiens 2042417	
<400> 234 ggataatgtt atatatacct tgtggatata caccttgtgg tgtgtgtatr tatatatata	60
tatatatata catacacaca caccttgtgg agtgcagtgt	100
<210> 235 <211> 100 <212> DNA <213> Homo sapiens 2048326	
<400> 235 aaaaagaatg gcatttggca agatgagact ttggttccta actgctctar gacctaggct	60
ggettgaggt tcaaagtgce ggttactete catttgactg	100
<210> 236 <211> 100 <212> DNA <213> Homo sapiens 2052832	,
<400> 236	60
gggatgatac actctttggt cctaccaaac agctttcatc ttgagatcgy cctccatctt	100
cattgtggcc cttcctctca gttctctttt gtgtggtccc	100
<210> 237 <211> 100	

<212> DNA	
<400> 237 agcagtggtg agaggggaag agatttttct atgccatgtt tcctgacagk agcagaacct	60
ageageggeg agaggggaag agaeeeeee aegeeaegee eeeegaeagk ageagaaeee	
gicticagac cigaaacaci ictccittia tagicactat	100
<210> 238	
<211> 100 <212> DNA	
<213> Homo sapiens 2064168	
<400> 238 getggacace tgggagggge agcetgtetg tettetgete tgccetttey tgetgtgtte	60
tggaagcata ggcaagcatt ccacagtatc ctgcctgcag	100
<210> 239	
<211> 100 <212> DNA	
<213> Homo sapiens 1412302	
<400> 239	
tcacataatg gaagaatggc ccggcctaga ccagaggttc ctaattaggr gtccatgagc	60
	100
cacgttcaat tatatatatt ttttcttctg gtatttttct	100
<210> 240 <211> 100	
<212> DNA	
<213> Homo sapiens 291030	
<400> 240	
agtgtgtcac tectaactet tettacetee tttetttggt ttaaggtetm ttagtgtetg	60
tactgggtag tgttttcaca gctttctcct ctgacatggc	100
<210> 241	
<211> 100	
<212> DNA	
<213> Hómo sapiens 713970	
<400> 241	•
ccaaacagca ctcagaaaat cataaaatgg caagcaaaat gagcaattcy gcacgcagca	60
cgtgacatgg agtatgagct cgcatttgcc gaaggagcaa	100
<210> 242	
<211> 100	
<212> DNA	
<213> Homo sapiens 718830	
<400> 242	
acatatttga tgacagagac acaatccctg tcctcaagga gctttcatty tagtagagaa	60

gatgaaaacc agtacagttt ggtaagttag atgatattgg	100
<210> 243 <211> 100 <212> DNA <213> Homo sapiens 723053	
<400> 243 aagaggaagg gggatgtgaa acatatacag tacattactc tctgcgatcy tcctctgtgc	60
tatcaacaag caagaaaaa gagatctgga gaaaacagat	100
<210> 244 <211> 100 <212> DNA <213> Homo sapiens 725979	
<400> 244 gtcactgggc agtccatgac tgattgccgt gctagttgtg tgggcactam tccctttaga	60
tetttagagg aggetgagtt caetttagae agagaggttt	100
<210> 245 <211> 100 <212> DNA <213> Homo sapiens 726733	
<400> 245 atgtttttga gtttgttttg ggaaatctac tttgcacatt ttgttaattr tctttgaact	60
caagetttee acaggtteaa etgetattet teecaettag	100
<210> 246 <211> 100 <212> DNA <213> Homo sapiens 728612	·
<400> 246 agagagctgt actgggagcc tggaaaagga aggtctaatc ggcaatttty ctttctctac	60
	100
ccatgtattt ggaaaaaata atgaattatc tggttgtggg	100
<210> 247 <211> 100 <212> DNA <213> Homo sapiens 734356	
<400> 247	60
cgtgaagcaa aatctcattg gtaagtctga gttgcccaat gagggtttay aagaatacat	
tcacgttttc taacttcatt accttttctt acttgaattt	100
<210> 248 <211> 100	

<212> DNA

<213> Homo sapiens 742737			
<400> 248			
ctggtaataa cttggttggt ctgaacccat gcacagctca gaagtgtgar tgagttaata	60		
coddonaean cooddoodda codaaccear daaadaaaa daadaaaa adadaaaaa			
cccccaagga aaacttttga ctaattgggg gaagaaagca	100		
•			
<210> 249			
<211> 100			
<212> DNA <213> Homo sapiens 752331			
VZIJV NOMO Baptens /JZJJI			
<400> 249			
cttccttaat gatcagaaca gagataaaaa tagtgtttag aatgtcctgr aagcctttgg	60		
gaatgttaaa ttagaagaca tctattttga tacagatgtg	100		
<210> 250			
<211> 100			
<212> DNA			
<213> Homo sapiens 756827			
<400> 250	60		
aaacatttcc ttctgttcca cccctggtat tcttgtatgt gtgttctgcr taggagggcc	60		
tgggagttcc tgttgcttct ctgtgaaggg gggaggactt	100		
tgggagttee tgttgtteet tegtgaaggg gggaggaett,			
<210> 251			
<211> 100			
<212> DNA			
<213> Homo sapiens 761057			
<400> 251			
tcaggttcac atgtgcattt ggttcattat gtgtctttag tgttttaaty tggaacagtt	60		
Couggeood degegeneer ggreenes gages and			
ctcttcccac ctttggatgt aagtggcaca tttgtttccc 100			
<210> 252			
<211> 100 <212> DNA			
<213> Homo sapiens 879784	,		
<400> 252			
gcaggaccgc gcacagagta ggcagcattt gtgtgtgcct cgcacgtgtr tggtggcact	60		
h to a character casteraine consentes	100		
gtatgcactt gtagatactc aaatacgtag ggaaagatga	100		
<210> 253			
<211> 100			
<212> DNA			
<213> Homo sapiens 881929			
.400. 253			
<400> 253 acceagtggt tetgagetet ggeeetteta etgeetettg ceagetetgk gteteageet	60		
tectatetgt gagttagaca ecaggtaget ggaggggaaa	100		

<210><211><211><212><213>	254 100 DNA Homo sapiens 902336	
<400> aatttg	254 gaaa ctatagtacg tatcaggtat cgagaaggaa gtgaatataw ctcctataag	60
ctaact	ttot agtottttt tottgagoat tttttttca	100
<210>	255	
<211> <212>	100 DNA	
<213>	Homo sapiens 912265	
<400>	255	60
ggcttt	gggc ataaagcagc cggctggctg ttggatctac agagtcaaay gtgaacaggg	60
aggeet	tacg aagcaagcct tgttttgctg agagctctgt	100
<210> <211>		
<212>		
<213>	Homo sapiens 915941	
<400>	256	
eggets	gegte tatggteatg acgtetgaea gagegteeae eegtettegm eaggaeteta	60
tggtto	ettac gegegeagae agacegeeta tataageeat	100
<210>	257	
<211> <212>		
<213>		
<400>	257	
tctag	tgaaa agactaatta ttttttgtat tatcctgttt ccaattttgy ggatcacttc	60
atact	aaact gaagacaaac tootatataa toottatgto	100
<210>	258	
<211>	•	
	DNA	
<213>	Homo sapiens 985627	
<400>	258	
tacat	tgaag accaagccat totcagtotg otottcactt ctaattctam ggttattotg	60
attgc	teagg acaatgetgt etgggaagae tgeeceacag	100
<210≈	259	
<211>		
<212>		
<213>	Homo sapiens 997929	

<400> 259 tccatgtgcc caacttggaa ttctcttctc tgattttccc ataacttaay tgaagtgaga 100 actttgggga gttcaagcac tgtttcactt atttcctttt <210> 260 <211> 100 <212> DNA <213> Homo sapiens 998188 <400> 260 ggcctcttat ggctgtattc tattagctct gggtttactg tgatattatr tatatttgtc 100 ttgggccaaa cagcttgctt gcctcagttg aagtctaaaa <210> 261 <211> 100 <212> DNA <213> Homo sapiens 998262 <400> 261 ggcatgaata tatatcaaag tttcttttt cccaaggata aacaaattgy cccagcacca 100 ttaattgatg tactaatgtg ctcaaaacac aaaataaatg <210> 262 <211> 100 <212> DNA <213> Homo sapiens 1008520 <400> 262 agtotatota ttoatttatt tttootocaa acctgottat tttttttaar taccaataco 60 100 tgaaaattgt attttattga actgcatatt aatgcattct <210> 263 100 <211> <212> DNA <213> Homo sapiens 1009127 <400> 263 tgttttacta aagttgattg aaattagtta gttgatattc atcctaccak attttgagtt 60 100 agagaaattt agatagcaat atttgcacct aaactatttt <210> 264 <211> 100 <212> DNA <213> Homo sapiens 1016491 <400> 264 actacaacag aaacacactg tcacacttgc tagaatataa ctgtacttgr gcttctcctt 60 100 tcctgtgaag tagtgctggg ctttctagag tttaattctc

<210> 265	
<211> 100	
<212> DNA	
<213> Homo sapiens 1019178	
<400> 265 · · · · · · · · · · · · · · · · · · ·	
tcaagtgtat tttaccacaa aggcctcttc agggacattt ataacatatm ccagatgtag	60
ttttccaagg agagtgcttt gagaaacact gaccaagcct	100
212	
<210> 266	
<211> 100	
<212> DNA	•
<213> Homo sapiens 1240709	
<400> 266	
ggacttccag gcaggatgct gaaggataaa tgtggacacg ggtgagggar gcacctgtgg	60
32accccord 2cr32ac2cc 2ar32acras c2c32acras 23c2c22 = 2 22	
	100
aatctgcagg gaggetggac tecagteetg atgteggeee	100
<210> 267	
<211> 100	
<212> DNA	
<213> Homo sapiens 1375224	
2213> NOMO BADIENB 13/3224	
<400> 267	
aataagcatt ctcaattctg caattcagtg gcattgcata tatcttaggy ggtgtttatg	. 60
acttcatatt tataatttgg cattcatgtg tttatcttct	100
acttcatatt tataatttgg cattcatgtg tttatcttct	100
acttcatatt tataatttgg cattcatgtg tttatcttct	100
	100
<210> 268	100
<210> 268 <211> 100	100
<210> 268 <211> 100 <212> DNA	100
<210> 268 <211> 100	100
<210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381	100
<210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268	
<210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268	
<210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381	
<210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca	
<210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268	60
<210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca	60
<210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa	60
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269</pre>	60
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100</pre>	60
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa </pre> <pre><210> 269 <211> 100 <212> DNA</pre>	60
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100</pre>	60
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa </pre> <pre><210> 269 <211> 100 <212> DNA</pre>	60
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa </pre> <pre><210> 269 <211> 100 <212> DNA</pre>	60
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100 <212> DNA <213> Homo sapiens 1455576 <400> 269</pre>	60 100
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa </pre> <pre><210> 269 <211> 100 <212> DNA <213> Homo sapiens 1455576</pre>	60 100
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100 <212> DNA <213> Homo sapiens 1455576 <400> 269 acatgaaaga atttttggcc aggcaatgga gatagacagt accattagcy gaaaacaata</pre>	60 100
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100 <212> DNA <213> Homo sapiens 1455576 <400> 269</pre>	60 100
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100 <212> DNA <213> Homo sapiens 1455576 <400> 269 acatgaaaga atttttggcc aggcaatgga gatagacagt accattagcy gaaaacaata</pre>	60 100
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100 <212> DNA <213> Homo sapiens 1455576 <400> 269 acatgaaaga atttttggcc aggcaatgga gatagacagt accattagcy gaaaacaata agacaatgac atgccatact agaaatatta aatcattctt</pre>	60 100
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100 <212> DNA <213> Homo sapiens 1455576 <400> 269 acatgaaaga atttttggcc aggcaatgga gatagacagt accattagcy gaaaacaata agacaatgac atgccatact agaaatatta aatcattctt <210> 270</pre>	60 100
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100 <212> DNA <213> Homo sapiens 1455576 <400> 269 acatgaaaga atttttggcc aggcaatgga gatagacagt accattagcy gaaaacaata agacaatgac atgccatact agaaatatta aatcattctt</pre>	60 100
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100 <212> DNA <213> Homo sapiens 1455576 <400> 269 acatgaaaga atttttggcc aggcaatgga gatagacagt accattagcy gaaaacaata agacaatgac atgccatact agaaatatta aatcattctt <210> 270</pre>	60 100
<pre><210> 268 <211> 100 <212> DNA <213> Homo sapiens 1395381 <400> 268 ctaaagttcc cttgtcctca gagtgagagt tcatagggct catttcaaar ctctcagcca atgaagaaat taaactggac tgtcccaaag aagatgaaaa <210> 269 <211> 100 <212> DNA <213> Homo sapiens 1455576 <400> 269 acatgaaaga atttttggcc aggcaatgga gatagacagt accattagcy gaaaacaata agacaatgac atgccatact agaaatatta aatcattctt <210> 270 <211> 100</pre>	60 100

·	
<400> 270 tgctctgtat tggctgtggg gtgggggtgg cggtggaact gacccctggs gtctgcattt	60
ctcaggetee cagggetgtg getgaetttg gecaatggga	100
·	
<210> 271	
<211> 84 <212> DNA	
<213> Homo sapiens 242143	
<400> 271	
ctccaggcga ttcagatgca ggcttaagtc tgagaaccac tgttgtaggy ggttgtggtt	60
ggccccgttc ttagagtaga attc	. 84
<210> 272	
<211> 100	
<212> DNA	
<213> Homo sapiens 593930	
<400> 272 agcattgctt tgagaataga aatagtcagg aatggttggc tcatgcctgy aatcccaaca	60
ctttgacaag ctgagactat cacttgagac cacgagttca	100
<210> 273	
<211> 100	
<212> DNA <213> Homo sapiens 732148	
(213) Notice Supreme / Salie	
<400> 273 cctcatgatt tgttctgcca cttttagaat actaaatgct cacttatcty taagatctta	60
	100
tctagagcta tcatattaag tttaggacta ttcctctacc	100
<210> 274	
<211> 100	
<212> DNA <213> Homo sapiens 1377478	
<400> 274 ggtaaaatac tgacttgggt cacatgatct acctcaccat agggagaggk atactcactg	60
	100
acctggtcag agccatacac ccaaggagat aacctttgcc	100
<210> 275	
<211> 100	
<212> DNA <213> Homo sapiens 2011741	
<400> 275	
<400> 275 aagccaatat totatgttot atttotoagt ttotgtoato aatagatatm taggccotto	60
tcccacattc actgaatatc tacagccaac ctatccaata	100

	A.T.	
<210>	276	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 218855	
<400>	276	
	atca tgatcttagg tcttctgata tttctaactt caatcctcaw tgctgttatg	60
gattaa	atct tttaacccac ttgctattat attgatggga	100
gaccaa	acce elegacedad englished and Janggan	
-10		
<210>	277	
<211>	100	
<212>		
<213>	Homo sapiens 347867	
<400>	277	
ttaaac	ttcc ttccagttag ttttccctgc ctacctcttc cttgctgtcs tttcagactg	60
trator	ctgc ctacaagaga cagtggctgg gattcgggaa	100
cgacci		
	0.00	
<210>	278	
<211>	100	•
<212>	DNA	
<213>	Homo sapiens 523200	
<400>	278	
cttgad	caagg gtggcagaga ttaaaaaacc ccacgcttta aacaaacatm tttgggggtg	60
actate	ggcac cttgaaaaac ccacaaccag cttgacatga	100
goodo	5,5000 00050000000000000000000000000000	
<210>	279	
<211>		
<212>	DNA	
<213>	Homo sapiens 734908	
<400>	279	
ccctg	gctaa ggacgccacc ggcctccagg acagggccct gcctcacagk gtgctgcctc	60
_		
cccac	tggct ccgcagcaga agcttcagca ggccattcca	100
		٠,
<210>	280	
<211>		
	DNA	
<213>	Homo sapiens 743221	
<400>	280	60
tccac	ctata ccatgettta aaatgagtat ttgggcagge tgetetager tetatgtaca	00
		100
gtgad	agcaa aaccttcata ttatgaaggg ggccactcgg	100
<210>	281	
<211>	•	
<212		
<213>	Hollo Babteria 200022	
	201	
<400;	281	

cettececca aaacaacata ttatetggte tgteatagaa aetgaaatty geatattatt	60
cttccagttg gattagaagc tatgcattaa ttaatatgca	100
<210> 282 <211> 100	
<212> DNA	
<213> Homo sapiens 992864	
<400> 282 ttcactcttt tgggaaattt acttacctct tagcttttgg ttccaggatr aatatcacta	60
	100
ataggogtta gtotgotoaa ttaaattoaa taaatatgaa	100
	•
<210> 283 <211> 100	
<212> DNA	
<213> Homo sapiens 1076556	
<400> 283	60
tagaacageg tgcacatetg taagcatetg aagactacae teaagtgeer gatatatagt	
tattetgaaa acatgtaaca tgtagcaaat gggtttttet	100
<210> 284	
<211> 100 <212> DNA	
<213> Homo sapiens 1360554	
<400> 284 tototaaaat caatgacota aattgagaga ataotgatta ttaaaagaar ataatgtoaa	60
	100
caaagatgtc agaaatgtta ttaaggtgaa attgtaacca	100
<210> 285 <211> 100	
<212> DNA	
<213> Homo sapiens 1530044	
<400> 285	
ccacatttct cctagttttg tagcaattgc aacacagata tgttaagtgw gtataaagga	- 60
aaacgttatg gctatttcat cagcaagtgt tttttgccca	100
<210> 286	
<211> 100	
<212> DNA <213> Homo sapiens 1985080	
<400> 286 · ggagagaatg gagggaaaca aaggatgtga aaacccagag ggaatgaggr taagtggctc	60
	100
actgtacaat ccacaagtca gaaggtatag ggatgacagg	100

•	
<211> 100	
<212> DNA	
<213> Homo sapiens 2039055	
<400> 287	
aatttottga agtottggaa atgaaaattt toagaggtot ttttatgacm ttatagtoot	60
	200
tttggttaaa gcaaaagaat agactgtttt attttgcaaa	100
·	
	•
<210> 288	
<211> 100	
<212> DNA	
<213> Homo sapiens 327219	
<pre><400> 288 cttccttcct tccttccttc cttccttcct tccttccttc cttccttccy tccctctct</pre>	60
direction footies freezest confermed accompany	
tototttott totttottto ttttattttt gttttgggtg	100
EGEGEEEGEE EGEEEGEEEGE GEEEGEGGGGGGG	
<210> 289	
<210> 289 <211> 100	
•	
<213> Homo sapiens 379601	
-220-	
<220>	
<pre><221> misc_feature <222> (100)(100)</pre>	
<223> n is any nucleotide	
<400> 289	
tttttagctg caatcttttc acggaaaaac aatgtaaaag gcataacatr gagaatagag	60
coolinging carcologic auggantance mangements growing growing	
atggaaaaga aagatacaga cttgtaattg cctgacattn	100
uoggaaaaga aagaaaaga oo agaaa oo ag	
<210> 290	
<211> 100	
<212> DNA	
<213> Homo sapiens 398158	
•	
<400> 290	
tcacttcttt caagtctctg ttcaaatgtc cccttattag caaggccttm tttgacctcc	. 60
ctatgtagaa cagtaaaagc ccacccctt tttcctccta	100
<210> 291	
<211> 100	
<212> DNA	
<213> Homo sapiens 492933	
<400> 291	. 60
gtgaggacct ttttactttt tgaaaaatgg caagccaagg acttaacaay tgccaactaa	. 00
the section of	100
aacacaaatc cttcacttca caaaataaaa ttagatgatc	100
•	
•	

<211> 100	
<212> DNA	
<213> Homo sapiens 508118	
<400> 292	60
gtcacagtta agccacagta tcaaactggt gaggcagaaa ggggagagtm atgtaactgg	00
tagtcaagaa acctggattc tatgattaat actaaaacta	100
tagecaagaa acceggaece taagaceaac accaaaa	
<210> 293	
<211> 100	
<212> DNA	
<213> Homo sapiens 689594	
100 000	
<400> 293 aataccatca ctttccctac tttgatggca taccttatag tctttcttcy cttcacatgt	60
adeacoacca cocceced conductation encouraged accountable	
agtttttctt attagtatgt aagaagcagc ataaggtagt	100
<210> 294	
<211> 100	
<212> DNA	
<213> Homo sapiens 699863	
<400> 294	
<400> 294 taaggtacta taattgatat acttgtatac agggtcctta taaaatacar ctcataccta	60
caaggeacea caacegacas acceguates agggeetes and	
aatccagata tttaataatt gggcccatta agtccaatca	100
<210> 295	
<211> 100	
<212> DNA	
<213> Homo sapiens 815147	
<400> 295	
gtgtgtatgc ttttgtatat gtatgtgtat atatatatat atataatttw aaaaatcttc	60
tctatggctt tgataatget gtttccccat catgtctata	100
<210> 296	
<211> 100 <212> DNA	
<213> Homo sapiens 878213	
10.10 Paperson	
<400> 296	
gtcatgaaaa ggttaatcta caatgcactt agacagctaa taataagtar aacatttcca	60
	100
aagaaatagg aacctgggat tgtctctttt taataaatta	
<210> 297	
<211> 100	
<212> DNA	
<213> Homo sapiens 914432	
-	
<400> 297	-
atagttcact gctaaattat ctttggtgca aagtagggta caaaacaaar aaagtcaaaa	60

ccacaaatga tcaattccta attatgcaga atagccacca	100
<210> 298 <211> 100 <212> DNA <213> Homo sapiens 946685	•
<400> 298 gaacccaggc aaggggcatt tgctgactac agtgtgtctt tcagcaaacr cagccggagg	60
cccccaccc atggaaggga cagccttgcc aagaaaactg	100
<210> 299 <211> 100 <212> DNA <213> Homo sapiens 957452	
<400> 299 tatacaggat ggggattcca gacaatcaga gtatgatgtt ctcttgtcay tgatcacatt	60
cactagtgac aactaattag cacatccgag taatcaaaaa	100
<210> 300 <211> 100 <212> DNA <213> Homo sapiens 1004660	
<400> 300 gcaccgcctg totataceag tgctgatccc acctctccc tcccccttcy gccctcctgc	60
ctttgcaatt ggctgtttcc ctccagcttt gtccatcctc	100
<210> 301 <211> 100 <212> DNA <213> Homo sapiens 1011024	
<400> 301 agatttggga atcactgaat tgaactgccg gccattggtg gctggtgagr ctgtcctggc	. 60
agtgatgcag tattctggtc cctgtgccag cactgatggg	100
<210> 302 <211> 100 <212> DNA <213> Homo sapiens 1015736 <400> 302	45
atagtttgac aggaggaacc atccaggctg taattgtggt aaggaatacm tggttgaatt tcactttta tgtctttatt ttggaattct gtagctaggt	60 100
Coaccicia (gioriciali leggaacce giagelagge	
<210> 303	

<211> 100

<212> <213>	DNA Homo sapiens 1198718	
(213>	nomo Baptens 1190/10	
<400>	303 tatt attotataag aacaagatto agatttagga taaagotots tgoatataga	60
agctact	tgaa tggtaaattg ataaagttaa tctttaatat	100
<210>	304	
<211> <212>	100 DNA	
<213>	Homo sapiens 1377313	
<400>	304	
	agat taaatcacat tcagaagaaa acaacatttg ggagaagacr tctagagaca	60
		100
aaaatt	gtca aaagtttatt gtaaggctat gaaaaatgag	
<210> <211>	305 100	
<212>	DNA	
<213>	Homo sapiens 1384866	
<400>	305	
	ctta aactatcaaa tetttgatga ggtatttett tgactaagtr taaacacaaa	60
+ a+ a+ +	agac tagaatgcat caatcctcaa ctgtccaccc	100
CCCCCC	agac cagaatgeat caaceeecaa otgeeeacoo	
<210>	306	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 1418978	
<400>	306	
	ggca agatcagatt acctcaaggt cacattactg ggcaaagagy tagacttctg	60
caggg	caga agaggtttgg gagctcattt catccattcc	100
<210>	307 100	
	Homo sapiens 1936999	
	,	
<400>	307 :tttt ttgagatgga gtotcagtot gtcacccagg ctggagtgor gtggcatgat	60
	titti tigagatgga gitticagitti gittacootagg ooggagoger goggenegat	
cttggc	etcac.tgcaacetet gcateceage ttcaagcaat	100
<210>	308	
<211>		
<212>		
<213>	Homo sapiens 1955983	
<400>		
caatat	taaat ccatggcaga aagattttta aaaagaacta tttttataar tataaaataa	60

aacaattaga aaacattgtt ctaacttaat tggcagcatt	100
<210> 309	
<211> 100	
<212> DNA	
<213> Homo sapiens 2031262	
22137 NONO Sapiens 2031202	
<400> 309	
attotgatag agaccatatg acctocaagt ggattagott toaatgaatk otacaaacca	60
	100
gaagagtett teeceeatet teatetteat tetetaaett	100
<210> 310	
<211> 100	
<212> DNA	
<213> Homo sapiens 2047198	
<400> 310	
tggtagcaaa cctaaagtca aatttttggc tcaattactt caaccaacay agatctaaat	60
agcttatttt ttcttttatt cgggcctgag ctcttattct	100
<210> 311	
<211> 100	
<212> DNA	
<213> Homo sapiens 2049590	
<400> 311	
ttatgttaag gacatttatg atatattata tataatatac cataggtcam tagatacaaa	60
aacatatatg tcaaggatat atataacata caatacatgt	100
<210> 312	
<211> 100	
<211> 100 <212> DNA	
<213> Homo sapiens 2052972	
400 210	
<400> 312 table table and appropriate transport of the contract of t	60
tatatgtaac gacctatttg cagacacaca agtctcaaag aggggtttay atgcagtgac	
ttttggtaca atccctgtgg ttagtggaag catttatacc	100
<210> 313	
<211> 100	
<212> DNA	
<213> Homo sapiens 518116	
<400> 313	60
taaggtgaga gaggcagaag gagggtggaa agaatggcgt ttttggtgcr gtgcaaacta	00
atttgcccaa actgaaaaag ctggtgctct caccattatg	100
<210> 314	
<211> 100	
<212> DNA	

<213> Homo sapiens 546445	
<400> 314 cagtccttcc gccgccgggg ctaacacaat agtgaaagct gcgctcctgr tcgtcggcag	60
gctggcactg ttatacagga aaagggtcgt gtgcgacctt	100
<210> 315 <211> 100	•
<212> DNA <213> Homo sapiens 645142	·
<400> 315 tcaacatatt tggtattcca tggagagttc tcttagtgga gatagttcas gaaggggttt	60
attttcttaa tttgctttct cagttataaa atcttcccaa	100
<210> 316 <211> 100	
<212> DNA <213> Homo sapiens 715005	
<400> 316 gcttttaggt ctttaaagca gcacatgatt ttcaagtgag ttattcaaty aatcgtttct	60
gtatcactct gatgcccctt tggactgtct gcttattgaa	100
<210> 317	
<211> 100	
<212> DNA	
<213> Homo sapiens 763003	
<400> 317 ctgagctcag ccagggacca ggaaccaggg accagagtca ggggagccgk ctagaacaga	60
agggcacgtc ggtgctaggc tgggggaggc gcagagatgg	100
<210> 318	
<211> 100	
<212> DNA <213> Homo sapiens 768312	
<400> 318 tctgagctgc atcctgaagg ccacagacaa ggcacagaaa gtcactccar aaagagggaa	60
caggatgtac agaatcatgg aacatctact gtgtgccagg	100
<210> 319 <211> 100	
<211> 100 <212> DNA	
<213> Homo sapiens 873893	
<400> 319	
attgaaaaca cctcctggca aagaaggtgt gatttccacg ggacaatgck gatgactttt	60
taattaagat tettgtette ageetgetge ettaceaggt	100

<210><211><211>	320 100 DNA	
<213>	Homo sapiens 913315	
<400>	320	
	gaagg ggaggccaca ggagttacag aggactctgg aaatacagar agatctgtga	60
tttact	cagt gggtatttga tagactaaga aggcaaattc	100
<210>	321	
<211>	100	
<212>		
<213>	Homo sapiens 974828	
<400>	321	
	agtit tecacatgga attectette tigeactitg geetaageay geattitite	60
3		
taatg	gtgga aaatattaaa cagacttttg ctttagaagg	100
<210>	322	
<211>	•	
<212>		
<213>		
	•	
<400>		
tataai	tacc cttacctaat tttctaatac agtcctaata ttttctagar ccctctccaa	. 60
a a contr	cagta cagactatca ctgcatttgt ttaagtccac	100
aagge	sagen cagaceacca cogonocoge coungrooms	
	•	
<210>	•	
<211>	•	
<212>		
<213>	Homo sapiens 1109525	
<400>	323	
tcago	ccaga gaaagagtca gtcccataat cccagatctc caccacatcy gcacagagge	60
		100
ctgtt	aggec agggggetge cetgeteece getgeeecea	100
<210>	324	
<211>	100	
<212>	DNA	
<213>	Homo sapiens 1325561	
<400>	224	
	aaggg aacttttat ctcatatttg aaacataagt cacaagaatr actaacattt	60
accaa		
atgct	ttaaa ttattaatcc tagttttcag aattttatta	100
-010	325	
<210> <211>		
<212>		
	Homo sapiens 1343877	

<400> 325 aaaagtgggt ttttggatgc tcttaaatga gaaatcattt cttaccagtw aaagagaaaa 100 aagtaccaag tagagaataa tggggatact acctaagaca <210> 326 <211> 100 <212> DNA <213> Homo sapiens 1382794 <400> 326 aaaactttta ccaataattt tcatggagga ttttatgaaa taaccaaaak ctattaaaaa 100 tgcttatgct gaaaaatata aaatattcag ttttctatta <210> 327 <211> 100 <212> DNA <213> Homo sapiens 1813081 <400> 327 tgcaaaagag aaggccacac cttgctcaga aagacctgat gtgtggggcy agtgcaaggg 60 100 gtcaaaaact gaggagaggc cagaaggcca aagcagcttg <210> 328 <211> 100 <212> DNA <213> Homo sapiens 1858955 caggotgtgg atgoogaaga gttoattota cotgottoat taaataactr atgtgtttot 100 gggcaatcag gcccaatctc agccctgcca cctataatta <210> 329 <211> 100 <212> DNA <213> Homo sapiens 2002962 tgatgatgta tgccacgtgc ttgcccctgt ctgcagaagg acagcagcay agtcctgaat 60 100 tagecaccgg ggccttcaga gaggatcccg ggacagtgtg <210> 330 <211> 100 <212> DNA <213> Homo sapiens 2008988 ggttattctt tccaaaattc cgcaaggctc caattaattc catgtgtatk gtatatttaa 60 100 cacgcatttc ttcctgagaa caacacaaga tggtgtactt

<210>	331	
<211>	100	
<212>	DNA	
	Homo sapiens 2051772	
(21)/	Nomo Baptens 2001/72	
.400		
	331	
ttactct	tet cageeccagg gagatgeatt ttaceageag ageeaceeay acceaagace	60
agagcta	atga ccaaggcata tgagctcaaa aagccccagc	100
<210>	332	
<211>	54	
	DNA	
	·	
(213)	Homo sapiens rs5443	
400		
<400>	332	
catcato	etge ggeateaegt cygtggeett eteceteagt ggeegeetae tatt	54
	•	
<210>	333	
<211>	82	
<212>	DNA	
	Homo sapiens rs4884	
	110000 Dapaona abaota	
<400>	333	
	geet acttetggge ggggateatg tegteratgg actggeettt etceaactte	60
geaggeg	geet activings gaggateaty togethatay actigateth thecaatete	00
		~~
ttctcca	attt ccaccatgag ct	82
<210>	334	
<211>	60	
<212>	DNA	
<213>	Homo sapiens rs4646	
<400>	334	
	ctcc tatgggttgt caccaagcta ggtgctattk gccatctgct cctgttcaca	60
geatact	gree targggrege carragera ggreetarik geraterger terigireara	00
<210>	335	
<211>	59	
<212>	DNA	
<213>	Homo sapiens rs1079598	
<400>	335	
	caga atcacctatt caaaaggcga atcygatcat gtggttcctg ctgccctta	59
-geege.	cada accadente cadadagada acolamenta acadanacea acadenacea	
	•	
<210>	336	
<211>		
<212>		
<213>	Homo sapiens rs1800498	
	•	
<400>	336	
	tgtg aagaaaagag ccttgggtty gactagggaa cctggggcca ctccttcctc	60
2223		
tgctac	taaa	70

WO 20	04/016768			PCT/US2003/0	126229
	34/010/00	63/68		1 € 1/032003/0	,2022)
<210>	337		•		
<211>	95				
<212> <213>	DNA Homo sapiens rs594689				
12137	nome supreme issued				
<400>		ttaaaaaaaa	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~		60
994666	cctc cctttgtagg gtaggagaaa t	ccgggggac	gggcaaggag	yccaccigig	80
tcccc	acct cggggtctct agcccgctgt (ctctg			95
<210>	338				
<211> <212>	97 DNA				
<213>					
400					
<400>	338 taat ggttacatct ctgatgtaaa g	gttattgtat	aaaattcaaq	agaacatgcr	60
ttatgg	ttgc gcttcagggt acaaaaccac	tggaggg			97
	•				
<210>	339				
<211> <212>	63 DNA				
<213>	Homo sapiens AF100634				
<400>	339 aacc tgatggccct cattagtgct (taactattav	cttqqaaqqa	cagggggtga	60
900049	auce ogaoggoese caecagegoe (caaccacal	cccggaagca	caggegeega	00
cag					63
<210>	340				•
<211> <212>	70 DNA				
<213>	Homo sapiens rs285				
400					
<400>	340 ggtt ttgcttaatt ctcaattcaa (tatetettea	tcttttagva	actatagaat	60
		-3		55-5555-	
tttgtt	gttg				70
<210>	341				
<211> <212>	100 DNA				
<213>					
<400>	341				
	acgo ottocacago caggagotos (gcaggacgct	caaggaggtg	ctgacrtgct	60
					100
cctggt	gage geggtgeaeg egetttaagt (grgergggea			100
212	240			•	
<210> <211>	342 75				
<212>	DNA				
<213>	Homo sapiens rs1800404				
<400>	342				
-2007					

gcggtaa	attt cctgtgcttc tttccagatc gtgcacagaa ctctggcrgc catgctgggt	60
tcccttg	gcag cactg	75
<210>	343	
	62	
	DNA	
	Homo sapiens rs2065160	
<400>		
gcctctc	cgat gagtaaatat gggycaaatg aagtaaacag actatgcgta ggtatcatca	60
aa		62
<210>	344	
<211>	69	
<212>	DNA	
<213>	Homo sapiens rs1042602	
<400>		60
ttatgtg	gtca atggatgcac tgcttggggg atmtgaaatc tggagagaca ttgattttgc	60
ccatga	agc	69
	·	
<210>	345	
<211>	79	
<212>	DNA	
<213>	Homo sapiens rs3287	
<400>	345	
ccatcc	tcca acacacaca aaataagcag ctaatggcaa trctagtggt cttcccaatt	60
cacaag	acct gtgcttcaa	79
<210>	346	
<211>	52	
<212>	DNA	
<213>	Homo sapiens rs17203	
<400>		
cttcaa	attg ctttaagtac tctasgaaga ccttgactgt tggatttttg ag	52
<210>	347	
<211> <212>		
<213>		
<400>	347	50
ccattt	cttt gtaaaataac aataactttr aaggcaaaag caagattctg taaaccaaca	60
ttggaa	aaag	69
<210>		
<21.1>	66	

<212> DNA

WO 2004/016768 PCT/US2003/026229

<213>	Homo sapiens	rs2763				
<400> gtgacco	348 etgt gaggtcaggt	ccccagatt	gasgtctgag	tgtgggcaag	tgtgtcaaaa	60
ggggct						66
	240					
<210> <211>	349 62				•	
	DNA					
	Homo sapiens	rs2161			•	
<400>	349					
caggcca	aga gcgtcctatg	aatcatrcat	ttgttcctgt	tattgctgtt	cacagagtgg	60
ca						62
<210>	350	•				
<211>	60		·			
	'DNA					
<213>	Homo sapiens	rs2695				
<400>	350					
atttgtg	gttg ggtggtctag	rctattcaga	aaatctcaga	ggaggacaaa	tgatagtgca	60
<210>	351				,	
<211>	84			•		
<212>	DNA					
<213>	Homo sapiens	rs2862				
<400>	351				,	
	agtg caagataaaa	aggaaatagc	aattcaaggc	acaaagctaa	gvacat.gcaa	60
cacceg	agog caagacaaa		aassoaaggs		97	
caatata	agat gatttggggg	tggg				84
				·		
<210>	352					
<211>	.67				•	
<212>	DNA					
<213>	Homo sapiens	rs2891				
<400>	352					
	gcag gacattcca	ggctctctaa	caygagtgtc	tgcagcccca	ttcgctttga	60
gatgtg	э.					67
<210>	353					
<211>	80					
<212>	DNA					
<213>	Homo sapiens	rs2816				
<400>	353					
	ggca gggatttgtg	g acactgcatt	gctgggctgt	gttccycggg	ctcttctgga	. 60
ccttgc	accg tggatacca	3				. 80

	ne.	
<210>	354	
<211>	66	
<212>		
<213>	Homo sapiens rs3309	
<400>	354	
gcctgag	gaat cccacacatt tcagtccaag awaaccttcc tcaaattttc catctcccat	60
cagagg		66
555		
<210>	355	
<211>	60	
<212>		
	•	
<213>	Homo sapiens rs3317	
	•	
<400>	355	
	cagt gtatcacatc ttcaggatag gtrataacag tgtgaagggt gtgctcattt	60
CCCacc	cage graceacare recaggaray greataacay egradaggge graceacer	60
	·	
<210>	356	
<211>		
<212>	DNA	
<213>	Homo sapiens rs140864	
<220>		
<221>	misc_feature •	
	(29)(29)	
<223>		
<223>	n is ite or absent	
<400>	356	
		56
caaaac	ctgc tecatgteca atetgetena agtteaacea acetecetaa ecettg	50
<210>	357	
<211>		
<212>	DNA	
<213>	Homo sapiens rs16626	
	· · · · · · · · · · · · · · · · · · ·	
<220>		
<221>	_	
<222>	(28)(28)	
<223>	n is CGTGAAGTCC or absent	
<400>	357	
atatto	acet ceagecacea gteaccenta tgggttggga cetetgg	47
454550	2000 00000000 3000000000 03530003550 0000055	
	·	
<210>	358	
<211>	63	
<212>		
<213>	Homo sapiens rs16434	
<220>		
	wine feeture	
	misc_feature	
<222>	(41)(41)	
42225	n is TCCCACCCACTCTCCTCCACCTTC or absent	

<400> tctgctt	358 :ete	ttgagataca	tagttaaagt	agacacggcc	nagacttagg	aggattgcct	60
gtt							63
<210> <211> <212> <213>	359 465 DNA Homo	o sapiens :	X00238				
<220> <221> <222> <223>	(152 n is TAAC		TAGAGGGCTAGA AAATCCTGCAGA GTTAG				
<400> ccacago	359 stgt	aacattgtgt	tttccttgtc	tgtgccaggg	acaccttggc	atcagatgcc	60
tgaaggt	agc	agcttgtccc	tetttgeett	ctctaattag	atatttctct	ctctctcc	120
ctctctc	ccat	aaagaaaact	atgagagagg	gntttcctaa	ccaagtttga	gggtatgaac	180
atactct	cat	tttccttttc	tataaagctg	aggagaagag	tgagggagtg	tgggcaagag	240
aggtggd	ctca	ggctttccct	gggcctgatt	gaactttaaa	acttctctac	taattaaaca	300
acactg	ggct	ctacactttg	cttaaccctg	ggaactggtc	atcagccttt	gacctcagtt	360
cccct	cctg	accagetete	tgccccaccc	tgtcctctgg	aacctctgcg	agatttagag	420
gaaagaa	acca	gttttcaggc	ggattgcctc	agatcacact	atctc		465
<210><211><211><212><213>	360 97 DNA Homo	sapiens	X53550				٠
<222>	(58)	c_feature)(58) s (Alu) or	absent				
<400> aagtgc	360 tgta	ggccatttag	attagagaag	aaaagagcct	tctcaagaag	gccttctnaa	60
aactcc	tctg	agtctgtata	cgctgtcatc	gaagact			97
<210><211><211><212><213>		o sapiens	AL392048		·		
<400> cccggg	361 tgtg	cgtagggcgg	gegeeaagge	ggctcggcgg	ggatccgtcc	tcgccagggg	60

ccggrtcccg gcgggaggag gcgccctccc tgcccccgc cacggcggag cgtctgcaga 141 atggtgacag gattctgggt t <210> 362 <211> 127 <212> DNA <213> Homo sapiens M57427 <220> <221> misc_feature <222> (77)..(77) <223> n is (Alu) or absent . <400> 362 aactgggaaa atttgaagag aaagtacaca gatacatttg taaggttgtc tgttacttag gctacagagg aaagaanaga attccctctc tctaaacaca ctctaacaca caggagttga 120 127 gaactca <210> 363 <211> 136 <212> DNA <213> Homo sapiens AF015160 <220> <221> misc_feature <222> (106)..(106) <223> n is (Alu) or absent <400> 363 totaggecca gatttatggt aactgataat tattgataat ttetttagat ttgttgatat 60 tctacttgat gaaacatcca aattttactt tatgtaaatg tggccnaaaa gtgtcagaaa 120 136 ataaccaatt gtgctt

(19) World Intellectual Property Organization International Bureau



(43) International Publication Date 26 February 2004 (26.02.2004)

(10) International Publication Number WO 2004/016768 A3

(51) International Patent Classification7: C12Q 1/68

G01N 33/48,

(74) Agent: HAILE, Lisa, A.; Gray Cary Ware & Freidenrich LLP, 4365 Executive Drive, Suite 1100, San Diego, CA

(21) International Application Number:

PCT/US2003/026229

(22) International Filing Date: 19 August 2003 (19.08.2003)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

60/404,357 60/467,613 19 August 2002 (19.08.2002) US 2 May 2003 (02.05.2003) US

(63) Related by continuation (CON) or continuation-in-part (CIP) to earlier applications:

US Filed on US

60/404,357 (CIP) 19 August 2002 (19.08.2002) 60/467,613 (CIP)

Filed on

2 May 2003 (02.05.2003)

(71) Applicant (for all designated States except US): DNAPRINT GENOMICS, INC. [US/US]; 900 Cocoanut Avenue, Sarasota, FL 34236 (US).

(72) Inventors; and

(75) Inventors/Applicants (for US only): FRUDAKIS, Tony, N. [US/US]; 3707 Plumosa Terrace, Bradenton, FL 34210 (US). SHRIVER, Mark, D. [US/US]; 720 Hemlock St., Boalsburg, PA 16827 (US).

92121-2133 (US).

(81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

- with international search report
- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments
- (88) Date of publication of the international search report: 22 December 2005

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

2004/016768 A3

(54) Title: COMPOSITIONS AND METHODS FOR INFERRING ANCESTRY

(57) Abstract: Ancestry informative marker (AIMs) polynucleotides that contain a single nucleotide polymorphism or that contain an insertion or deletion, are provided, as are methods of using panels of the AIMs to draw an inference as to a trait of an individual. The trait can be, for example, biogeographical ancestry, a pigmentation trait, responsiveness to a drug, or susceptibility to a disease. Also provided are methods of determining the proportional ancestry of an individual. Reagents and kits also are provided.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US03/26229

A. CLASSIFICATION OF SUBJECT MATTER IPC(7) : G01N 33/48; C12Q 1/68		-
US CL : 702/19; 435/6		
According to International Patent Classification (IPC) or to both na	tional classification and IPC	•
B. FIELDS SEARCHED		
Minimum documentation searched (classification system followed b U.S.: 702/19; 435/6	oy classification symbols)	
Documentation searched other than minimum documentation to the	extent that such documents are included in	the fields searched
·		
Electronic data base consulted during the international search (name Please See Continuation Sheet	e of data base and, where practicable, search	n terms used)
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category • Citation of document, with indication, where a		Relevant to claim No.
Y,P WO 02/097047 A2 (Frudakis) 5 December 2002 (5.1	2.2002).	1-11, 24-58
Y,P US 2003/012065 (Sorenson et al) 11 Sept. 2003 (11.	9.2003).	1-11, 24-58
Y Parra, E. J., Am. J. Hum. Genet., 1998, Vol. 63, page	es 1839-1851.	1-11, 24-58
Y .McKeigue, P. M., Ann. Hum. Genet., 2000, Vol. 64,	pages 171-186.	3, 8-11, 32-34, 39, 47- 48,
Y,P Fernandez, J. R., Obesity Res., 7 July 2003, Vol. 11,	No. 7, pages 904-911.	1-11, 24-58
Y Durham-Pierre, D., Hum. Mutation, 1996, Vol. 7, pa	ges 370-373.	3, 8-11, 32-34, 39, 47- 48,
Y Collins-Scheramm, H. E., Am. J. Hum. genet., 11 Fe	b. 2002, Vol. 70, pages 737-750.	1-11, 24-58
Y Chakraborty, R., Ethnicity & Disease, 1991, Vol. 1,	pages 245-256.	1-11, 24-58
Y Chakraborty, R., PNAS, 1988, Vol. 85, pages 9119-9	9123.	1-11, 24-58
Further documents are listed in the continuation of Box C.	See patent family annex.	
Special categories of cited documents:	"T" later document published after the inter	
"A" document defining the general state of the art which is not considered to be of particular relevance	date and not in conflict with the applica principle or theory underlying the inver	ntion
"E" earlier application or patent published on or after the international filing date	"X" document of particular relevance; the considered novel or cannot be considered when the document is taken alone	
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"Y" document of particular relevance; the considered to involve an inventive step combined with one or more other such	when the document is
"O" document referring to an oral disclosure, use, exhibition or other means	being obvious to a person skilled in the	
"P" document published prior to the international filing date but later than the priority date claimed	"&" document member of the same patent f	amily
Date of the actual completion of the international search	Date of mailing of the international search	2005
30 September 2005 (30.09.2005) Name and mailing address of the ISA/US	Authorized officer	/ //
Mail Stop PCT, Atm: ISA/US		1 Chair
Commissioner of Patents P.O. Box 1450	Marjorie Moral	X X X
Alexandria, Virginia 22313-1450 Facsimile No. (703) 305-3230	Telephone No (\$71)272-1600	000

Form PCT/ISA/210 (second sheet) (July 1998)

PCT/US03/26229 INTERNATIONAL SEARCH REPORT Continuation of B. FIELDS SEARCHED Item 3: East, STN, Biosis, Compuscience, Biotedhds, Biotechno, Medline, Embase, WPI covering terms: SNP, polymorphism, ancestry, marker, primer, array, geographical, biogeographical, marker panel, African ancestry, Indian, Hispanics.

	/26229

INTERNATIONAL SEARCH REPORT

ategory *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim 1	
Y	NCBI record of a sequence ID U73638, Human Chromosome 11 Cosmid cSRL16b6 complete sequence, publicly available 25 July 1997.	4-6, 36-37	
Y	NCBI record of a sequence ID AP000685, Human Chromosome 11 clone CMB9-8M6 map 11q23, publicly available 20 May 2000.	4-6, 36-37	
٠	· ·		
	·		
		,	
		•	
	·		
		•	
	·		
,			
	•		
	·		

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US03/26229

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)		
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:		
Claim Nos.: because they relate to subject matter not required to be searched by this Authority, namely:		
Ctaim Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:		
3. Claim Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).		
Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)		
This International Searching Authority found multiple inventions in this international application, as follows: Please See Continuation Sheet		
 As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.: 1-11 and 24-58 		
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:		
Remark on Protest The additional search fees were accompanied by the applicant's protest.		
No protest accompanied the payment of additional search fees.		

INTERNATIONAL SEARCH REPORT	PCT/US03/26229
BOX II. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING	
This application contains the following inventions or groups of inventions which concept under PCT Rule 13.1. In order for all inventions to be examined, the app	
Group I, claim(s) 1-30, drawn to a method of inferring a trait of an individual.	
Group II, claim(s) 31-58, drawn to a method of estimating proportional ancestry.	
Group III, claim(s) 59-71, drawn to an article of manufacture.	
Group IV, claim(s) 72-82, drawn to a kit.	
This application contains claims directed to more than one species of the generic invention because they are not so linked as to form a single general inventive cond	
In order for more than one species to be examined, the appropriate additional examined.	mination fees must be paid. The species are as follows:
Species A: ancestry information markers of SEQ ID NOs: 1-331, recited in claims 4-6, 35-37, 72-73, and 75.	
Species B: a set of traits, as recited in claims 3 and 12-23.	
The inventions listed as Groups I-IV do not relate to a single general inventive coi 13.2, they lack the same or corresponding special technical features for the follow photograph of an individual of known proportional ancestry is known in the art. S	ring reasons: an article of manufacture comprising a
The species listed above do not relate to a single general inventive concept under species lack the same or corresponding special technical features for the following	
Species of group A, SEQ ID NOS. are unrelated, each has a different structure and	d function, and therefore are independent inventions.
Species of group B, different traits, are structurally unrelated, each has different fu different from data generated for any other trait.	unction, data generated for each trait is expected to be